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Supporting Information

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Rh^I-Catalyzed Two-Component [(5+2)+1] Cycloaddition Approach toward [5-8-5] Ring Systems

**Feng Huang, Zhong-Ke Yao, Yi Wang, Yuanyuan Wang, Jialing Zhang, and
Zhi-Xiang Yu^{*[a]}**

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Supporting Information

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1. General Methods of Synthesis

Air and moisture sensitive reactions were carried out in oven-dried glassware sealed with rubber septa under a positive pressure of dry CO mixed gas or nitrogen from a balloon, unless otherwise indicated. Similarly sensitive liquids and solutions were transferred via syringe. Reactions were stirred using Teflon-coated magnetic stir bars. Elevated temperatures were maintained using thermostat-controlled silicone oil baths. Organic solutions were concentrated using a Büchi rotary evaporator with a desktop vacuum pump. Tetrahydrofuran and diethyl ether were distilled from sodium and benzophenone prior to use. Dioxane (extra dry, water < 50 ppm), $[\text{Rh}(\text{CO})_2\text{Cl}]_2$ was commercially available and used as received. Analytical TLC was performed with 0.25 mm silica gel 60F plates with a 254 nm fluorescent indicator. The TLC plates were visualized by ultraviolet light and treatment with acidic *p*-anisaldehyde stain followed by gentle heating. Purification of products was accomplished by flash chromatography on silica gel and the purified compounds show a single spot by analytical TLC.

NMR spectra were measured on a Varian INOVA 600 (^1H at 600 MHz, ^{13}C at 150 MHz) magnetic resonance spectrometer. Data for ^1H -NMR spectra are reported as follows: chemical shift (ppm: s = singlet, d = doublet, t = triplet, q = quartet, dd = doublet of doublets, td = triplet of doublets, ddd = doublet of doublet of doublets, tdd = triplet of doublet of doublets, m = multiplet), coupling constant (Hz), and integration. Data for ^{13}C -NMR are reported in terms of chemical shift (ppm) relative to residual solvent peak. Infrared spectra were recorded on an AVATAR 330 Fourier transform spectrometer (FT-IR) and are reported in wavenumbers (cm^{-1}). Mass spectra (MS) and high-resolution mass spectra (HRMS) were recorded on a VG-ZAB-BS mass spectrometer (EI, 70 eV).

Abbreviations:

THF = tetrahydrofuran

PE = petroleum ether

EA = ethyl acetate

DCE = 1,2-dichloroethane

PCC: Pyridinium Chlorochromate

PDC: Pyridinium Dichromate

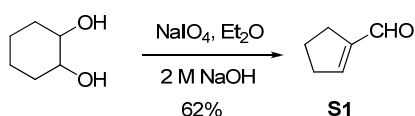
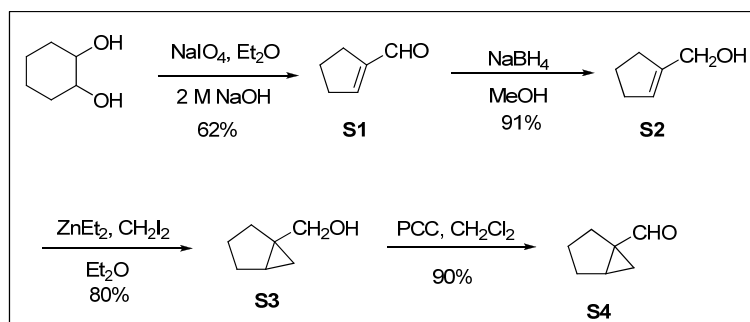
DIBAL-H: Diisobutylaluminum Hydride

DEAD = diethyl azodicarboxylate

m.p. = melting point

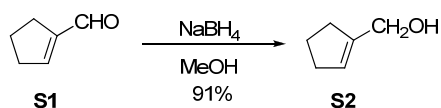
2. General Procedure for the Preparation of Ene-VCP Substrates

(1) Procedure for the Preparation of Bicyclo[3.1.0]hexane-1-carboaldehyde (S4)



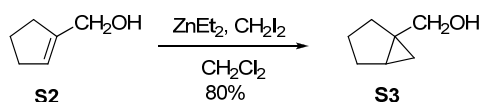
Cyclopent-1-enecarbaldehyde (S1)^[1]

To a solution of sodium periodate (20.3 g, 94.8 mmol) in H₂O (150 mL) at 0 °C was added 1,2-diol-cyclohexane (10.0 g, 86.2 mmol) dropwise over 10 min. After stirring for 10 min, a solution of Et₂O (80 mL) and 2 N NaOH (40 mL) was added consecutively. Then the reaction was stirred for 1 h at room temperature. The mixture was extracted with Et₂O (40 mL x 3). After washing with saturated water and brine, the combined organic layer was dried over MgSO₄ and concentrated. The residue was purified by flash column chromatography to afford 4.7 g (62%) **S1** as a colorless oil.



Cyclopentenylmethanol (S2)^[2]

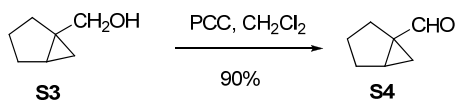
A solution of 5.17 g (53.8 mmol) **S1** dissolved in 50 mL of methanol was cooled to 0°C. Then NaBH₄ (0.6 g, 16 mmol) was added in small portion to the above prepared solution over 10 minutes. After 2 h, the reaction solution was evaporated to about one-half volume under reduced pressure. The reaction was then quenched with water and the resulting aqueous phase was extracted with ether and washed with brine. The combined ether phase was dried over anhydrous Na₂SO₄. After removal of the solvent, the residue was purified by flash column chromatography to give 4.8 g (91%) **S2** as a colorless liquid.



Bicyclo[3.1.0]hexane-1-methanol (S3)^[3]

Diethyl zinc solution (19.2 mL, 1.5 M in hexane, 28.8 mmol) and CH₂I₂ (4.6 mL, 57.4 mmol) were sequentially added to a solution of **S2** (2.8 g, 28.7 mmol) in anhydrous CH₂Cl₂ (40 mL) at 0 °C. The reaction mixture gradually became a white suspension while stirred at 0 °C. The reaction was monitored by TLC. After 12 h, the reaction was quenched with saturated NH₄Cl. The resulting mixture

was extracted with ether and the combined organic phase was dried over MgSO_4 and concentrated. The residue was purified by flash column chromatography to afford 2.5 g (80%) **S3** as a colorless oil.

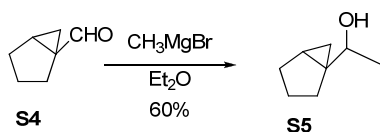
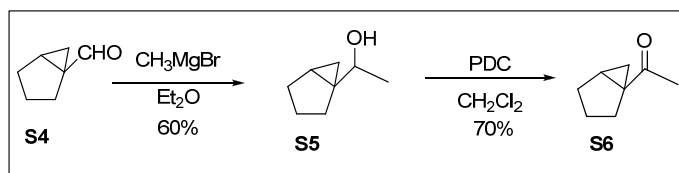


Bicyclo[3.1.0]hexane-1-carboaldehyde (**S4**)

SiO_2 (6.9 g) was added to a solution of 1.8 g (16.0 mmol) **S3** in 100 mL of CH_2Cl_2 . Two equivalents of PCC (6.9 g, 32.0 mmol) were added dropwise to the above solution over 10 minutes. The resulting mixture was stirred at room temperature for 2 h. Then the reaction mixture was filtered and the precipitate was washed with excess Et_2O . The organic layers were collected and dried over MgSO_4 . After concentrated, the residue was purified by flash column chromatography to afford the desired product **S4** 1.58 g as a colorless oil in 90% yield.

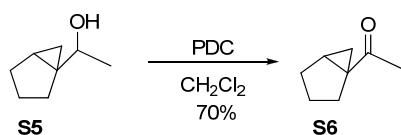
^1H -NMR (600 MHz, CDCl_3): δ 8.99 (s, 1H), 2.18-2.13 (m, 1H), 1.97-1.94 (m, 1H), 1.84-1.73 (m, 4H), 1.37-1.28 (m, 2H), 1.1 (t, $J = 6.0$ Hz, 1H). ^{13}C -NMR (150 MHz, CDCl_3): δ 200.7, 42.12, 28.57, 26.26, 24.54, 20.84, 15.39. IR (FT-IR): $\nu = 2941, 2868, 1684, 1450, 1288, 1227, 1192, 1153, 1040, 958, 930$ cm^{-1} . MS (EI): m/z (%) = 110 (M^+ , 100), 95 (53), 81 (100), 67 (65), 41 (100). HRMS calcd for $\text{C}_7\text{H}_{10}\text{O}$: 110.0732. Found: 110.0728.

(2) Procedure for the Preparation of 1-(Bicyclo[3.1.0]hexan-1-yl)ethanone (**S6**)



1-(Bicyclo[3.1.0]hexan-1-yl)ethanol (**S5**)^[4]

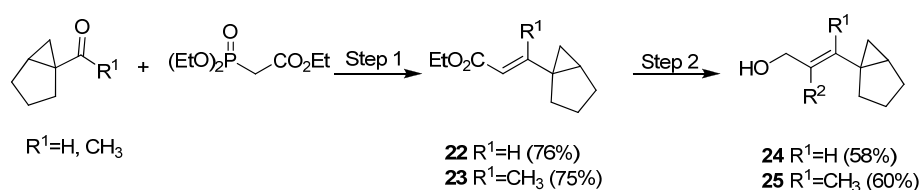
Methylmagnesium bromide (15.0 mL, 3 M, 45.0 mmol) was dissolved in 270 mL anhydrous dimethyl ether under N_2 , and then cooled to 0 °C. **S4** (45.0 mmol, in 10 mL Et_2O) was added slowly to the above solution. The solution was then stirred under 0 °C for 30 minutes before it was poured into the mixture of 100 g ice and 100 mL 1 M H_2SO_4 (aq). After extracted with Et_2O , washed with water, and brine, dried over MgSO_4 , and concentrated in vacuo, the crude mixture was purified by flash column chromatography to afford 3.4 g (60 %) **S5** as a light yellow liquid.



1-(Bicyclo[3.1.0]hexan-1-yl)ethanone (**S6**)^[5]

1-(Bicyclo[3.1.0]hexan-1-yl)ethanol **S5** (3.4 g, 27.0 mmol) was dissolved in 100 mL anhydrous CH_2Cl_2 and cooled to 0 °C. Then PDC (11.1 g, 30 mmol) was added in batches and the resulting solution was stirred for 10 h at room temperature. The product mixture was filtered through a short silica gel column. Then the filtrate was concentrated to get the product with 70% yield (2.7 g) as a light yellow liquid.

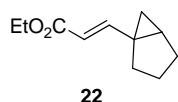
(3) General Procedure for the Preparation of Vinyl Cyclopropyl Alcohols



Step 1: To a flask containing NaH (1.2 equiv.) and THF at 0 °C was added triethyl phosphonoacetate (1.2 equiv.). After stirring at room temperature for 30 min, the corresponding cyclopropyl carbonyl compound (1.0 equiv.) was added dropwise and the reaction was allowed to stir overnight. After quenching with brine, extracting with Et_2O , and drying over MgSO_4 , concentration of the organic phase in vacuo gave a crude oil that was further purified by flash chromatography (petroleum ether/ EtOAc = 9:1). **22** or **23** was obtained as a clear, colorless oil.

Step 2: To a Schlenk flask charged with ester **22** or **23** (1.0 equiv.) in THF at -78 °C was added DIBAL-H (1 M in toluene, 2.2 equiv.) dropwise. The reaction was warmed to room temperature overnight and was quenched with ethylacetate and aqueous potassium tartrate tetrahydrate. Stirring was continued until the solution was clear. Extracted with Et_2O , washed with brine, dried over MgSO_4 , evaporation and purification by flash column chromatography (petroleum ether/ EtOAc = 4:1) provided alcohol **24** or **25** as a clear, colorless oil.

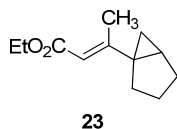
Physical Data



(*E*)-Ethyl 3-(bicyclo[3.1.0]hexan-1-yl)acrylate (**22**)

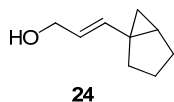
$^1\text{H-NMR}$ (600 MHz, CDCl_3): δ 6.80 (d, J = 15.6 Hz, 1H), 5.79 (d, J = 15.6 Hz, 1H), 4.18 (q, J = 7.2 Hz, 2H), 1.87-1.84 (m, 1H), 1.78-1.75 (m, 3H), 1.69-1.67 (m, 1H), 1.56-1.54 (m, 1H), 1.28 (t, J = 7.2 Hz, 3H), 0.95 (t, J = 5.4 Hz, 1H), 0.87-0.84 (m, 2H). $^{13}\text{C-NMR}$ (150 MHz, CDCl_3): δ 167.1, 155.2, 116.2, 59.9, 31.8, 29.1, 28.1, 27.0, 20.7, 16.9, 14.3. IR (FT-IR): ν = 2934, 2862, 1638, 1446, 1308, 1275, 1209, 1161, 1095, 1044, 983 cm^{-1} . MS (EI): m/z (%) = 180 (M^+ , 35), 151 (33), 79 (47), 43 (100). HRMS

calcd for C₁₁H₁₆O₂: 180.1150. Found: 180.1148.



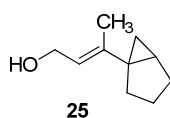
(E)-Ethyl 3-(bicyclo[3.1.0]hexan-1-yl)but-2-enoate (23)

¹H-NMR (600 MHz, CDCl₃): δ 6.74 (s, 1H), 4.17 (q, J = 7.2 Hz, 2H), 1.98-1.92 (m, 1H), 1.95 (s, 3H), 1.88 (d, J = 1.8 Hz, 1H), 1.84-1.78 (m, 1H), 1.73 (dd, J = 12.0 and 7.8 Hz, 1H), 1.68-1.63 (m, 1H), 1.40-1.38 (m, 1H), 1.28 (t, J = 5.4 Hz, 3H), 0.88-0.84 (m, 1H), 0.78 (t, J = 4.8 Hz, 1H), 0.67-0.64 (m, 1H). ¹³C-NMR (150 MHz, CDCl₃): δ 168.8, 145.4, 127.9, 60.3, 31.2, 28.8, 27.5, 27.0, 21.3, 14.7, 14.3, 13.2. IR (FT-IR): ν = 2956, 2924, 1708, 1461, 1261, 1111, 1090, 745 cm⁻¹. MS (EI): m/z (%) = 194 (M⁺, 95), 165 (77), 147 (63), 121(100), 79 (75). HRMS calcd for C₁₂H₁₈O₂: 194.1307. Found: 194.1306.



(E)-3-(Bicyclo[3.1.0]hexan-1-yl)prop-2-en-1-ol (24)

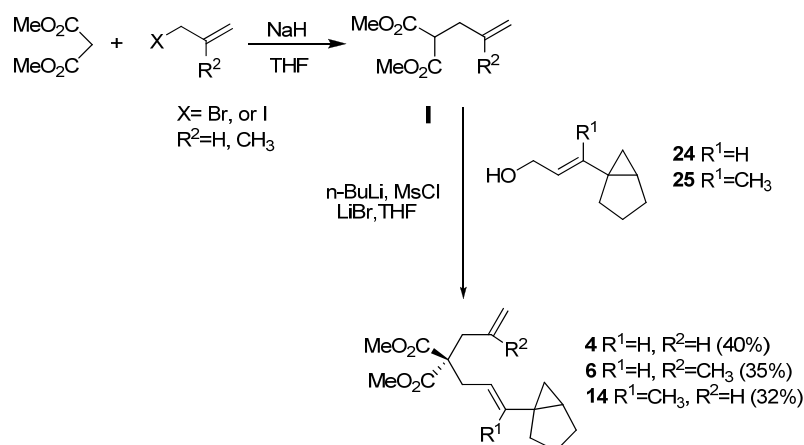
¹H-NMR (600 MHz, CDCl₃): δ 5.65-5.57 (m, 2H), 4.10 (d, J = 5.4 Hz, 2H), 1.86-1.81 (m, 1H), 1.77-1.69 (m, 3H), 1.66-1.61 (m, 1H), 1.53 (bs, 1H), 1.27-1.25 (m, 1H), 1.23-1.19 (m, 1H), 0.69 (t, J = 4.8 Hz, 1H), 0.59-0.57 (m, 1H). ¹³C-NMR (150 MHz, CDCl₃): δ 138.8, 124.6, 63.8, 30.2, 29.2, 27.2, 26.6, 20.8, 14.5. IR (FT-IR): ν = 3327, 2932, 2860, 1664, 1451, 1040, 1006, 966 cm⁻¹. MS (EI): m/z (%) = 138 (M⁺, 8.0), 120 (15), 91 (52), 79 (100), 41 (66). HRMS calcd for C₉H₁₄O: 138.1045. Found: 138.1044.



(E)-3-(Bicyclo[3.1.0]hexan-1-yl)but-2-en-1-ol (25)

¹H-NMR (600 MHz, CDCl₃): δ 5.52-5.49 (m, 1H), 4.18 (d, J = 6.6 Hz, 2H), 1.80-1.73 (m, 4H), 1.67 (bs, 1H), 1.60 (s, 3H), 1.39-1.37 (m, 1H), 1.27-1.22 (m, 2H), 0.62 (dd, J = 8.4 and 4.8 Hz, 1H), 0.54 (t, J = 4.2 Hz, 1H). ¹³C-NMR (150 MHz, CDCl₃): δ 142.2, 122.0, 59.6, 34.9, 30.8, 27.4, 23.8, 20.9, 14.3, 12.5. IR (FT-IR): ν = 3330, 2930, 2859, 1649, 1448, 1128, 997, 933 cm⁻¹. MS (EI): m/z (%) = 152 (M⁺, 11), 123 (47), 93 (75), 79 (73), 43 (66). HRMS calcd for C₁₀H₁₆O: 152.1201. Found: 152.1200.

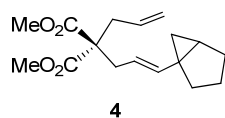
(4) General Procedure for the Preparation of Geminal Diester-Tether Substrates **4**, **6**, and **14**



Step 1: To a flask charged with NaH (1.8 equiv.) and THF at 0 °C was added dimethyl malonate (1.8 equiv.) dropwise. After stirred for 30 min, allyl iodide or allyl bromide (1.0 equiv.) was added and the reaction mixture was stirred at room temperature overnight. Quenching with brine, extracting with Et₂O, drying over MgSO₄, evaporation, and purification by flash column chromatography (petroleum ether/EtOAc = 9:1) provided **I** as a clear colorless oil.

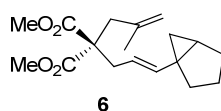
Step 2: (a) To a 250 mL flask containing alcohol **24** or **25** (1.0 equiv.) in THF at -78 °C was added *n*-BuLi (1.6 M in hexane, 1.2 equiv.), followed 10 min later by methane sulfonyl chloride (1.2 equiv.), and then immediately by lithium bromide (4.5 equiv.) in one portion. The reaction mixture was stirred for 30 min. (b) To a second flask charged with NaH (1.6 equiv.) in THF was added **I** (1.6 equiv.) dropwise over 15 min. After stirred for 30 min at room temperature, the solution was cooled to -78 °C and the solution of the first flask was transferred in via cannula. The reaction mixture was stirred for 2 h at -78 °C, then allowed to warm to room temperature. Quenching with water, extracting with Et₂O, drying over MgSO₄, evaporation and purification by flash column chromatography (petroleum ether/EtOAc = 9:1) provided geminal diester derivatives **4**, **6**, and **14** as clear colorless oils.

Physical Data



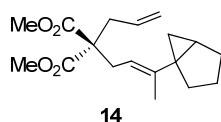
(*E*)-Dimethyl 2-allyl-2-(3-(bicyclo[3.1.0]hexan-1-yl)allyl)malonate (**4**)

¹H-NMR (600 MHz, CDCl₃): δ 5.69-5.61 (m, 1H), 5.42 (d, *J* = 15.6 Hz, 1H), 5.21-5.17 (m, 1H), 5.12-5.07 (m, 2H), 3.71(s, 6H), 2.65-2.58 (m, 4H), 1.76-1.75 (m, 2H), 1.68-1.65 (m, 2H), 1.62-1.59 (m, 2H), 1.21-1.16 (m, 1H), 0.62 (t, *J* = 4.8 Hz, 1H), 0.51-0.49 (m, 1H). ¹³C-NMR (150 MHz, CDCl₃): δ 171.5, 140.6, 132.7, 119.21, 119.17, 58.3, 52.5, 52.4, 37.2, 36.1, 30.7, 29.7, 27.4, 26.5, 21.1, 14.4. IR (FT-IR): ν = 2952, 2860, 1734, 1436, 1208, 994, 920, 858 cm⁻¹. MS (EI): *m/z* (%) = 292 (M⁺, 4.0), 219 (47), 120 (86), 91 (82), 79 (100). HRMS calcd for C₁₇H₂₄O₄: 292.1675. Found: 292.1676.



(E)-Dimethyl 2-(3-(bicyclo[3.1.0]hexan-1-yl)allyl)-2-(2-methylallyl)malonate (6)

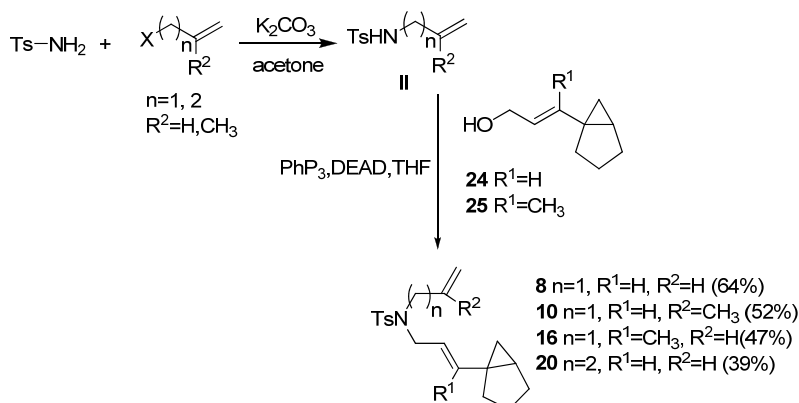
$^1\text{H-NMR}$ (600 MHz, CDCl_3): δ 5.40 (d, $J = 15.3$ Hz, 1H), 5.25-5.20 (m, 1H), 4.86-4.85 (m, 1H), 4.73 (t, $J = 0.6$ Hz, 1H), 3.69 (s, 6H), 2.69 (s, 2H), 2.61 (d, $J = 7.2$ Hz, 2H), 1.76-1.70 (m, 2H), 1.68-1.66 (m, 2H), 1.67 (s, 3H), 1.63-1.58 (m, 2H), 1.19-1.15 (m, 1H), 0.61 (t, $J = 4.8$ Hz, 1H), 0.51-0.48 (m, 1H). $^{13}\text{C-NMR}$ (150 MHz, CDCl_3): δ 171.5, 140.4, 139.9, 119.3, 115.4, 57.7, 51.9, 40.2, 35.8, 30.3, 29.4, 27.1, 26.1, 23.0, 20.7, 14.0. IR (FT-IR): $\nu = 2950, 2859, 1735, 1436, 1272, 1199, 1176, 968\text{ cm}^{-1}$. MS (EI): m/z (%) = 306 (M^+ , 5.0), 187 (36), 165 (34), 151 (100), 120 (68), 79 (78). HRMS calcd for $\text{C}_{18}\text{H}_{26}\text{O}_4$: 306.1831. Found: 306.1845.



(E)-Dimethyl 2-allyl-2-(3-(bicyclo[3.1.0]hexan-1-yl)but-2-enyl)malonate (14)

$^1\text{H-NMR}$ (600 MHz, CDCl_3): δ 5.69-5.62 (m, 1H), 5.08-5.05 (m, 3H), 3.70 (s, 6H), 2.63-2.61 (m, 4H), 1.81-1.64 (m, 5H), 1.56 (s, 3H), 1.29-1.23 (m, 2H), 0.51-0.47 (m, 2H). $^{13}\text{C-NMR}$ (150 MHz, CDCl_3): δ 171.5, 141.6, 132.6, 118.9, 116.3, 66.2, 58.0, 52.2, 37.0, 35.4, 31.2, 31.1, 27.6, 22.3, 12.5. IR (FT-IR): $\nu = 2951, 2859, 1735, 1436, 1269, 1139, 920, 862\text{ cm}^{-1}$. MS (EI): m/z (%) = 306 (M^+ , 6.0), 233 (20), 147 (30), 134 (100), 119 (52), 79 (46). HRMS calcd for $\text{C}_{18}\text{H}_{26}\text{O}_4$: 306.1831. Found: 306.1830.

(5) General Procedure for the Preparation of Tosylamide-Tether Substrates 8, 10, 16, and 20

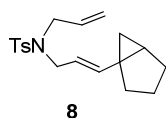


Step 1: To a flask was added TsNH_2 (1.1 equiv.), K_2CO_3 (1.5 equiv.) and acetone at $0\text{ }^\circ\text{C}$. After stirring for 30 min, allyl bromide or 4-bromobut-1-ene (1.0 equiv.) was added and the reaction mixture was refluxed for 7 h. Quenching with brine, extraction with Et_2O , drying over MgSO_4 , evaporation, and purification by flash column chromatography (petroleum ether/ $\text{EtOAc} = 2:1$) provided **II** as a white solid.

Step 2: To a solution of alcohol **24** or **25** (1.0 equiv.), **II** (1.2 equiv.), and PPh_3 (2.2 equiv.) in THF was

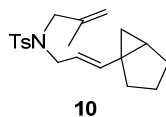
added DEAD (2.2 equiv.) at room temperature and the reaction mixture was stirred for 20 h at room temperature. The resulting mixture was diluted with Et₂O, washed with water and brine, and dried over Na₂SO₄. After removal of the solvent, the residue was purified by flash column chromatography with Et₃N-impregnated silica gel (petroleum ether/EtOAc = 9:1) to give **8**, **10**, **16**, and **20** as colorless oils.

Physical Data



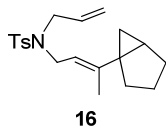
(*E*)-N-Allyl-N-(3-(bicyclo[3.1.0]hexan-1-yl)allyl)-4-methylbenzenesulfonamide (**8**)

¹H-NMR (600 MHz, CDCl₃): δ 7.69 (d, *J* = 8.4 Hz, 2H), 7.28 (d, *J* = 7.8 Hz, 2H), 5.66-5.59 (m, 1H), 5.39 (d, *J* = 10.2 Hz, 1H), 5.16-5.11 (m, 3H), 3.80-3.75 (m, 4H), 2.42 (s, 3H), 1.75-1.58 (m, 6H), 0.88-0.84 (m, 1H), 0.64 (t, *J* = 4.8 Hz, 1H), 0.48-0.47 (m, 1H). ¹³C-NMR (150 MHz, CDCl₃): δ 143.0, 141.0, 133.0, 129.5, 127.3, 119.4, 118.6, 49.0, 48.8, 30.2, 29.1, 27.2, 26.5, 21.5, 20.8, 14.4. IR (FT-IR): ν = 2917, 2860, 1340, 1157, 1090, 1032, 952, 814, 753 cm⁻¹. MS (EI): *m/z* (%) = 331 (M⁺, 2.0), 250 (18), 224 (38), 155 (48), 91 (100), 41 (84). HRMS calcd for C₁₉H₂₅NO₂S: 331.1606. Found: 331.1614.



(*E*)-N-(3-(Bicyclo[3.1.0]hexan-1-yl)allyl)-N-(2-methylallyl)-4-methylbenzenesulfonamide (**10**)

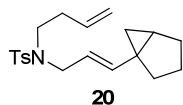
¹H-NMR (600 MHz, CDCl₃): δ 7.69 (d, *J* = 7.8 Hz, 2H), 7.28 (d, *J* = 7.8 Hz, 2H), 5.57-5.50 (m, 1H), 5.34 (d, *J* = 15.6 Hz, 1H), 5.06-4.84 (m, 2H), 3.74-3.67 (m, 4H), 2.42 (s, 3H), 1.70 (s, 3H), 1.69-1.53 (m, 6H), 0.88-0.83 (m, 1H), 0.60 (t, *J* = 4.8 Hz, 1H), 0.45-0.40 (m, 1H). ¹³C-NMR (150 MHz, CDCl₃): δ 142.9, 141.2, 140.4, 133.4, 129.2, 127.3, 118.9, 114.2, 52.5, 48.8, 30.2, 29.0, 27.2, 26.4, 21.5, 21.4, 20.8, 14.3. IR (FT-IR): ν = 2927, 2859, 1337, 1157, 1093, 1002, 912, 814, 763 cm⁻¹. MS (EI): *m/z* (%) = 345 (M⁺, 4.0), 264 (16), 238 (38), 190 (46), 91 (100), 55 (100). HRMS calcd for C₂₀H₂₇NO₂S: 345.1763. Found: 345.1767.



(*E*)-N-Allyl-N-(3-(bicyclo[3.1.0]hexan-1-yl)but-2-enyl)-4-methylbenzenesulfonamide (**16**)

¹H-NMR (600 MHz, CDCl₃): δ 7.69 (d, *J* = 8.4 Hz, 2H), 7.29 (d, *J* = 7.8 Hz, 2H), 5.69-5.63 (m, 1H), 5.15-5.11 (m, 2H), 5.01-4.98 (m, 1H), 3.83 (d, *J* = 6.6 Hz, 2H), 3.77 (d, *J* = 6.0 Hz, 2H), 2.42 (s, 3H), 1.70-1.68 (m, 2H), 1.64-1.60 (m, 2H), 1.59-1.58 (m, 2H), 1.57 (s, 3H), 0.89-0.83 (m, 2H), 0.47-0.45 (m, 1H). ¹³C-NMR (150 MHz, CDCl₃): δ 142.9, 142.5, 133.3, 129.53, 129.51, 127.2, 118.3, 117.1, 49.4, 44.6, 35.0, 30.8, 27.4, 23.6, 21.5, 20.9, 14.3, 12.3. IR (FT-IR): ν = 2926, 2855, 1339, 1156, 1089, 1037, 922, 814, 756 cm⁻¹. MS (EI): *m/z* (%) = 345 (M⁺, 2.0), 238 (32), 224 (36), 155 (64), 91 (100), 41 (78).

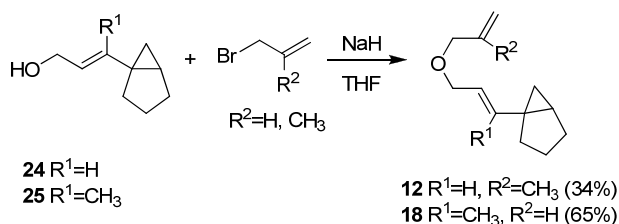
HRMS calcd for C₂₀H₂₇O₂S: 345.1763. Found: 345.1771.



(E)-N-(3-(Bicyclo[3.1.0]hexan-1-yl)allyl)-N-(but-3-enyl)-4-methylbenzenesulfonamide (20)

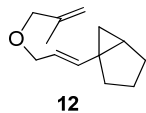
¹H-NMR (600 MHz, CDCl₃): δ 7.69 (d, *J* = 8.4 Hz, 2H), 7.28 (d, *J* = 7.8 Hz, 2H), 5.74-5.68 (m, 1H), 5.44 (d, *J* = 18 Hz, 1H), 5.20-5.15 (m, 1H), 5.05-5.01 (m, 2H), 3.78-3.74 (m, 2H), 3.18-3.15 (m, 2H), 2.42 (s, 3H), 2.27 (q, *J* = 6.6 Hz, 2H), 1.73-1.58 (m, 6H), 1.19-1.18 (m, 1H), 0.64 (t, *J* = 5.4 Hz, 1H), 0.49 (m, 1H). ¹³C-NMR (150 MHz, CDCl₃): δ 143.2, 140.9, 137.7, 135.2, 129.7, 127.4, 120.2, 117.1, 50.3, 46.6, 33.3, 30.5, 29.4, 27.4, 26.7, 21.7, 21.0, 14.6. IR (FT-IR): ν = 2930, 2860, 1338, 1156, 1090, 1031, 926, 813, 746 cm⁻¹. MS (EI): *m/z* (%) = 345 (M⁺, 6.0), 304 (12), 190 (14), 155 (20), 121 (100), 91 (68), 79 (58). HRMS calcd for C₂₀H₂₇O₂S: 345.1762. Found: 345.1757.

(6) General Procedure for the Preparation of Ether-Tethered Substrates 12 and 18



To a solution of NaH (1.2 equiv.) in THF was added a solution of alcohol **24** or **25** (1.0 equiv.) in THF at 0 °C and the mixture was refluxed for 2 h. The solution was cooled to room temperature, then allyl bromide (1.2 equiv.) was added and the mixture was refluxed overnight. After being cooled to room temperature, the reaction was quenched with water and extracted with Et₂O. Washed with brine, dried over MgSO₄, evaporation to remove the solvent, and purification by flash column chromatography (petroleum ether/EtOAc = 9:1) gave **12** and **18** as colorless oils.

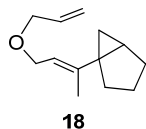
Physical Data



(E)-1-(3-(2-Methylallyloxy)prop-1-enyl)bicyclo[3.1.0]hexane (12)

¹H-NMR (600 MHz, CDCl₃): δ 5.61-5.53 (m, 2H), 4.96-4.95 (m, 1H), 4.89-4.88 (m, 1H), 3.91 (d, *J* = 5.4 Hz, 2H), 3.88 (s, 2H), 1.87-1.82 (m, 1H), 1.74 (s, 3H), 1.72-1.68 (m, 2H), 1.64-1.60 (m, 2H), 1.27-1.20 (m, 2H), 0.68 (t, 1H), 0.59-0.57 (m, 1H). ¹³C-NMR (150 MHz, CDCl₃): δ 142.6, 140.1, 122.2, 112.2, 74.1, 71.1, 30.5, 29.4, 27.5, 26.7, 21.1, 19.8, 14.8. IR (FT-IR): ν = 2929, 2859, 1450, 1113, 1077, 971, 891 cm⁻¹. MS (EI): *m/z* (%) = 192 (M⁺, 12), 121 (32), 119 (42), 93 (74), 86 (100), 67

(70), 47 (100). HRMS calcd for C₁₃H₂₀O: 192.1514. Found: 192.1520.



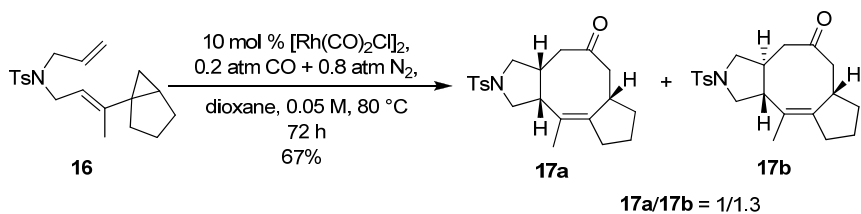
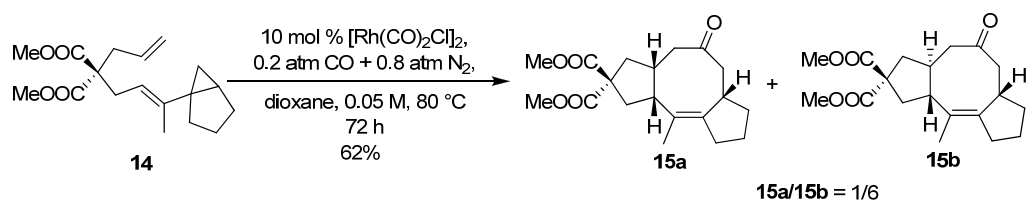
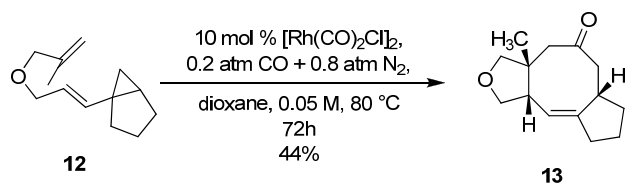
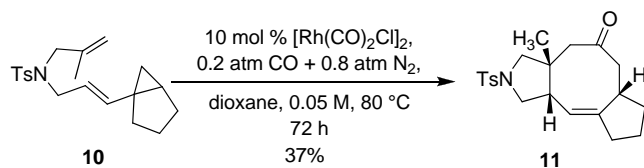
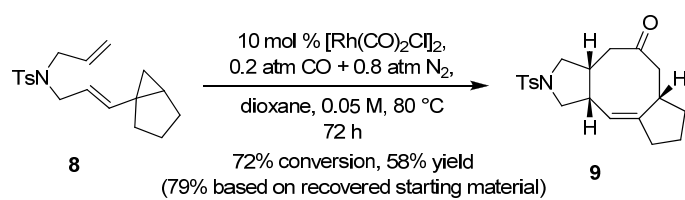
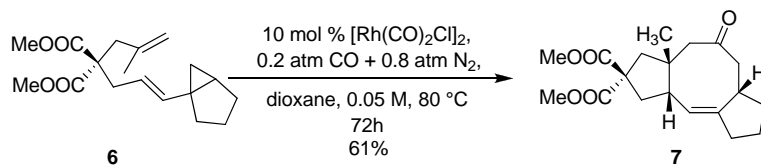
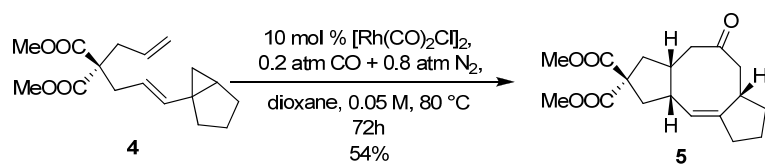
(*E*)-1-(4-(Allyloxy)but-2-en-2-yl)bicyclo[3.1.0]hexane (18)

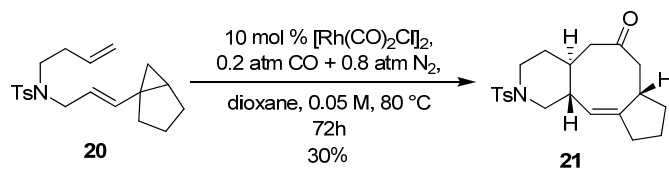
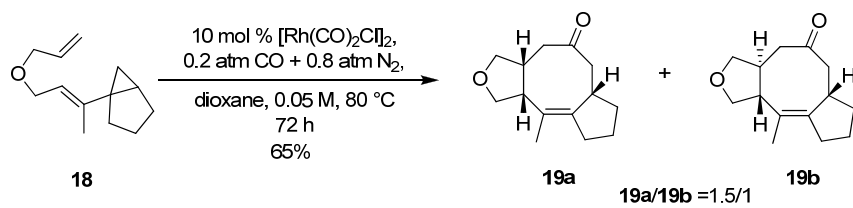
¹H-NMR (600 MHz, CDCl₃): δ 5.94 (m, 1H), 5.45 (m, 1H), 5.27 (m, 1H), 5.17 (dd, J = 10.2 and 1.2 Hz, 1H), 4.01 (d, J = 7.2Hz, 2H), 3.96 (d, J = 2.4Hz, 2H), 1.81-1.78 (m, 2H), 1.77-1.69 (m, 2H), 1.62-1.59 (m, 2H), 1.59 (s, 3H), 1.25-1.20 (m, 1H), 0.62-0.60 (m, 1H), 0.26 (t, J = 4.8Hz, 1H). ¹³C-NMR (150 MHz, CDCl₃): δ 142.5, 135.1, 119.5, 116.9, 71.1, 66.9, 35.0, 30.8, 27.5, 23.7, 21.0, 14.5, 12.5. IR (FT-IR): ν = 2926, 2860, 1450, 1066, 920, 800 cm⁻¹. MS (EI): m/z (%) = 192 (M⁺, 6.0), 134 (22), 121 (30), 91 (38), 81 (48), 91 (68), 41 (100). HRMS calcd for C₁₃H₂₀O: 192.1514. Found: 192.1515.

3. General Procedure for the [(5 + 2) + 1] Cycloaddition Reactions

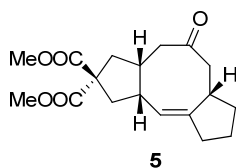
[Rh(CO)₂Cl]₂ (10 mol % to the substrate) was charged in a base-washed, oven-dried Schlenk flask under an atmosphere of nitrogen, and then a solution of the ene-VCP substrate in degassed dioxane (0.05 M) was added. The solution was bubbled with the mixed gas of CO and N₂ (0.2 atm CO + 0.8 atm N₂) for 5 min. The reaction mixture was stirred at 80 °C under balloon pressured mixed gas of CO and N₂ (0.2 atm CO + 0.8 atm N₂) until TLC indicated the completion of the reaction (72 h). After cooled to room temperature, the mixture was concentrated and the residue was purified by flash column chromatography with silica gel to afford the cycloaddition product.

4. Summary of all [(5 + 2) + 1] Cycloaddition Reactions



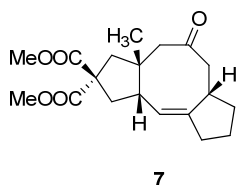


5. Physical Data for Cycloadducts



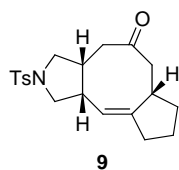
(3R*, 7R*, 11S*)-5,5-Bis(methoxycarbonyl)tricyclo[9.3.0.0^{3,7}]tetradec-1-ene-9-one (**5**)

Light yellow oil, 54% yield. ¹H-NMR (600 MHz, CDCl₃): δ 5.28-5.24 (m, 1H), 3.74 (s, 3H), 3.73 (s, 3H), 2.69-2.58 (m, 2H), 2.52-2.43 (m, 4H), 2.42-2.32 (m, 4H), 2.28-2.19 (m, 2H), 2.05-1.95 (m, 1H), 1.78-1.69 (m, 2H), 1.64-1.56 (m, 1H), 1.49-1.45 (m, 1H). ¹³C-NMR (150 MHz, CDCl₃): δ 212.2, 173.1, 172.9, 147.5, 121.8, 59.0, 53.3, 52.82, 52.81, 43.2, 41.4, 41.0, 40.0, 39.5, 37.8, 34.5, 33.2, 24.1. IR (FT-IR): ν = 2916, 2848, 1731, 1434, 1245, 1166, 848 cm⁻¹. MS (EI): m/z (%) = 320 (M⁺, 38), 302 (55), 203 (90), 91 (100), 79 (94), 41 (58). HRMS calcd for C₁₈H₂₄O₅: 320.1624. Found: 320.1621.



(3R*, 7R*, 11S*)-5,5-Bis(methoxycarbonyl)-7-methyltricyclo[9.3.0.0^{3,7}]tetradec-1-ene-9-one (**7**)

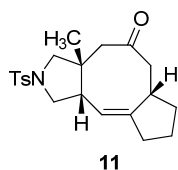
Yellow oil, 61% yield. ¹H-NMR (600 MHz, CDCl₃): δ 5.39 (d, *J* = 1.8 Hz, 1H), 3.73 (s, 3H), 3.71 (s, 3H), 2.61-2.50 (m, 4H), 2.44-2.38 (m, 2H), 2.29-2.18 (m, 4H), 2.03 (dd, *J* = 13.8 and 9.0 Hz, 1H), 1.89-1.84 (m, 1H), 1.69-1.65 (m, 1H), 1.61-1.56 (m, 2H), 1.53-1.50 (m, 1H), 1.20 (s, 3H). ¹³C-NMR (150 MHz, CDCl₃): δ 211.6, 173.9, 173.5, 147.4, 128.1, 57.9, 53.56, 53.53, 52.0, 51.2, 47.5, 47.1, 44.7, 41.2, 36.4, 35.3, 34.8, 26.4, 23.4. IR (FT-IR): ν = 2954, 2924, 1732, 1434, 1264, 1198, 1065, 847 cm⁻¹. MS (EI): m/z (%) = 334 (M⁺, 84), 316 (100), 217 (40), 122 (46), 91 (52), 41 (44). HRMS calcd for C₁₉H₂₆O₅: 334.1780. Found: 334.1772.



(3S*, 7S*, 11S*)-N-Tosyl-5-azatricyclo[9.3.0.0^{3,7}]tetradec-1-ene-9-one (**9**)

Light yellow oil, 72% conversion, 58% yield (79% based on recovered starting material). ¹H-NMR (600 MHz, CDCl₃): δ 7.72 (d, *J* = 7.8 Hz, 2H), 7.34 (d, *J* = 8.4 Hz, 2H), 4.96 (dd, *J* = 8.4 and 1.8 Hz, 1H), 3.41 (d, *J* = 10.2 Hz, 1H), 3.39 (d, *J* = 9.6 Hz, 1H), 3.22 (dd, *J* = 10.2 and 4.8 Hz, 1H), 2.82 (t, *J* = 10.2 Hz, 1H), 2.61-2.47 (m, 3H), 2.45 (s, 3H), 2.42-2.33 (m, 3H), 2.32-2.16 (m, 3H), 1.99-1.96 (m, 1H), 1.73-1.70 (m, 1H), 1.62-1.58 (m, 1H), 1.48-1.47 (m, 1H). ¹³C-NMR (150 MHz, CDCl₃): δ 211.9, 149.5, 144.3, 130.4, 128.1, 119.7, 54.5, 53.9, 51.8, 41.5, 39.6, 38.7, 35.2, 34.0, 30.4, 30.0, 24.8, 22.2. IR (FT-IR): ν = 2955, 2921, 1700, 1466, 1343, 1162, 1090, 1045, 803 cm⁻¹. MS (EI): m/z (%) = 359 (M⁺, 24), 301 (90), 204 (78), 146 (72), 91 (100), 42 (86). HRMS calcd for C₂₀H₂₅NO₃S: 359.1555. Found:

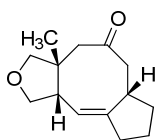
359.1550.



11

(3S*, 7S*, 11S*)-N-Tosyl-7-methyl-5-azatricyclo[9.3.0.0^{3,7}]tetradec-1-ene-9-one (11)

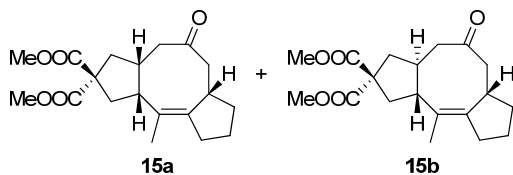
Colorless crystals, 37% yield, m.p.150 °C. ¹H-NMR (600 MHz, CDCl₃): δ 7.73 (d, *J* = 7.8 Hz, 2H), 7.34 (d, *J* = 7.8 Hz, 2H), 4.83 (dd, *J* = 8.4 and 1.2 Hz, 1H), 3.34 (dd, *J* = 9.8 and 4.8 Hz, 1H), 3.31 (d, *J* = 10.2 Hz, 1H), 3.10 (d, *J* = 9.6 Hz, 1H), 2.91 (d, 9.0Hz, 1H), 2.63 (d, *J* = 11.4 Hz, 1H), 2.57 (m, 1H), 2.45 (s, 3H), 2.40-2.30 (m, 3H), 2.28-2.19 (m, 3H), 1.97-1.90 (m, 1H), 1.72-1.67 (m, 1H), 1.60-1.57 (m, 1H), 1.44-1.42 (m, 1H), 0.87 (s, 3H). ¹³C-NMR (150 MHz, CDCl₃): δ 210.6, 149.0, 144.2, 134.9, 130.3, 127.9, 121.7, 59.3, 53.69, 53.66, 48.1, 47.5, 43.8, 38.0, 35.0, 33.4, 24.3, 23.9, 22.2. IR (FT-IR): ν = 2957, 2869, 1697, 1384, 1344, 1156, 1091, 1051, 738 cm⁻¹. MS (EI): *m/z* (%) = 373 (M⁺, 30), 315 (60), 218 (78), 160 (94), 91 (76), 42 (100). HRMS calcd for C₂₁H₂₇NO₃S: 373.1711. Found: 373.1720. This structure was confirmed by X-ray crystallographic analysis.



13

(3S*, 7S*, 11S*)-7-Methyl-5-oxatricyclo[9.3.0.0^{3,7}]tetradec-1-ene-9-one (13)

Light yellow oil, 44% yield. ¹H-NMR (600 MHz, CDCl₃): δ 5.38 (dd, *J* = 7.8 and 1.8 Hz, 1H), 4.13 (q, *J* = 13.8 Hz, 1H), 3.81 (d, *J* = 8.4, 1H), 3.52 (d, *J* = 7.2 Hz, 1H), 3.38 (d, *J* = 7.8Hz, 1H), 2.71-2.68 (m, 2H), 2.53-2.37 (m, 5H), 2.03 (d, *J* = 11.4 Hz, 1H), 1.98-1.93 (m, 1H), 1.78-1.72 (m, 1H), 1.65-1.61 (m, 1H), 1.49-1.46 (m, 1H), 1.21 (s, 3H). ¹³C-NMR (150 MHz, CDCl₃): δ 210.9, 148.7, 123.5, 79.3, 75.0, 53.4, 48.7, 47.1, 44.7, 38.2, 35.0, 33.2, 24.4, 24.3. IR (FT-IR): ν = 2954, 2873, 1695, 14548, 1055, 882, 754 cm⁻¹. MS (EI): *m/z* (%) = 220 (M⁺, 6.0), 162 (100), 135 (18), 91 (30), 79 (32), 41 (28). HRMS calcd for C₁₄H₂₀O₂: 220.1463. Found: 220.1473.



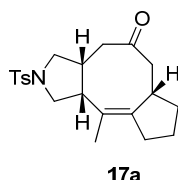
(1 : 6)

(3R*, 7S*, 11S*)-2-Methyl-5,5-bis(methoxycarbonyl)tricyclo[9.3.0.0^{3,7}]tetradec-1-ene-9-one (15b)

Yellow oil, 62% yield. ¹H-NMR (600 MHz, CDCl₃): δ 3.75 (s, 3H), 3.74 (s, 3H), 3.33-3.29 (m, 1H), 2.71-2.66 (m, 1H), 2.59-2.51 (m, 3H), 2.50-2.46 (m, 1H), 2.45-2.38 (m, 2H), 2.37-2.22 (m, 4H), 1.97-1.88 (m, 1H), 1.83-1.74 (m, 1H), 1.82-1.74 (m, 3H), 1.59 (s, 3H). ¹³C-NMR (150 MHz, CDCl₃): δ 211.2, 173.0, 172.9, 142.3, 125.7, 57.7, 52.9, 52.7, 52.6, 48.9, 45.2, 40.0, 39.2, 37.6, 37.4, 32.7, 29.6, 22.9, 15.4. IR (FT-IR): ν = 2954, 2923, 1731, 1435, 1257, 1199, 1066, 864 cm⁻¹. MS (EI): *m/z* (%) =

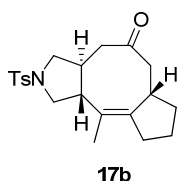
334 (M^+ , 94), 316 (82), 256 (54), 145 (82), 91 (80), 43 (100). HRMS calcd for $C_{19}H_{26}O_5$: 334.1780. Found: 334.1775.

Compound **15a** is the minor product generated in the [(5+2)+1] cycloaddition, which is inseparable from the major product **15b**. Therefore, no characterization data for **15a** was provided.



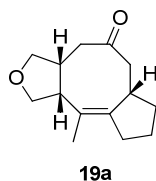
(3S*, 7S*, 11S*)-2-Methyl-N-tosyl-5-azatricyclo[9.3.0.0^{3,7}]tetradec-1-ene-9-one (17a)

Colorless crystals, 29% yield, m.p. 154-155 °C. 1H -NMR (600 MHz, $CDCl_3$): δ 7.71(d, J = 7.8 Hz, 2H), 7.33 (d, J = 7.8 Hz, 2H), 3.63 (d, J = 9.6 Hz, 1H), 3.48 (dd, J = 9.0 and 6.0 Hz, 1H), 3.18 (dd, J = 6.0 and 5.4 Hz, 1H), 2.86 (t, J = 7.2 Hz, 1H), 2.82 (d, J = 10.8 Hz, 1H), 2.56 (d, J = 6.6 Hz, 1H), 2.52-2.46 (m, 1H), 2.30-2.34 (m, 1H), 2.27-2.22 (m, 2H), 2.09 (dd, J = 4.2 and 3.6 Hz, 1H), 2.01-1.99 (m, 1H), 1.81-1.78 (m, 2H), 1.73-1.71 (m, 1H), 1.66 (s, 1H), 1.60-1.56 (m, 2H). ^{13}C -NMR (150 MHz, $CDCl_3$): δ 211.8, 143.5, 142.7, 134.0, 129.8, 127.2, 123.5, 52.6, 51.4, 50.8, 43.9, 40.6, 40.0, 39.2, 34.8, 30.4, 23.0, 21.5, 16.2. IR (FT-IR): ν = 2955, 2922, 1697, 1378, 1342, 1160, 1094, 1055, 816 cm^{-1} . MS (EI): m/z (%) = 373 (M^+ , 42), 315 (10), 218 (100), 160 (48), 91 (78), 42 (50). HRMS calcd for $C_{21}H_{27}NO_3S$: 373.1711. Found: 373.1714. This structure was confirmed by X-ray crystallographic analysis.



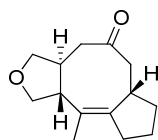
(3S*, 7R*, 11S*)-2-Methyl-N-tosyl-5-azatricyclo[9.3.0.0^{3,7}]tetradec-1-ene-9-one (17b)

White solid, 38% yield, m.p. 142-144 °C. 1H -NMR (600 MHz, $CDCl_3$): δ 7.71(d, J = 8.4 Hz, 2H), 7.33 (d, J = 8.4 Hz, 2H), 3.53 (d, J = 8.4 Hz, 1H), 3.45 (dd, J = 8.4 and 7.8 Hz, 1H), 3.27 (t, J = 10.5 Hz, 1H), 3.17 (t, J = 6.0 Hz, 1H), 2.99 (t, J = 9.9 Hz, 1H), 2.72 (dd, J = 8.4 Hz, 1H), 2.54-2.51 (m, 1H), 2.48 (d, J = 10.8 Hz, 1H), 2.45 (s, 3H), 2.36-2.25 (m, 3H), 2.20-2.04 (m, 1H), 1.87 (m, 1H), 1.77-1.59 (m, 3H), 1.54-1.52 (m, 1H), 1.45 (s, 3H). ^{13}C -NMR (150 MHz, $CDCl_3$): δ 209.8, 143.9, 143.5, 133.7, 129.7, 127.5, 123.1, 53.0, 52.2, 50.4, 48.1, 43.0, 39.0, 37.5, 32.6, 29.3, 22.3, 21.5, 15.7. IR (FT-IR): ν = 2955, 2922, 1697, 1378, 1342, 1160, 1094, 1055, 816 cm^{-1} . MS (EI): m/z (%) = 373 (M^+ , 42), 315 (10), 218 (100), 160 (48), 91 (78), 42 (50). HRMS calcd for $C_{21}H_{27}NO_3S$: 373.1711. Found: 373.1714.



(3S*, 7S*, 11S*)-2-Methyl-5-oxatricyclo[9.3.0.0^{3,7}]tetradec-1-ene-9-one (19a)

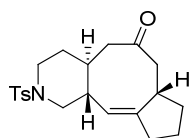
Light yellow oil, 39% yield. $^1\text{H-NMR}$ (600 MHz, CDCl_3): δ 4.11 (d, $J = 8.4$ Hz, 1H), 4.01 (t, $J = 7.8$ Hz, 1H), 3.84 (dd, $J = 5.4$ and 3.6 Hz, 1H), 3.21 (dd, $J = 8.4$ Hz and 7.8, 1H), 2.94 (t, $J = 12$ Hz, 1H), 2.70-2.68 (m, 1H), 2.57-2.46 (m, 2H), 2.38-2.36 (m, 2H), 2.38-2.36 (m, 2H), 2.26-2.20 (m, 2H), 2.19-1.74 (m, 3H), 1.71 (s, 3H), 1.67-1.57 (m, 2H). $^{13}\text{C-NMR}$ (150 MHz, CDCl_3): δ 212.1, 142.0, 124.0, 71.3, 71.0, 52.5, 45.2, 41.2, 40.0, 39.3, 34.9, 30.3, 23.0, 16.5. IR (FT-IR): $\nu = 2946, 2860, 1689, 1438, 1047, 897, 754\text{ cm}^{-1}$. MS (EI): m/z (%) = 220 (M^+ , 50), 134 (48), 105 (60), 91 (87), 79 (68), 41 (100). HRMS calcd for $\text{C}_{14}\text{H}_{20}\text{O}_2$: 220.1463. Found: 220.1468.



19b

(3S*, 7R*, 11S*)-2-Methyl-5-oxatricyclo[9.3.0.0^{3,7}]tetradec-1-ene-9-one (19b)

Light yellow oil, 26% yield. $^1\text{H-NMR}$ (600 MHz, CDCl_3): δ 4.05 (t, $J = 12$ Hz, 1H), 3.95 (t, $J = 12$ Hz, 1H), 3.84 (dd, $J = 12$ and 12.6 Hz, 1H), 3.43 (dd, $J = 13.2$ Hz and 13.8 Hz, 1H), 2.98 (d, $J = 13.2$ Hz, 1H), 2.69-2.60 (m, 2H), 2.42-2.30 (m, 2H), 2.28-2.16 (m, 1H), 2.15-2.00 (m, 1H), 1.62-1.56 (m, 2H), 1.53 (s, 3H). $^{13}\text{C-NMR}$ (150 MHz, CDCl_3): δ 210.4, 143.4, 124.1, 72.8, 70.7, 53.1, 49.9, 43.3, 41.0, 37.3, 32.5, 29.1, 22.3, 15.9. IR (FT-IR): $\nu = 2946, 2860, 1689, 1438, 1047, 897, 754\text{ cm}^{-1}$. MS (EI): m/z (%) = 220 (M^+ , 50), 134 (48), 105 (60), 91 (87), 79 (68), 41 (100). HRMS calcd for $\text{C}_{14}\text{H}_{20}\text{O}_2$: 220.1463. Found: 220.1468.



21

(3S*, 8R*, 12S*)-N-Tosyl-5-azatricyclo[10.3.0.0^{3,8}]pentadec-1-ene-10-one (21)

Colorless crystals, 30% yield, m.p. 186 °C. $^1\text{H-NMR}$ (600 MHz, CDCl_3): δ 7.63 (d, $J = 8.4$ Hz, 2H), 7.32 (d, $J = 7.8$ Hz, 2H), 4.95 (dd, $J = 8.4$ and 1.8 Hz, 1H), 3.80 (m, 1H), 3.73 (m, 1H), 3.01 (m, 1H), 2.55 (dd, $J = 12$ and 4.8 Hz, 1H), 2.49 (dd, $J = 12$ and 5.4 Hz, 1H), 2.44 (s, 3H), 2.40-2.36 (m, 1H), 2.32-2.23 (m, 3H), 2.20-2.15 (m, 2H), 2.01-1.60 (m, 5H), 1.63-1.49 (m, 1H), 1.31-1.26 (m, 1H), .089-0.83 (m, 1H). $^{13}\text{C-NMR}$ (150 MHz, CDCl_3): δ 212.1, 149.5, 144.1, 133.9, 130.4, 128.2, 120.9, 53.5, 51.8, 47.0, 46.8, 42.3, 38.1, 37.9, 33.8, 32.9, 31.6, 24.2, 22.2. IR (FT-IR): $\nu = 2954, 2923, 1697, 1340, 1215, 1163, 1090, 1022, 750\text{ cm}^{-1}$. MS (EI): m/z (%) = 373 (M^+ , 30), 316 (8.0), 218 (100), 198 (28), 91 (56), 42 (34). HRMS calcd for $\text{C}_{21}\text{H}_{27}\text{NO}_3\text{S}$: 373.1712. Found: 373.1716. This structure was confirmed by X-ray crystallographic analysis.

6. X-Ray Structure for Cycloadducts **11**, **17a**, **21**, and NOESY Correlations.

CCDC 673996, 756195, 673997 contain the supplementary crystallographic data for **11**, **17a**, and **21**.

These data can be obtained free of charge from The Cambridge Crystallographic Data Centre via www.ccdc.cam.ac.uk/data_request/cif.

Figure S1. Chemical Structure and ORTEP Figure of Cycloadduct **11**

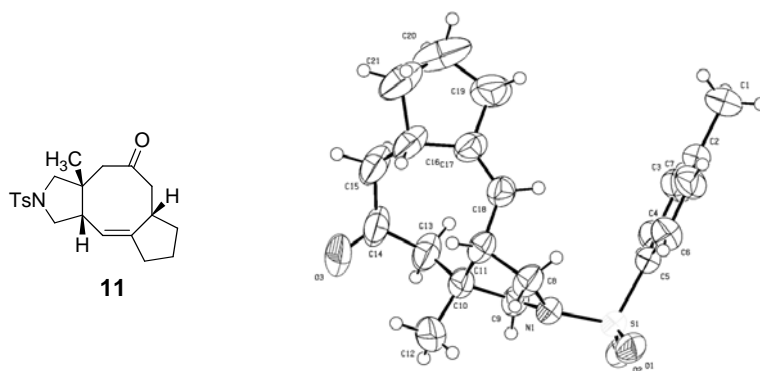


Figure S2. Chemical Structure and ORTEP Figure of Cycloadduct **17a**

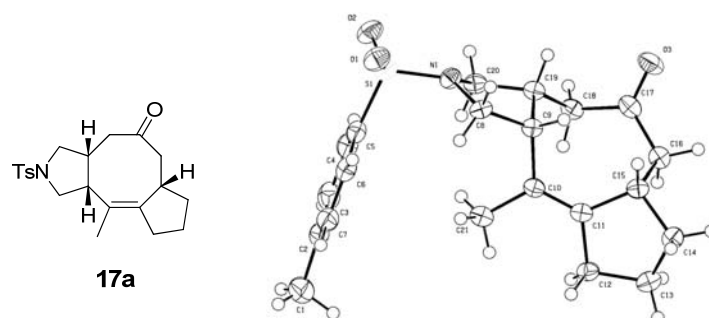


Figure S3. Chemical Structure and ORTEP Figure of Cycloadduct **21**

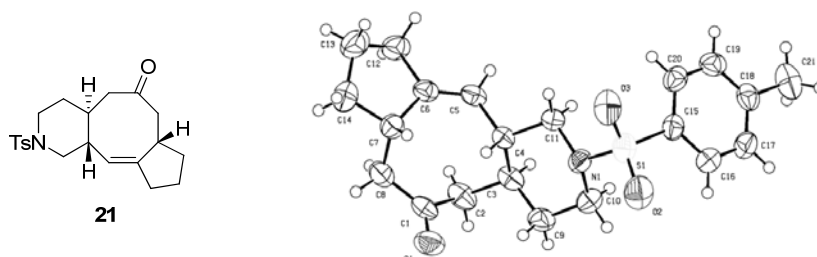


Figure S4. Selected NOESY Correlations for **5**.

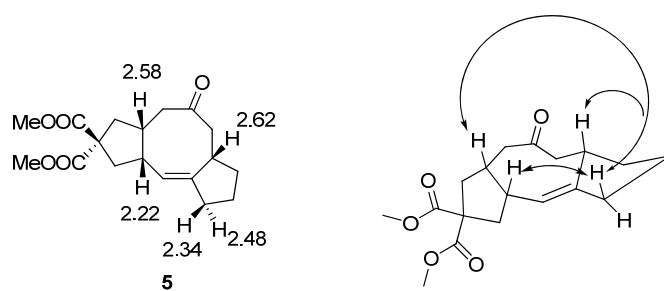


Figure S5. Selected NOESY Correlations for **7**.

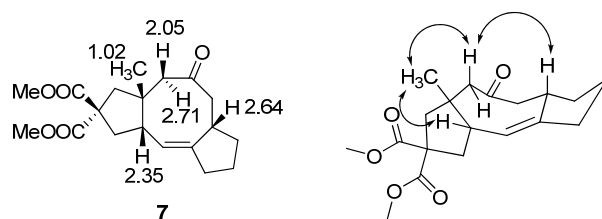


Figure S6. Selected NOESY Correlations for **9**.

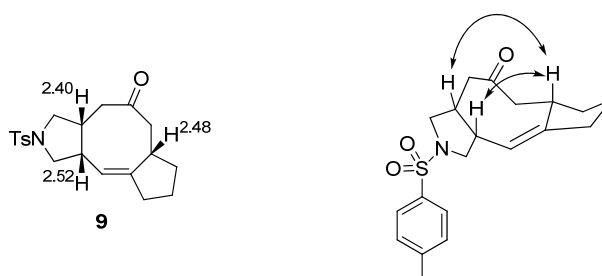
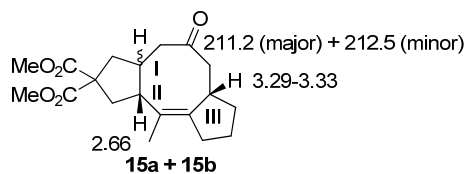
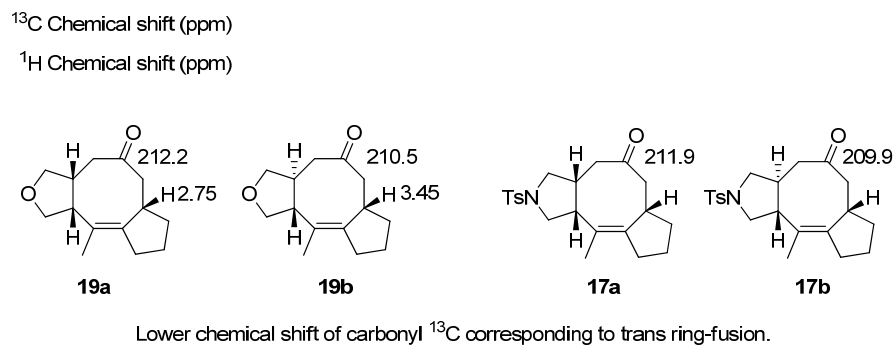


Figure S7. Selected NOESY Correlations for **13**.

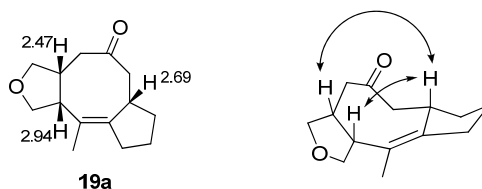


Figure S8. Stereochemistry Assignment for **15a/b**.



Since the carbonyl carbons in trans compounds have lower ¹³C NMR shift compared to those in the *cis* compounds (**19b** vs. **19a**, **17b** vs. **17a**), we assigned the major product in **15a/b** is **15b**, which has a *trans* configuration. The ratio of **15a/15b** is determined by ¹H NMR.

Figure S9. Selected NOESY Correlations for **19a**.



7. Stereochemistry Determination

Compounds **5**, **7**, **9**, **13**, and **19a** have a *cis*-fused A/B rings system and this assignment is supported by NOESY experiments. The stereochemistry of compound **7** was deduced by analogy to **11**. Compounds **15b** and **19b** have a *trans*-fused A/B rings system. This is deduced by analogy to the previous experimental findings (Y. Wang, J. Wang, J. C. Su, F. Huang, L. Jiao, Y. Liang, D. Z. Yang, S. W. Zhang, P. A. Wender, Z.-X. Yu, *J. Am. Chem. Soc.* **2007**, *129*, 10060). The assignment of the structure of **15b** was mainly based on ^{13}C NMR (see section 6 above). The configuration between the substituents at positions II and III is *cis*, as shown by XRD analysis of cycloadducts **11**, **17a**, and **21**. This can be well understood based on the computational model, thus, all the [(5+2)+1] cycloadducts were assigned to the same *cis* configuration for the substituents at positions II and III.

8. DFT Calculations and Cartesian Coordinates of Computed Species

Computational Details. All of the calculations were performed with the Gaussian 03 program.^[6] B3LYP^[7] functional was used to locate all the transition structures and intermediates. 6-31G* basis set is applied for all elements except for Rh, which uses the basis set of LANL2DZ+ECP.^[8, 9] This method has been successfully applied to predict structures and reaction mechanisms for reactions of dirhodium tetracarboxylate complexes and other Rh-catalyzed cycloadditions.^[10] Frequency calculations at the same level have been performed to confirm each stationary point to be either a minimum or a transition structure. The reported energies are Zero-point energy-corrected electronic energies ($\Delta E_{0\text{ K}}$), enthalpies ($\Delta H_{298\text{ K}}$), and Gibbs free energies ($\Delta G_{298\text{ K}}$).

Table S1. The Computed Energies and Other Thermal Parameters for the Three-Component [(5+2)+1] Reaction (in Hartree)

Structure	E_{ele}	$E_{0\text{ K}}$	$H_{298\text{ K}}$	$G_{298\text{ K}}$
S1	-1034.237214	-1034.4569113	-1034.222700	-1034.279901
TS	-1034.223202	-1034.4409475	-1034.208968	-1034.265139
S1a	-1034.232408	-1034.450763	-1034.217636	-1034.275472

S1:

C	-3.949464	0.107980	-0.338061
C	-3.623324	-0.716143	0.929695
C	-2.298877	-0.126013	1.463275
C	-1.581837	0.450140	0.237762
C	-2.590310	0.529405	-0.885573
C	-1.533615	-0.536514	-1.010051
C	-0.432100	1.364175	0.457246
C	0.345945	1.894117	-0.584199
Rh	0.896213	-0.197757	-0.114266
O	3.568510	0.428140	1.033618
C	2.537656	0.182668	0.578320
Cl	1.397667	-2.491898	-0.349142
H	-0.009496	1.794917	-1.610730
C	1.330364	3.018330	-0.376903
H	-0.298529	1.738617	1.470398
H	-1.692857	-0.860984	2.002663
H	-2.499208	0.705518	2.152354
H	-4.425390	-0.671937	1.672204

H	-3.490297	-1.772268	0.672367
H	-4.520305	1.008981	-0.080060
H	-4.541535	-0.458522	-1.065921
H	-2.541984	1.353903	-1.592664
H	-1.775503	-1.579205	-0.833947
H	-0.814242	-0.388467	-1.817963
H	2.204223	2.907585	-1.027105
H	0.861533	3.981499	-0.621974
H	1.678963	3.067739	0.659771

TS:

C	-1.152627	-0.702760	-1.207171
C	-1.463327	0.584989	0.324166
Rh	0.735349	-0.171359	-0.139513
Cl	1.271546	-2.506598	-0.254187
C	-2.267339	0.271750	-0.903203
C	-3.620746	-0.302724	-0.420411
C	-3.643308	-0.050131	1.106972
C	-2.155436	-0.046453	1.521803
C	-0.395917	1.537415	0.434792
C	0.367025	1.924348	-0.692407
C	1.459072	2.958146	-0.606116
O	3.428990	0.249494	1.274248
C	2.436561	0.114712	0.712135
H	2.274149	2.727570	-1.300231
H	1.069912	3.949383	-0.878970
H	1.878449	3.024813	0.403076
H	-0.077853	1.822468	-1.681599
H	-0.149342	1.915100	1.424296
H	-1.792321	-1.067433	1.681507
H	-1.948123	0.512609	2.440749
H	-4.084023	0.932301	1.316547
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H	-4.466923	0.171800	-0.926409
H	-3.672980	-1.374987	-0.640044

H	-2.353034	1.086943	-1.624149
H	-1.325631	-1.735420	-0.919194
H	-0.645069	-0.578858	-2.167279

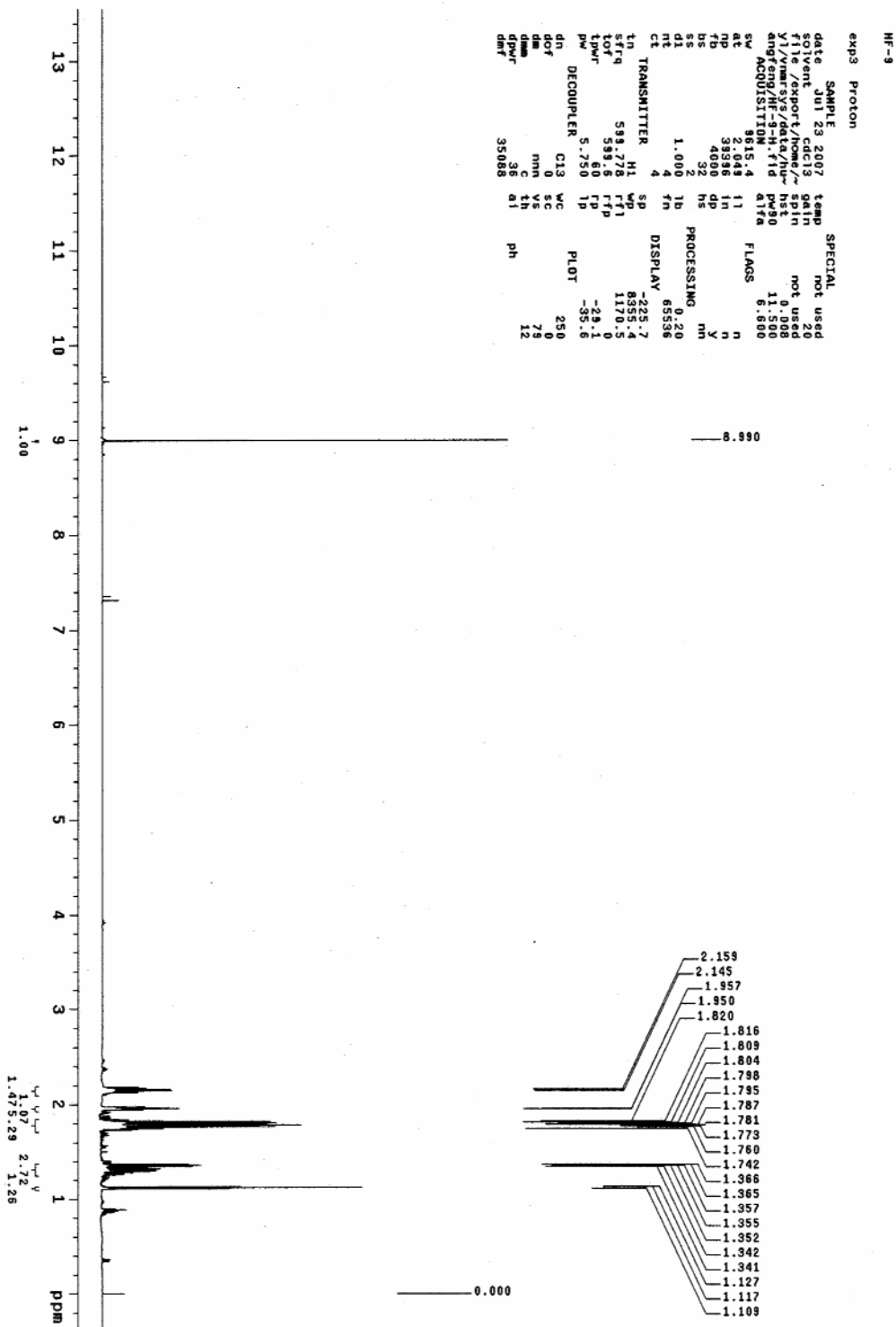
S1a:

C	-2.049183	-0.311600	-0.879838
C	-0.820993	-1.160525	-1.217556
C	-0.538734	1.681786	-0.179643
C	0.331143	1.496991	-1.301968
Rh	0.586421	-0.201884	-0.006904
C	1.512204	2.397445	-1.540314
Cl	1.932887	-2.141509	-0.125635
O	2.701675	0.893194	2.105782
C	1.919391	0.591602	1.327164
H	1.904859	2.816391	-0.607946
H	1.225356	3.232016	-2.196975
H	2.318933	1.851951	-2.040629
H	-0.089979	1.066885	-2.208207
H	-0.887421	-2.191281	-0.864999
H	-0.519231	-1.150294	-2.268039
H	-0.300900	2.446424	0.558895
C	-1.513922	0.726153	0.124982
C	-2.366305	0.724702	1.377582
C	-3.673094	0.042224	0.917593
C	-3.188647	-1.009688	-0.097534
H	-2.467938	0.206451	-1.755184
H	-1.883538	0.123601	2.159678
H	-2.505353	1.731947	1.786940
H	-4.321896	0.777367	0.423955
H	-4.238673	-0.389671	1.749078
H	-3.983576	-1.372965	-0.755944
H	-2.783202	-1.878484	0.436809

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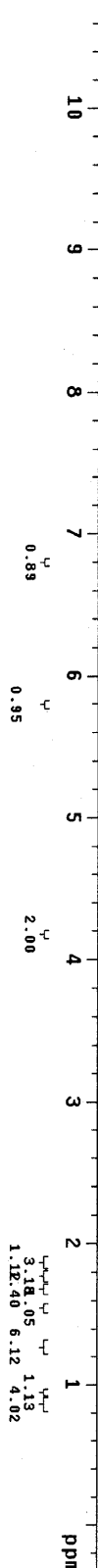
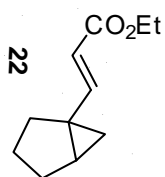
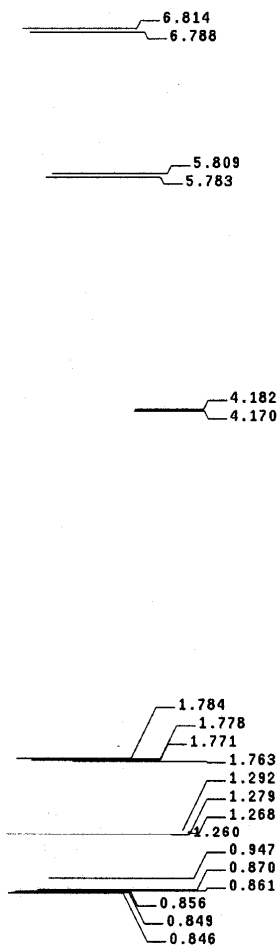
10. ¹H and ¹³C Spectra for all New Compounds



HF-1

exp3 Proton

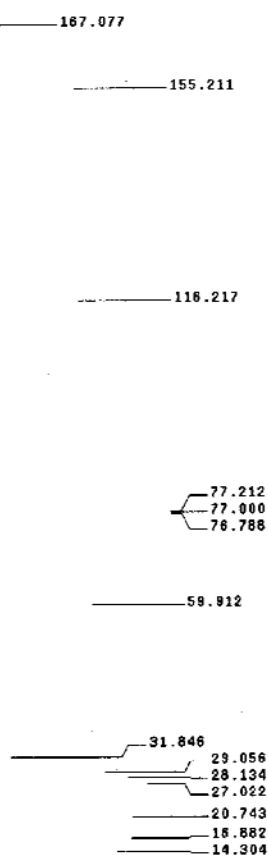
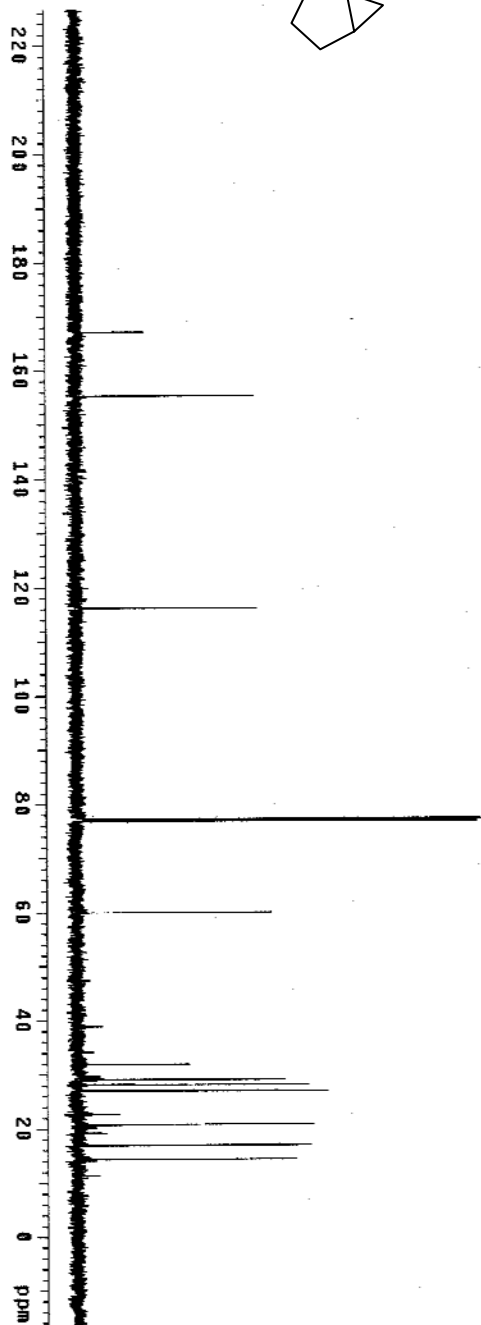
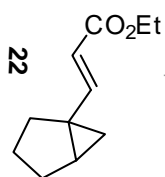
SAMPLE		SPECIAL	
date	Jul 23 2007	temp	not used
solvent	cdcl3	gain	20
file	/export/home/~	spin	not used
yl/vmr/sy/da/ta/hu	hst	0.006	
ang/enh/ff-1	ps90	11.500	
acq/si	alpha	6.600	
sv	1615.4	FLAGS	
at	2.049	n	
np	3936	in	
fb	4000	dp	
bs	32	hs	
ss	2	PROCESSING	
d1	1.000	lb	0.20
nt	4	fn	65536
ct	4	DISPLAY	
TRANSMITTER			
tn	H1	sp	-238.3
sfreq	599.778	wp	6713.6
iof	599.6	rfl	1194.3
tpwr	60	rtp	-84.2
pw	5.750	tp	-36.9
DECOUPLER			
dn	C13	mc	250
dof	0	sc	0
dm	mm	ys	167
clm	36	tn	15
dpwr	36	ph	
dmf	35088		



HF-1

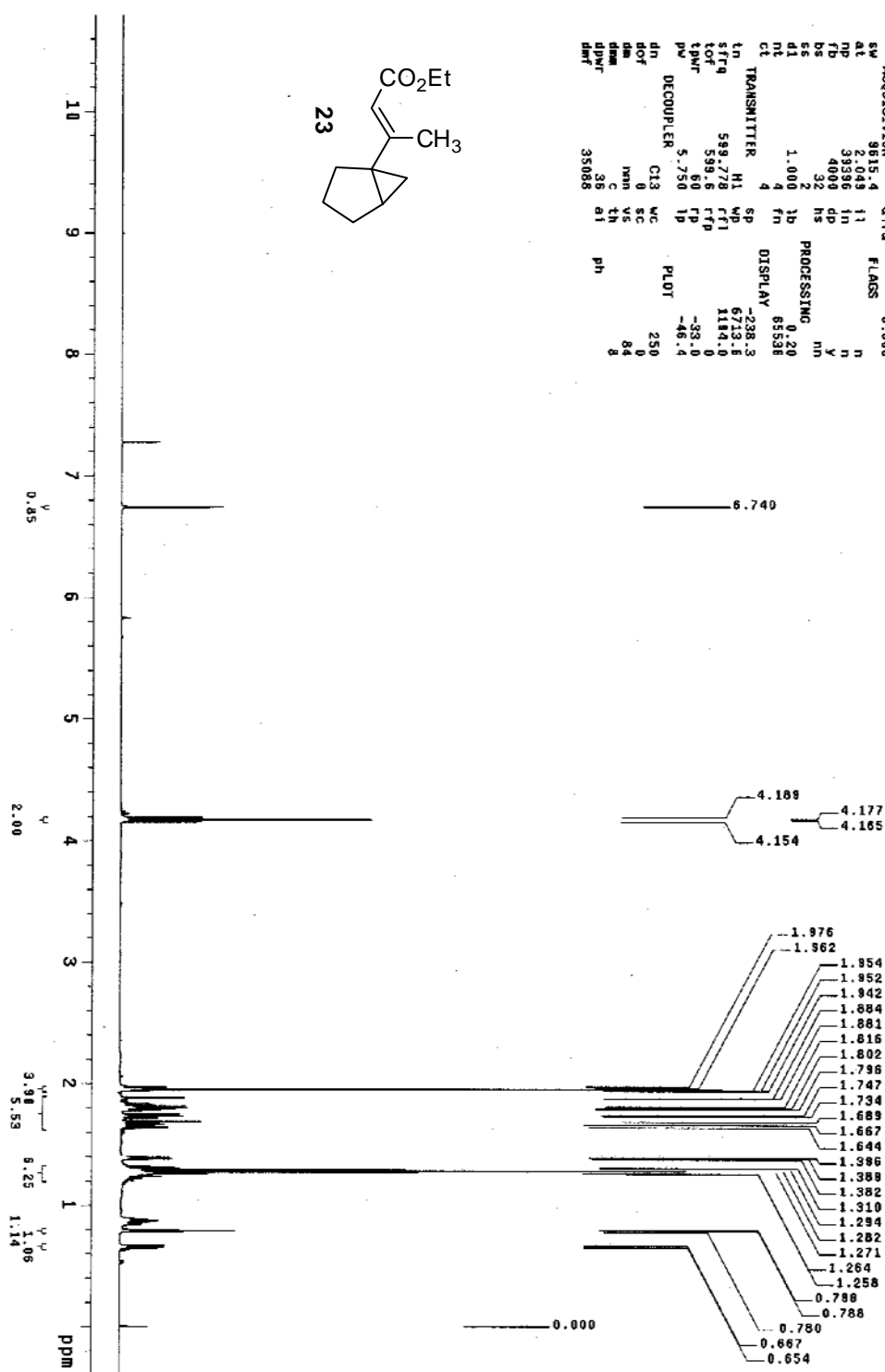
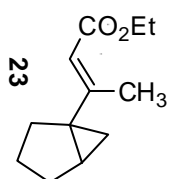
exp4 Carbon

SAMPLE		SPECIAL	
date	Jun 23 2007	temp	not used
time	15:00	spin	not used
file	/export/home/~	spin	not used
Y1/NameSYS/data/hu	not	spin	not used
angfreq/HF-1-C-fid	pw30	pw30	7.600
ACQUISITION	alpha	alpha	10.000
sw	36764.7	flags	
at	1.300	11	n
np	85624	1n	n
fb	17000	dp	y
bs	4	ns	PROCESSING
dl	1.000	1b	not used
nt	2.000	fn	not used
ct	48	fn	not used
tn	TRANSMITTER	C13	60
sfreq	150.829	wp	-2581.3
tof	2339.1	fti	36764.1
tpwr	55	ftf	14183.5
pw	3.800	fp	11612.5
DECOUPLER	1p	fp	73.0
dn	0	WC	-345.0
dot	0	WC	220
dm	yyy	sc	0
dms	yy	sc	3805
dpr	42	tn	3
dnt	12580	at	cdc
		ph	



exp3 Proton

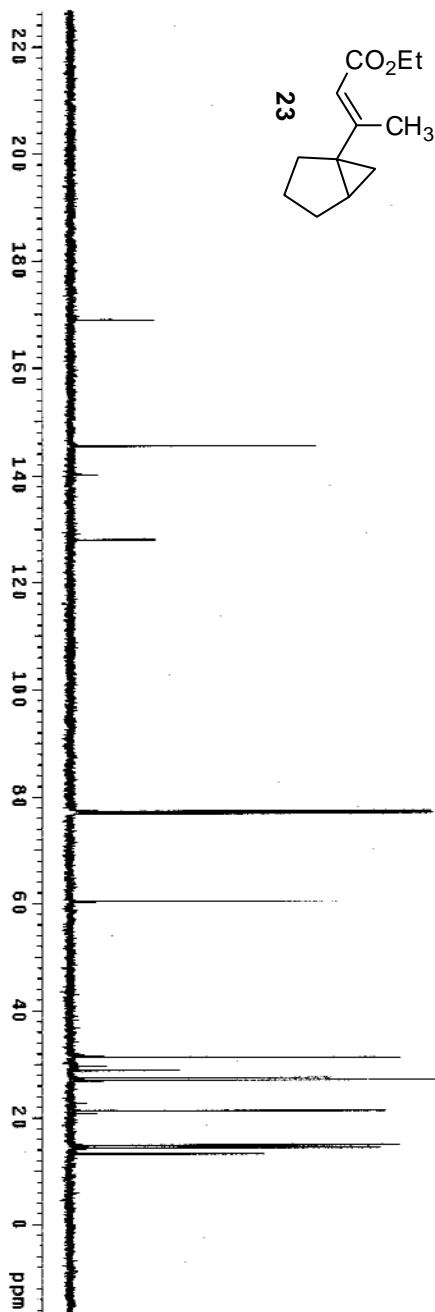
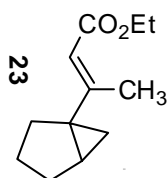
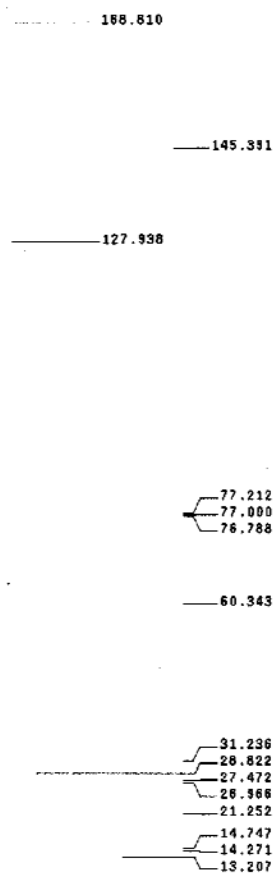
SAMPLE							SPECIAL					
date	JUN 23 2007	temp					not used					
solvent	cic13	gain					20					
file /export/home/~w	spn						not used					
y1/vr/yys/date/hu~	pst						0.988					
ang/rf-34t_rfd	pvd0						11.500					
ACQUISITION	a1ta						6.600					
av	5815.4						FCLASS					
ss	11											
fb	38586											
tp	4000											
cs	2						PROCESSING					
bs	32						0.20					
d1	1.000						65389					
nt	4											
ct	4						DISPLAY					
TRANSMITTER	4						-238.3					
tn	H1	sp					673.3					
sfreq	589.778	rft1					114.4					
tof	559.6	rtp					0					
tprv	60	fcp					-33.0					
	5.750	tp					-46.4					
DECOUPLER	C13	wc					PLOT					
dn	0	sc					250					
hof	0	vn										
dm	nrm	yn					84					
amr	32	at					ph					
idnr	35008											



HF-3

exp4 Carbon

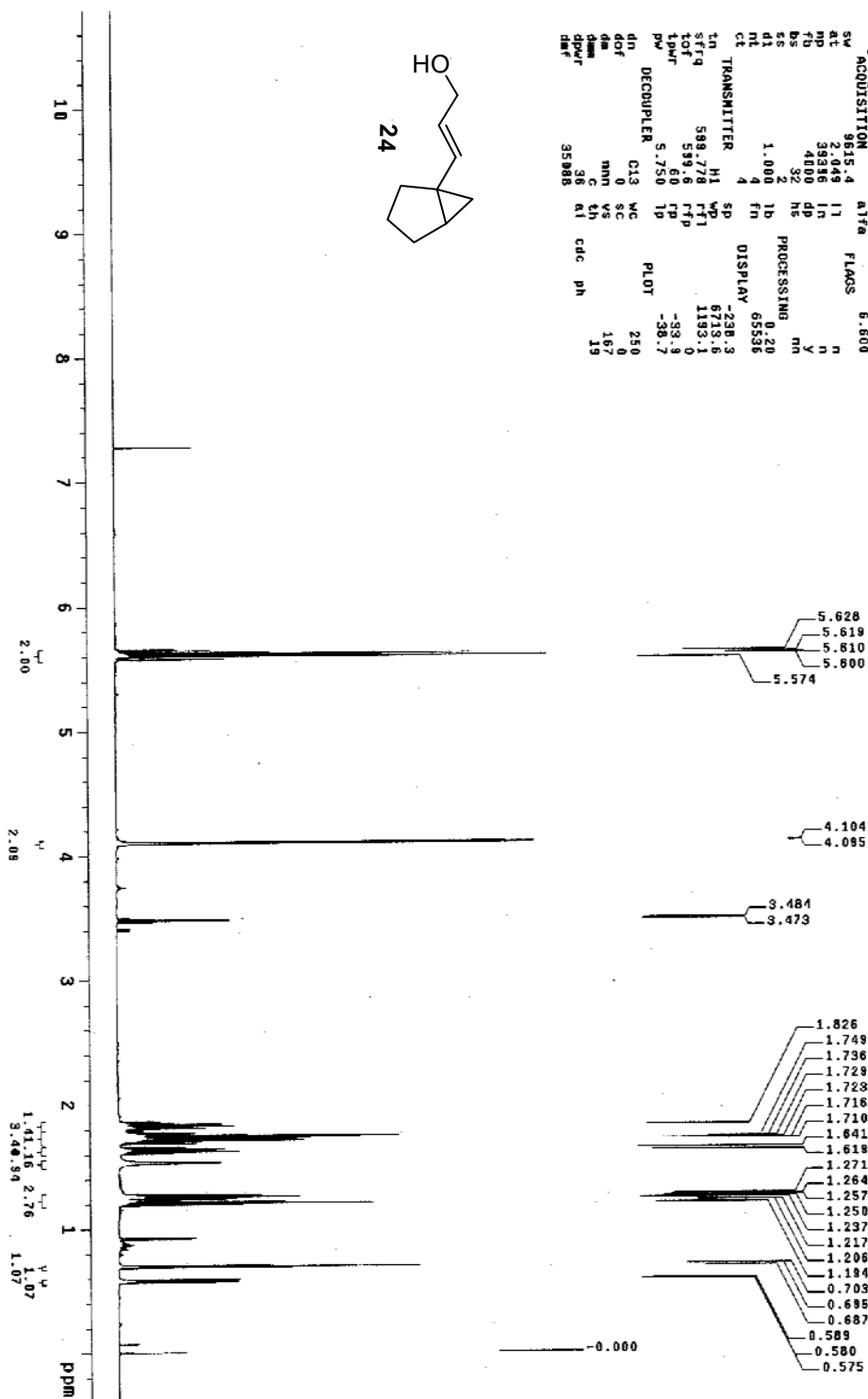
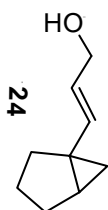
SAMPLE SPECIAL
 date Jul 23 2007 temp not used
 solvent cdc13 gain 40
 file /export/home/~ not used
 y1/vmr/sz/da/ta/hu~ not used
 angs/cm/230-718 p150 0.008
 sw ACQUISIT 10.000
 at 38764.7 11
 np 35824 in n
 fb 17000 dp y
 bs 4 ns
 ct 1.000 1b PROCESSING 2.00
 tn 48 fn not used
 TRANSMITTER C13 SP DISPLAY
 tn 2n -2580.8
 strq 150.828 wd 35764.1
 tot 2339.1 r1 14194.0
 lpr 35 r1p 11812.6
 pw 3.800 1p -269.1
 DECOUPLER H1 PLOT
 dn 0 wc 220
 dof 0 yy sc 0
 dm yy sc 0
 diam w vs 4885
 dpr 42 tn 13
 dmf 12580 at cdc ph



HF-2

exp3 proton

SAMPLE SPECIAL
 date Jul 28 2007 temp not used
 solvent cdcl3 gain 20
 file /export/home/~ not used
 y1/vmreys/data/hu~ hst 0.008
 angfreq/HF-2-H-1d pw80 11.500
 ACQUISITION
 sw 9615.4 11 flags
 at 2.000 1.0
 f2 3000 1.0
 f3 4000 1.0
 bs 32 he
 es 2
 nt 1.000 1b
 ct 4 4 fn
 TRANSMITTER M1 SP DISPLAY 238.3
 srq 589.728 613.6
 scf 589.728 1133.0
 tsmr 589.728 1133.0
 pw 5.750 1p
 DECOUPLER C13 WC PLOT 250
 dn 0 SC 0
 dof 0 SC 0
 dm mn VS 167
 dm mn VS 19
 spwr 38 kl cdc ph
 bat 35885



HF-2

expt4 Carbon

SAMPLE SPECIAL
 date Jul 23 2007 temp not used
 solvent cdcl3 gain 40
 f1h / export/home/~ spin not used
 y1h / export/home/~ p10 7.600
 shoftm/HF-2C.f1d p10 7.600
 ACQUISITION 36764.7 atfa 10.000
 sw 36764.7
 at 1.300 s1 n
 np 95824 in n
 fd 17000 dp y
 dt 1.000 hs
 nt 64 lb
 ct 44 fn not used
 TRANSMITTER C13 sp
 f1h 150.829 mp -2593.1
 f2h 23351 f1 38764.1
 f3h 23351 f1 1416.3
 pw 3.800 f1p -117.0
 DECOUPLER H1 1p -264.8
 dn 0 WC PLOT 250
 dot 0
 dm yyy sc 3834
 dnm 42
 shoft 12580 d1 cdc ph 20
 dnt

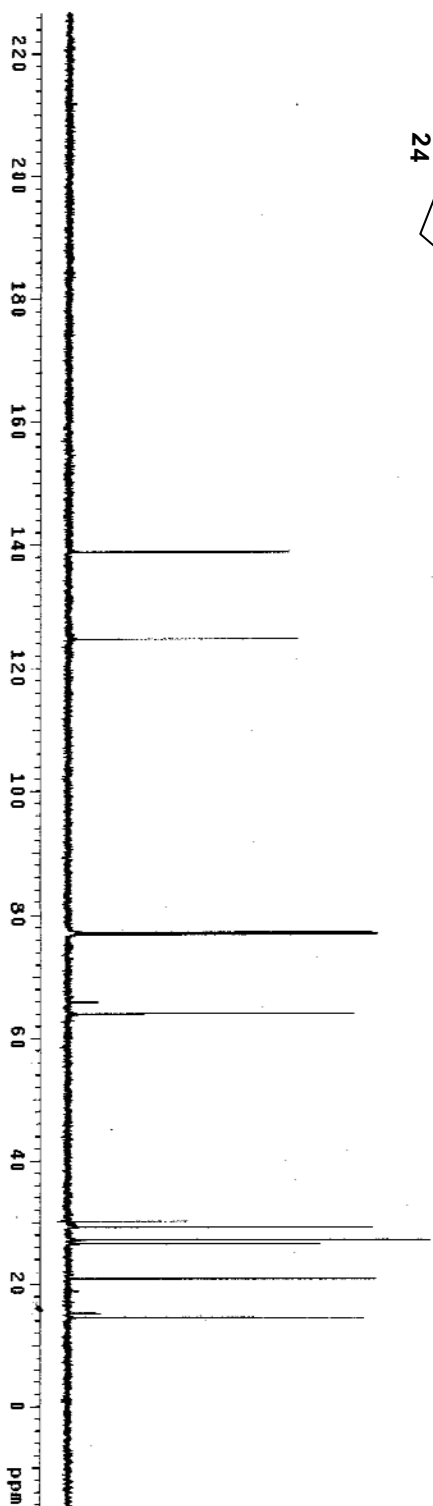
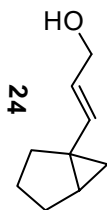
138.803

124.653

77.212
 77.000
 76.788

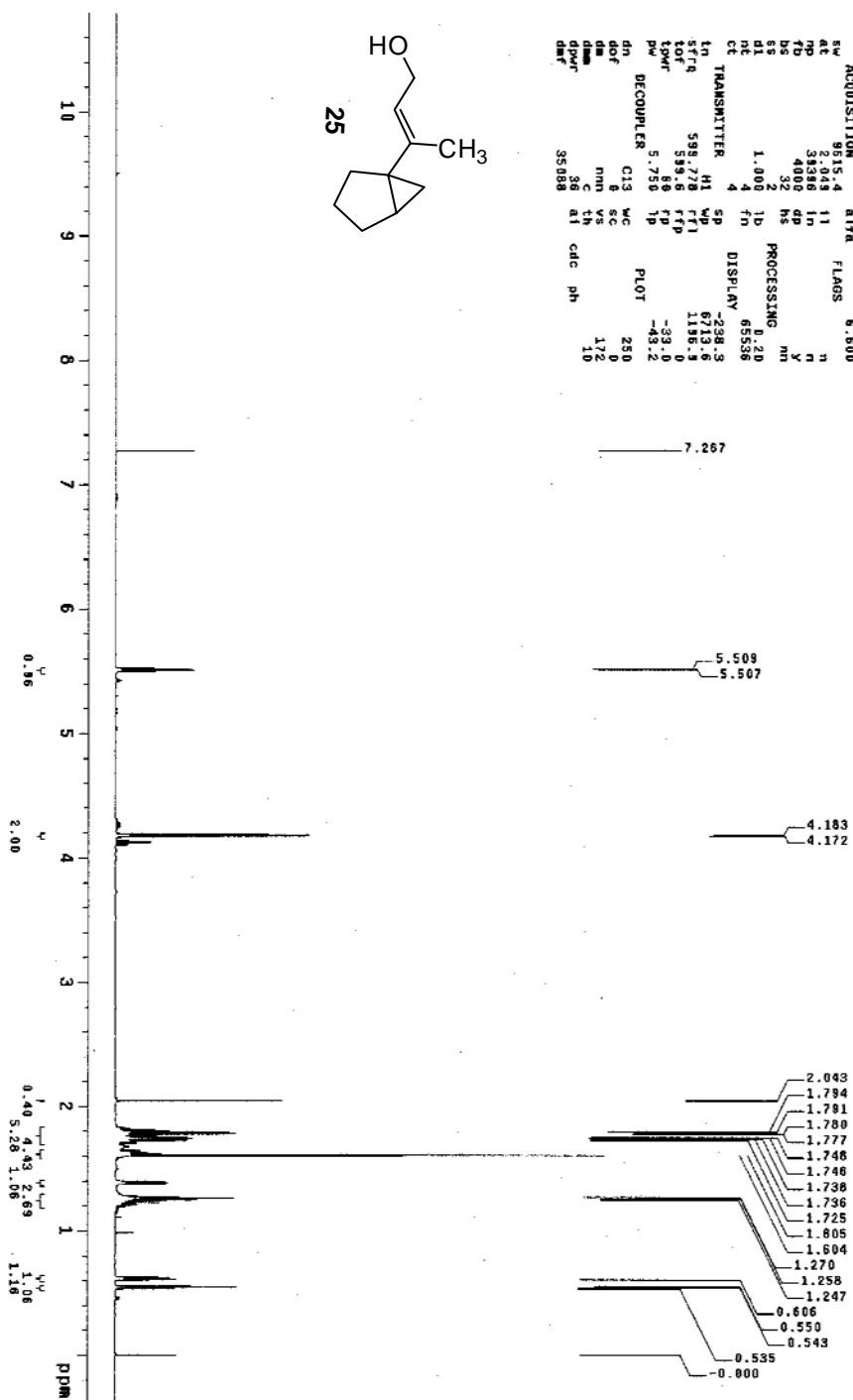
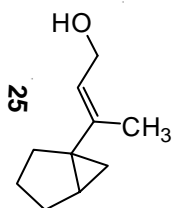
63.892

30.131
 29.220
 27.215
 26.575
 20.843
 14.523



exp3 Proton

SAMPLE										SPECIAL	
date	July 23 2007	temp	not used								
solvent	MeOH	saln									
concn	1000 mg/ml	not used									
Y1/Y2	nm/s	0.008									
nm/s	11,500										
nm/s	6,500										
nm/s	11,500										
nm/s	6,500										
nm/s	11,500										
nm/s	6,500										
nm/s	11,500										
nm/s	6,500										
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nm/s	6,500										
nm/s	11,500										
nm/s	6,500										
nm/s	11,500										
nm/s	6,500										
nm/s	11,500										



Hf-4

expd Carbon

SAMPLE
date Jul 28 2007 temp not used
solvent cdc13 gain 40
file /export/home/~ not used
y1/vmr/sys/date/hu- hst 0.008
eng/hf-4-c.fid pws0 7.600
ACQUISITION
36764.7
11 11
1304 11
95624 in
17000 dp
bs 4 hs
d1 1.000 1b
nt 64 1b
ct TRANSMITTER C13 50
sfreq 150.829 MP
tort 2839.1 rfi
tpr 55 rfp
pw 5.800 tp
DECOUPLER M1 MC
dn 10
scot 250
dnt 0
dnt 7512
dnt 10
12560 at cdc ph

142.166

122.001

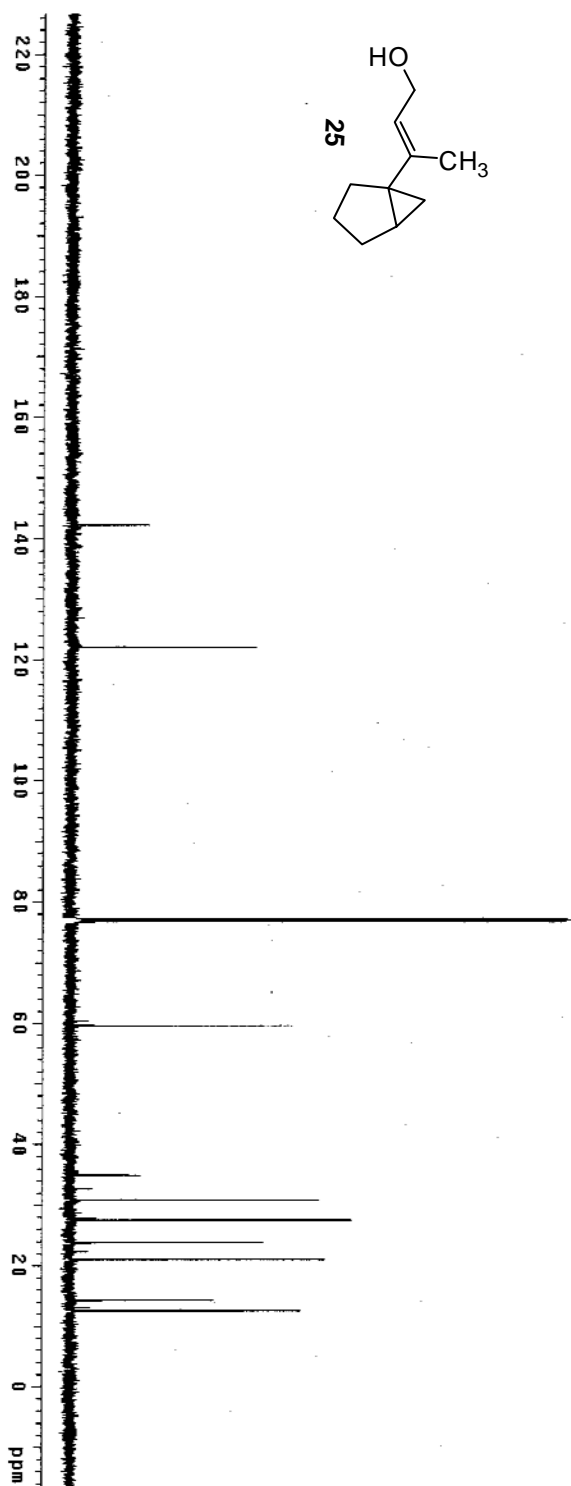
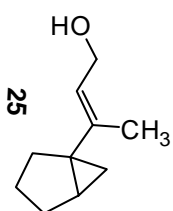
77.212
77.000
76.788

59.551

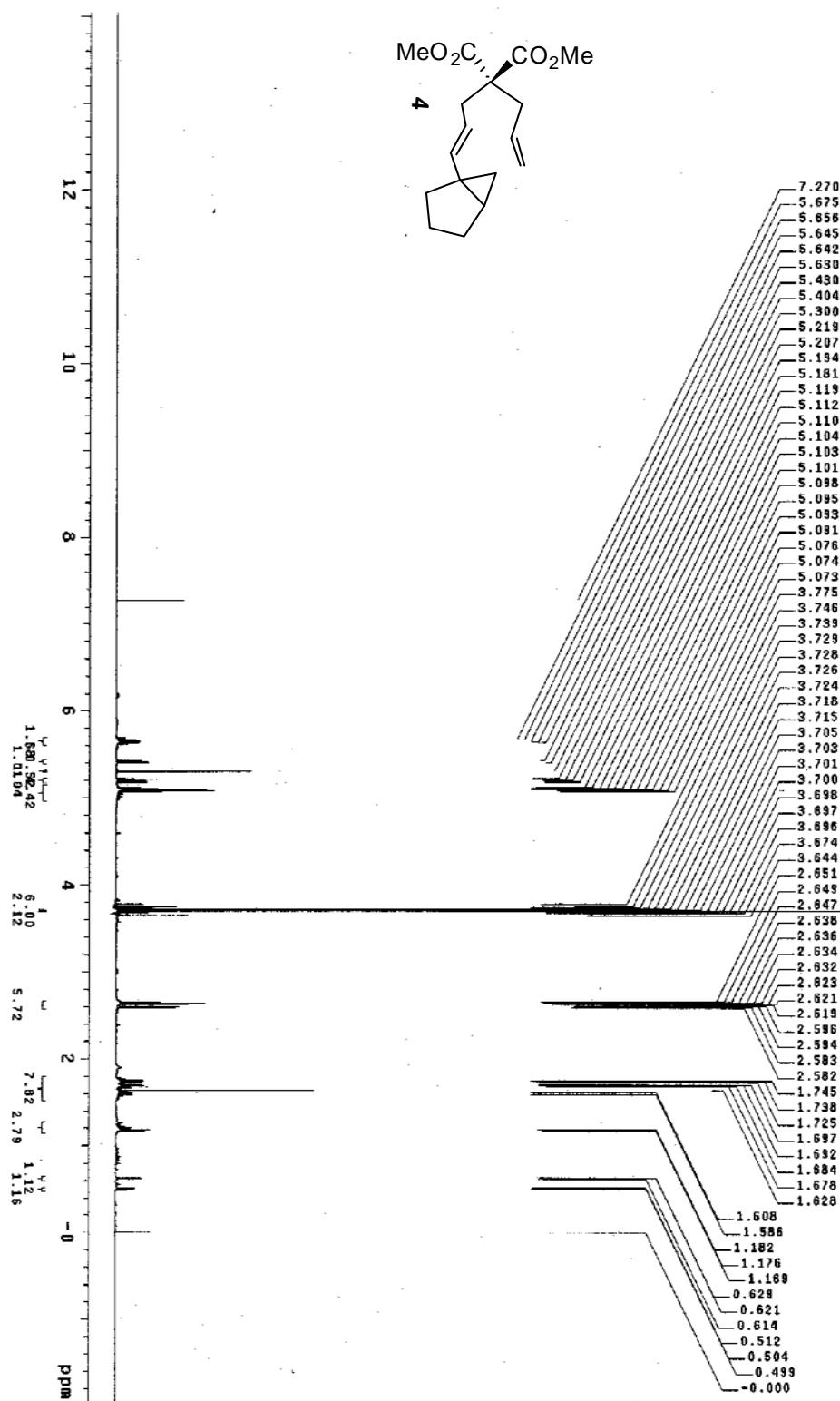
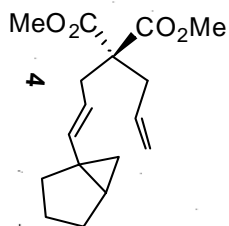
34.882

30.823
27.457
23.760
20.981

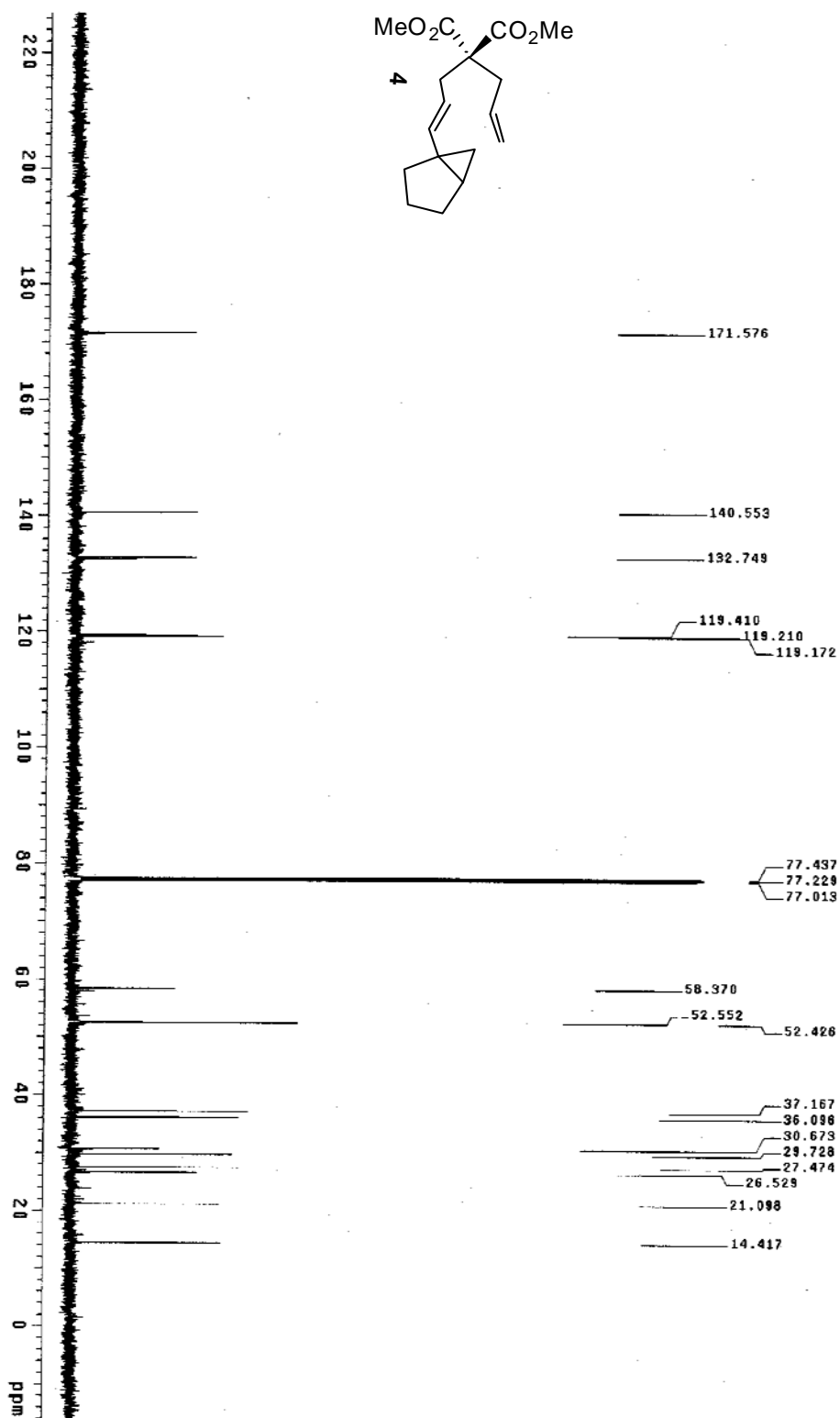
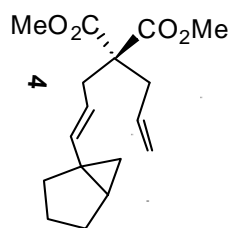
14.271
12.481



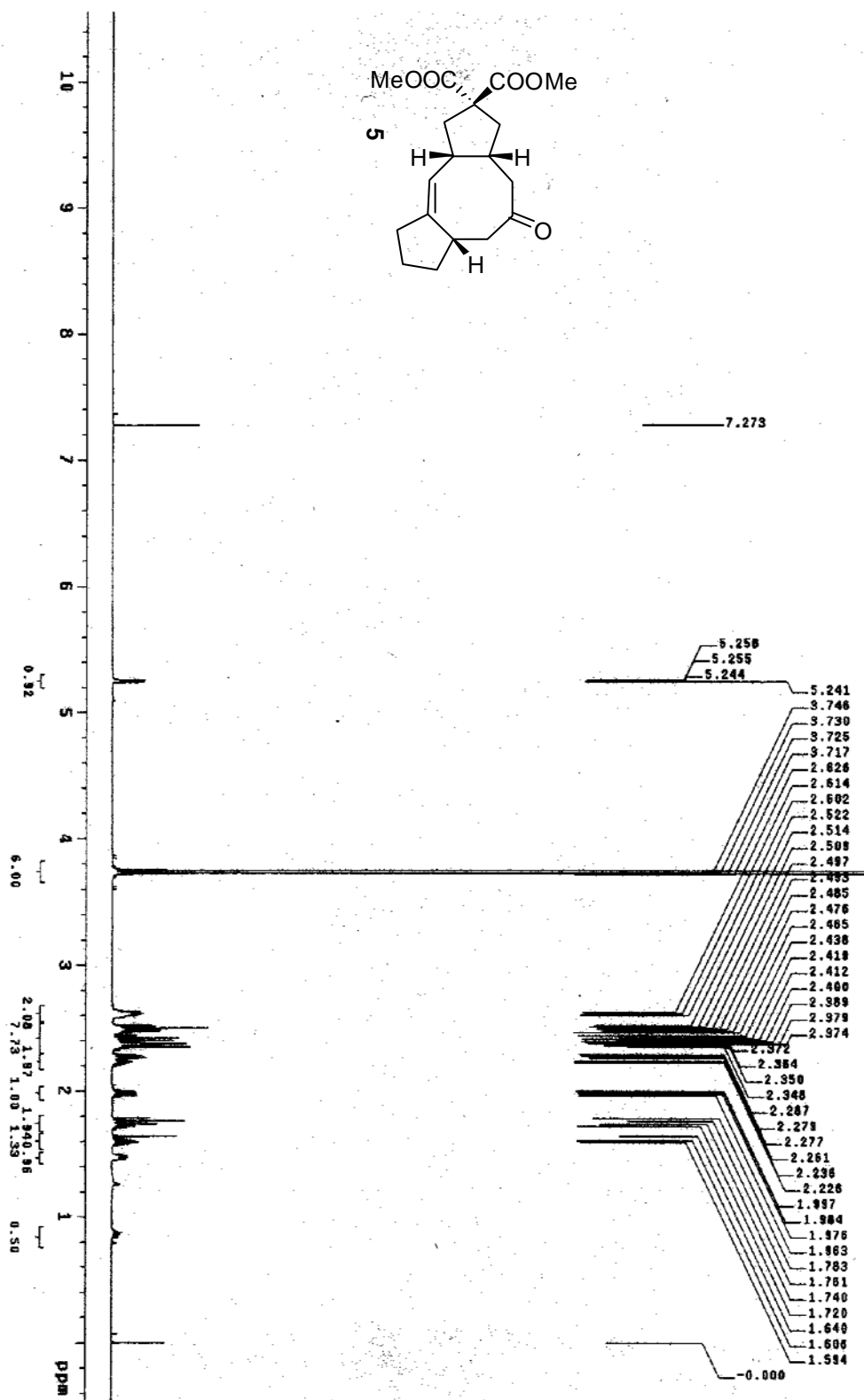
HF-13
 File: HF-13-20070821-H
 Pulse Sequence: s2pu1



HF-13
 File: HF-13-20070803-C
 Pulse Sequence: szpu1

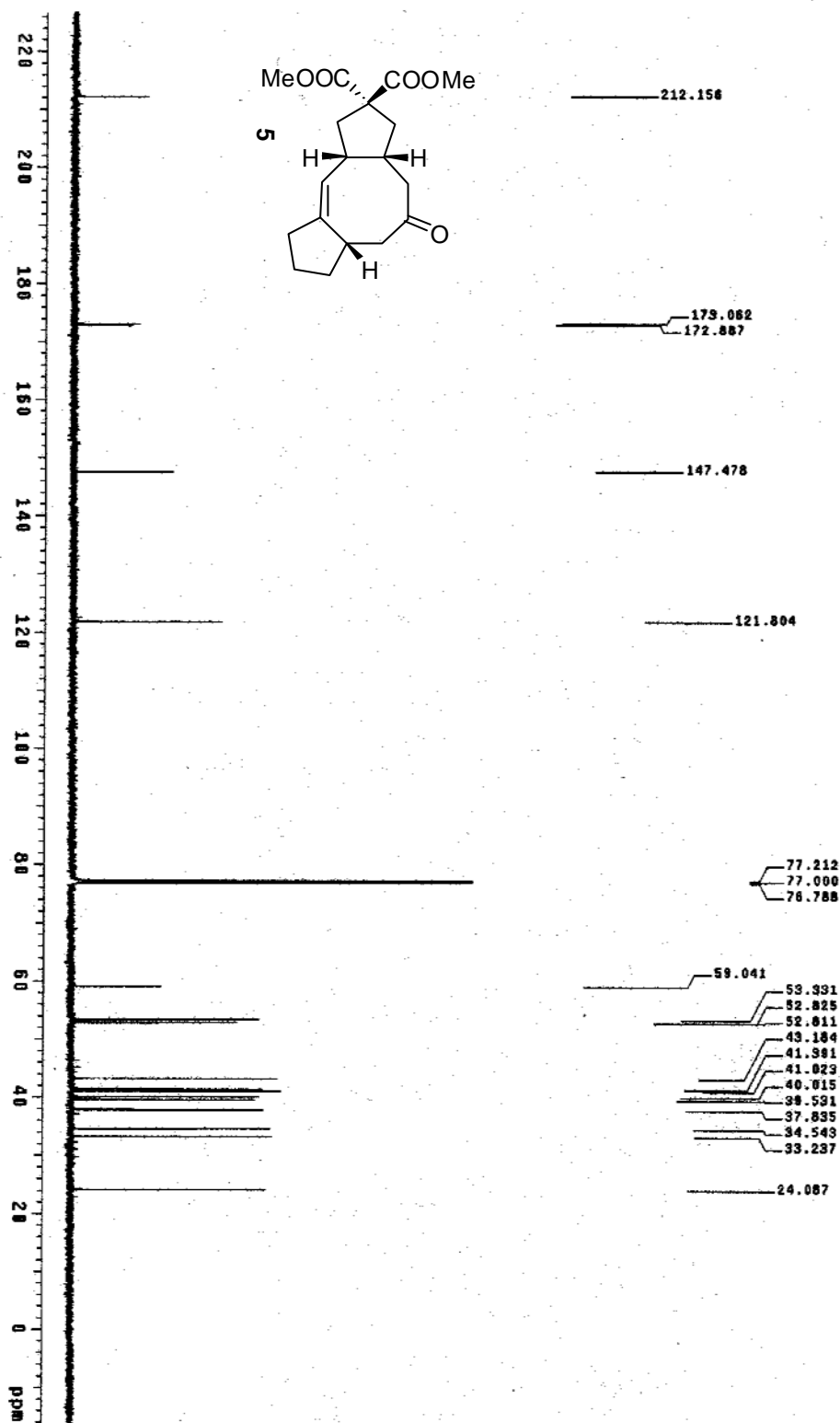


File: Proton
Pulse Sequence: zgpg30

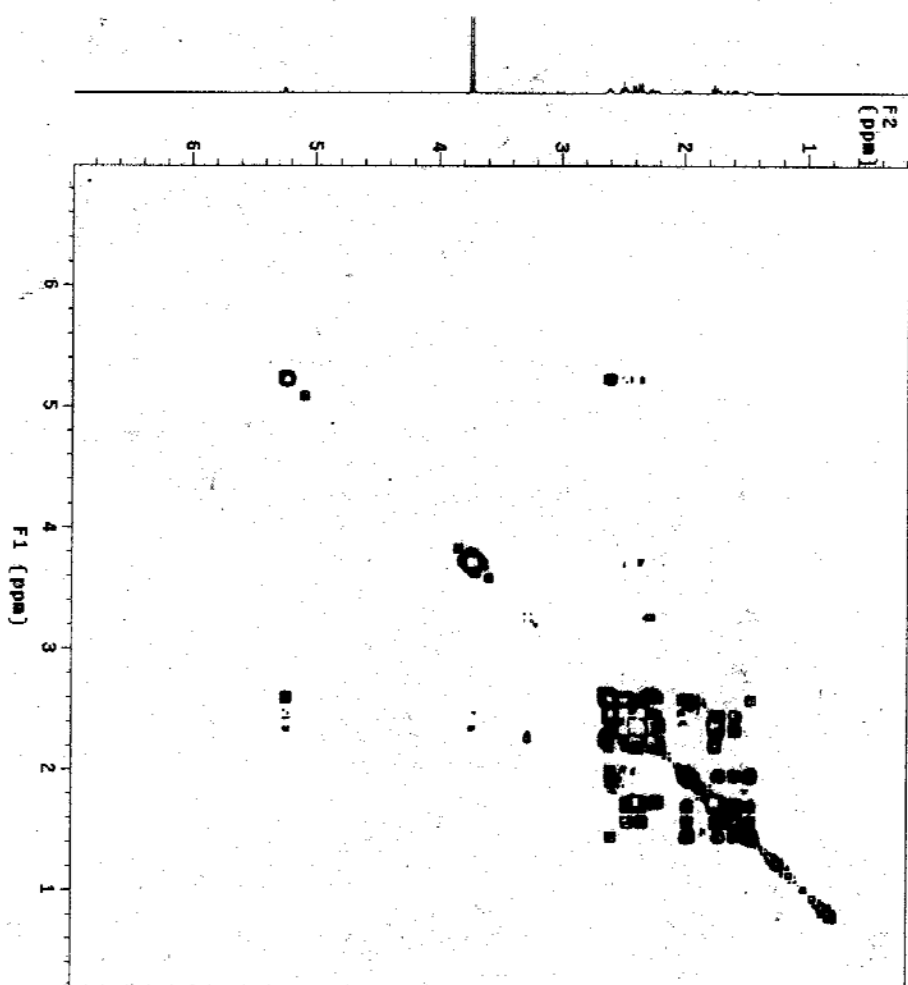
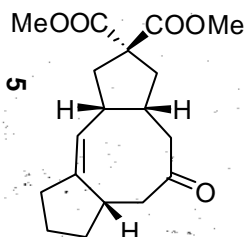


H

File: Carbon
Pulse Sequence: zgpg30

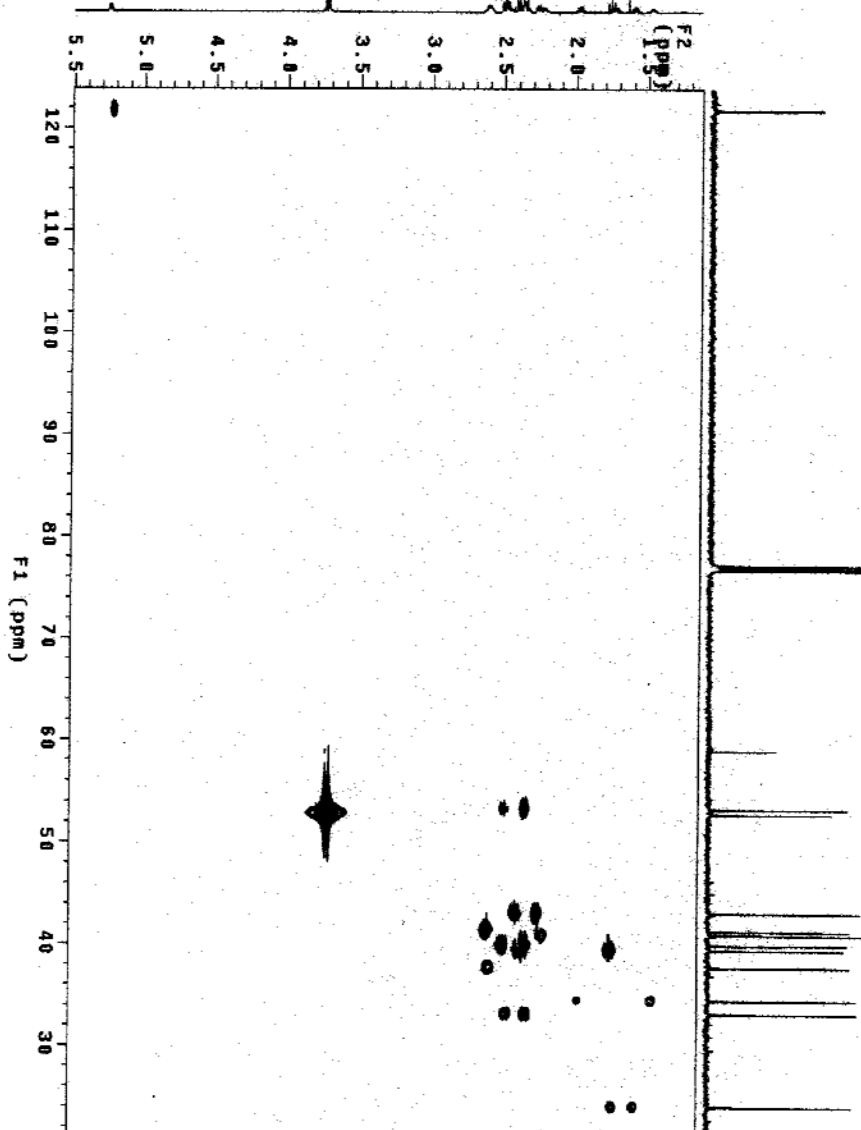
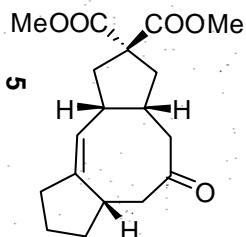


M
 File: Gcasy
 Pulse Sequence: gcosy
 Solvent: cdcl3
 Temp: 25.0 C / 298.1 K
 Operator: zly
 Vmax: 600 MHz
 Relax: delay 1.301 sec
 Acq: time 0.128 sec
 F2: 400.146 MHz
 2D Width 4005.0 Hz
 16 Repetitions
 128 Increments
 OBSERVE: H1, 599.7744908 MHz
 DATA PROCESSING
 Sg: sine bell 0.863 sec
 F2: DATA PROCESSING 0.18 sec
 F1: 120.021 MHz
 Total time 48 min, 45 sec



H

f1: 0h34c
 Pulse Sequence: ghsqc
 Solvent: cdc13
 Temp: 25.0 C / 298.1 K
 Operator: Y1
 VMMS-690 "R13QMR"
 Relax: delay 1.301 sec
 Acq. time 0.189 sec
 Width 4251.7 Hz
 2D Width 25841.0 Hz
 2D Resolution
 256 F2 increments
 OBSERVE N1: 589.7744986 MHz
 DECOUPLE C13: 100.626886 MHz
 Power 38 dB
 on during acquisition
 off during delay
 164.8000000 Hz
 DATA ACQUISITION
 Data: Apodization 0.002 sec
 F1 Data Processing 0.005 sec
 F1 size 488 x 2048
 Total time 27 min, 21 sec



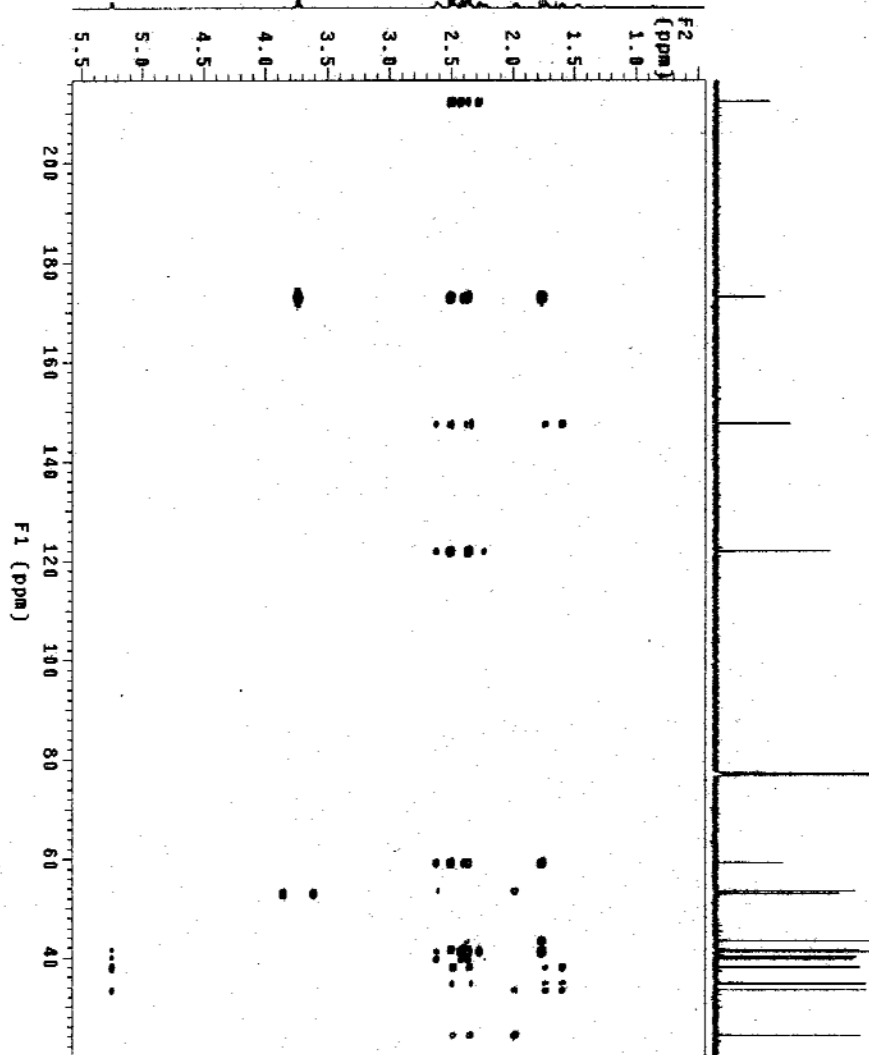
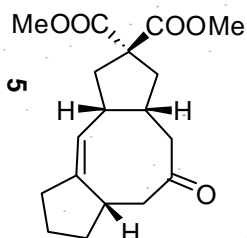
File: Gmmbc

Pulse Sequence: gnm3c
Solvent: cdcl3
Temp: 25.0 C / 298.1 K
Operator: y1
VMRS-500 "RICDMR"

```

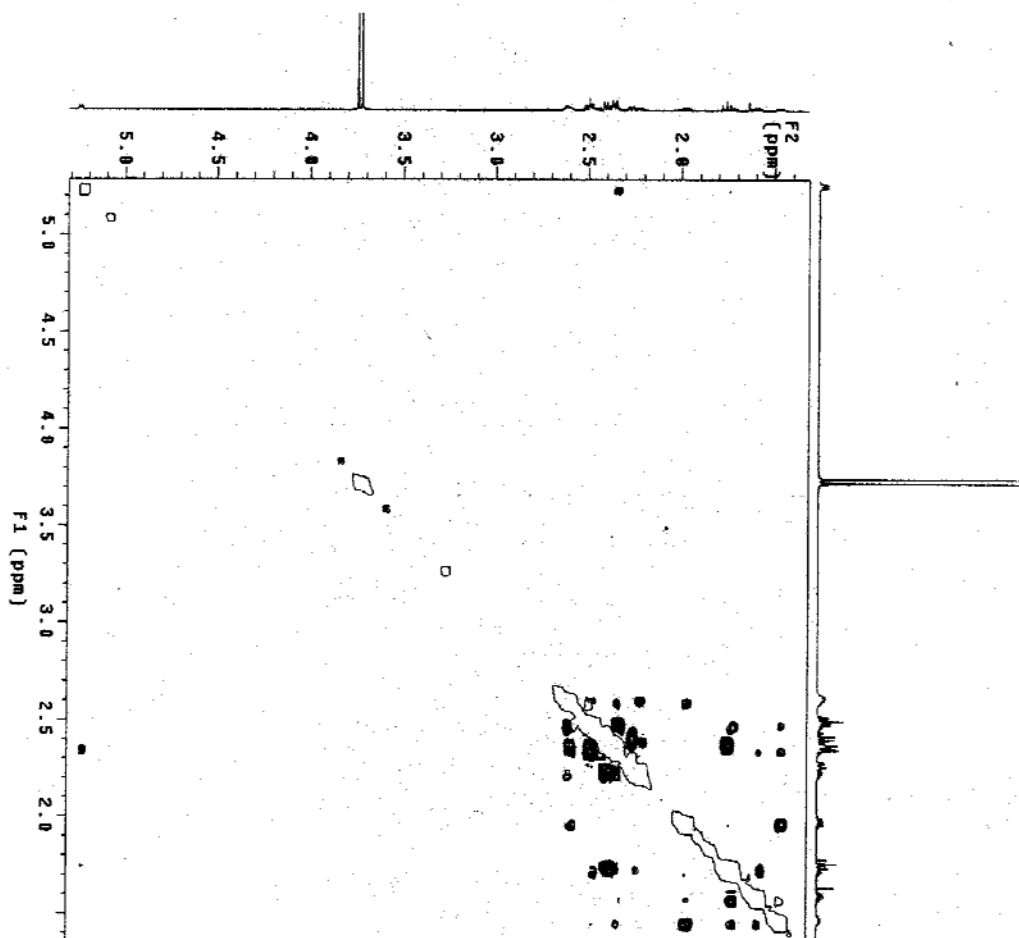
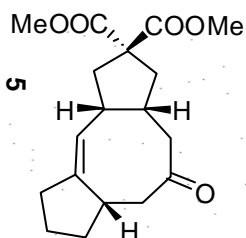
RateX-delay 1.800 sec
Mixing 0.060 sec
Acq. time 0.120 sec
Width 3078.6 Hz
2D Width 23761.4 Hz
8 repetitions
256 increments
DSSEV HI: 559.7744741 MHz
Sine rate 0.0485 sec
DATA PROCESSING
Sine rate 0.0485 sec
Sine rate 0.0485 sec
Sine rate 0.0485 sec
FT size 2048 X 4096
Total time 42 min, 4 sec

```



H

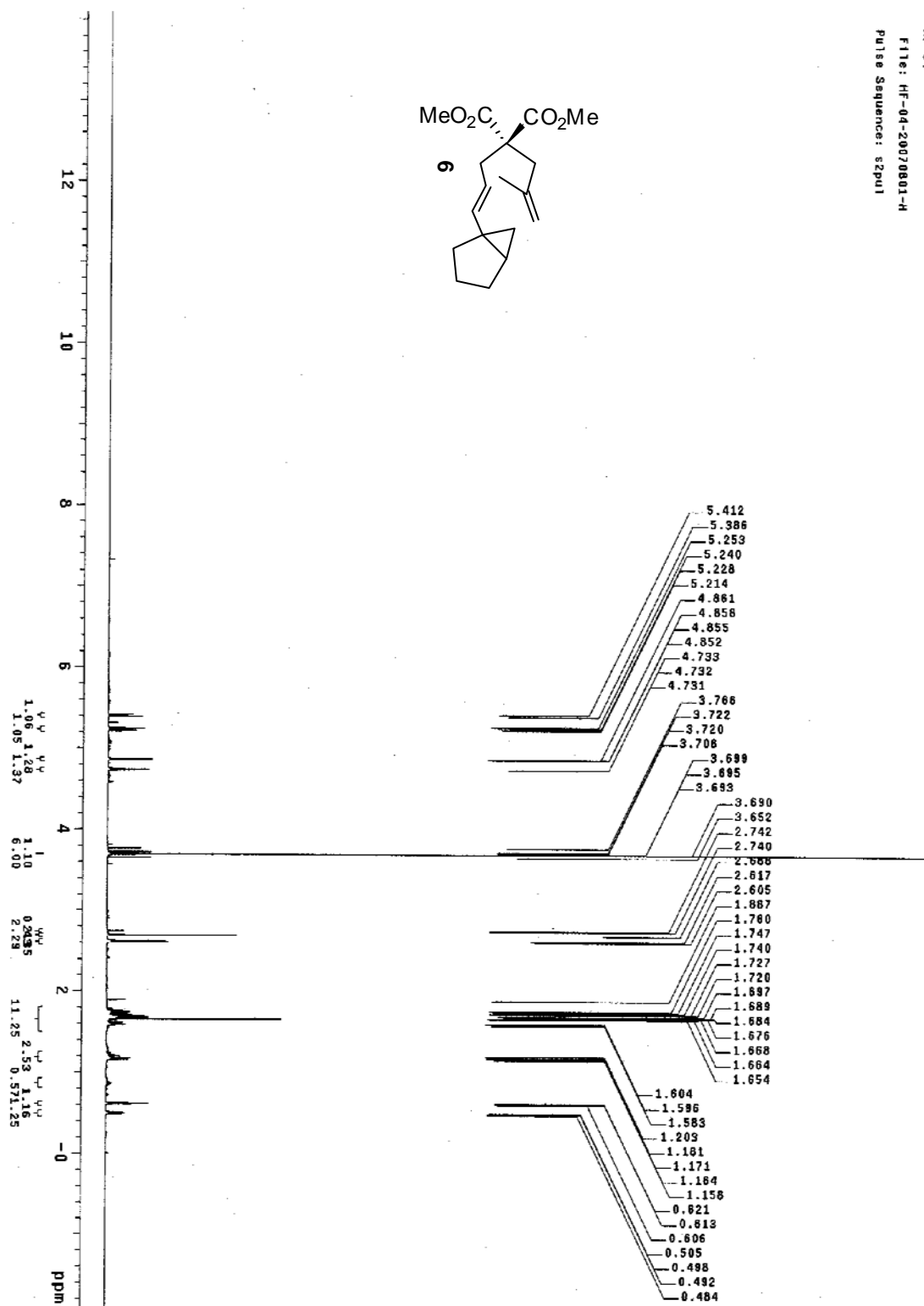
File: Noisy
 Pulse Sequence: MLEV
 Solvent: CDCl3
 Temp: 29.8 C / 298.1 K
 Operator: YI
 VNAME: 008 "RICMNR"
 Date: 4/2/91 1.008 sec
 N1: 500.1305 MHz
 Acq. Time: 8.177 sec
 Width: 2083.5 Hz
 20 Width: 2083.5 Hz
 10 Width: 2083.5 Hz
 20 Width: 2083.5 Hz
 10 Width: 2083.5 Hz
 OBSERVE: N1, 500.1305 MHz
 DATA PROCESSING
 Gauss Apodization: 0.086 sec
 F1 Data Processing: 0.016 sec
 FT Size: 1024 X 1024
 Total time: 2 hr, 16 min, 43 sec



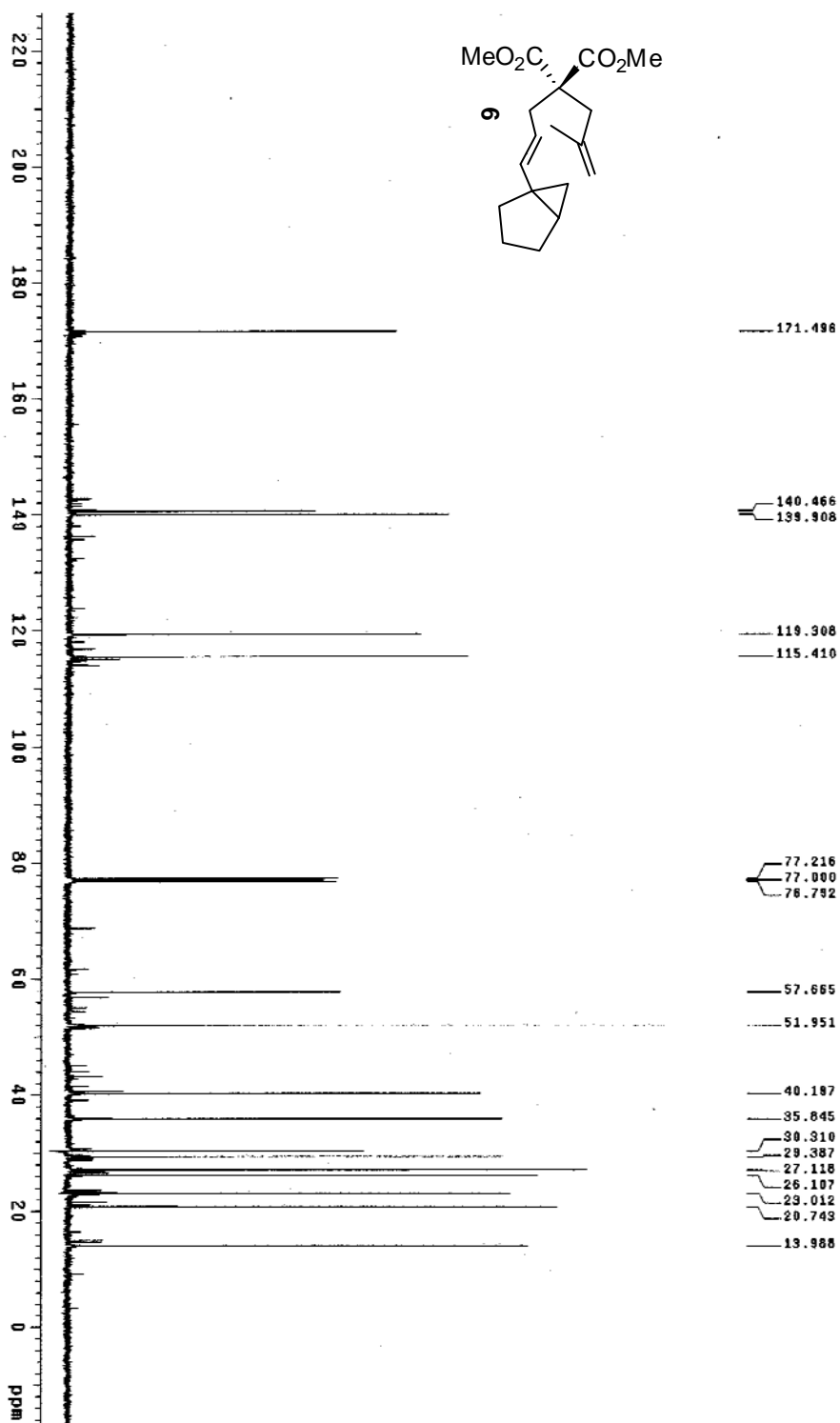
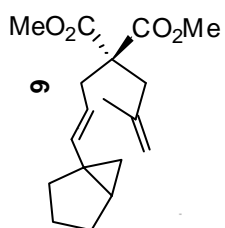
HF-04
 File: HF-04-20070801-H
 Pulse Sequence: szpul1



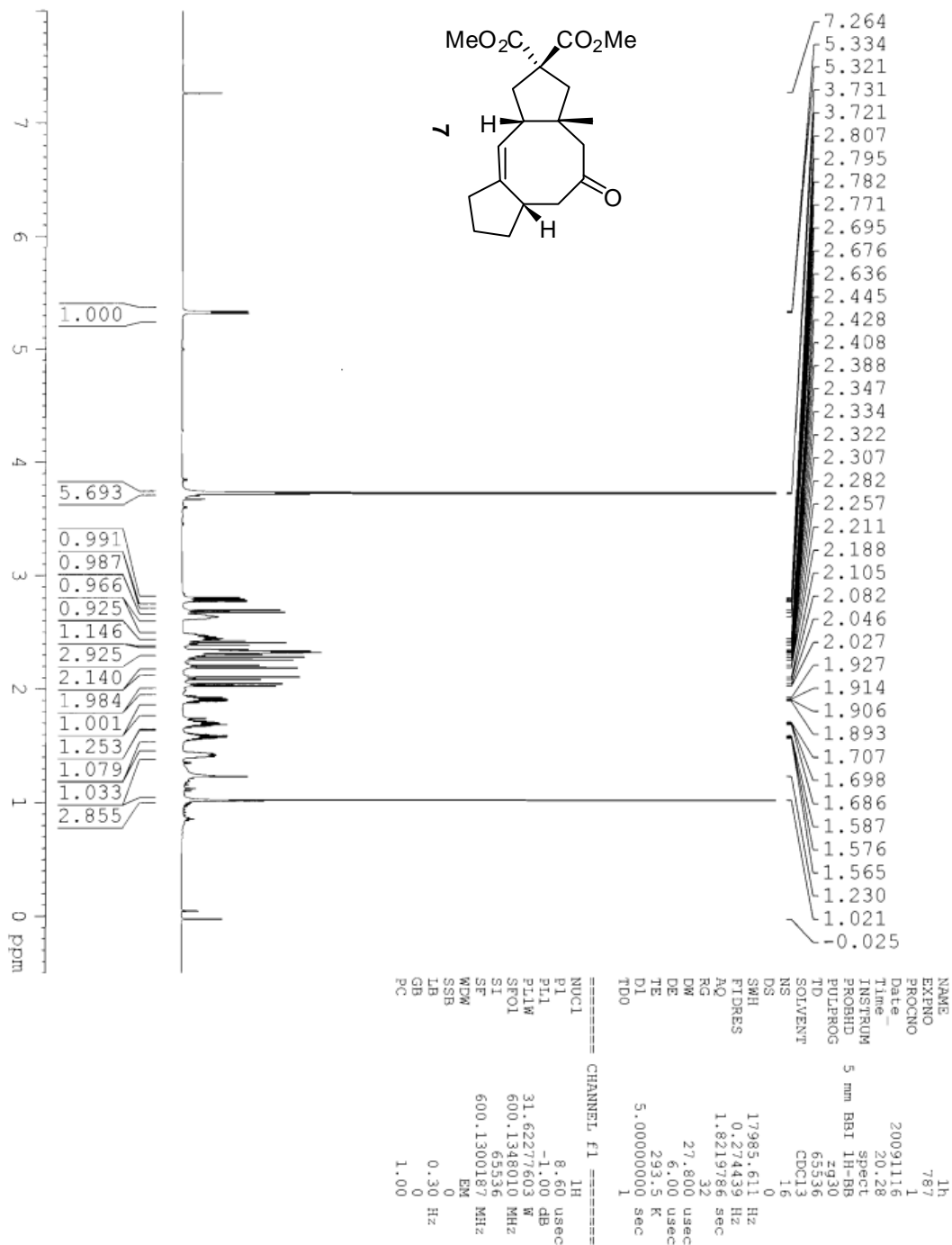
6



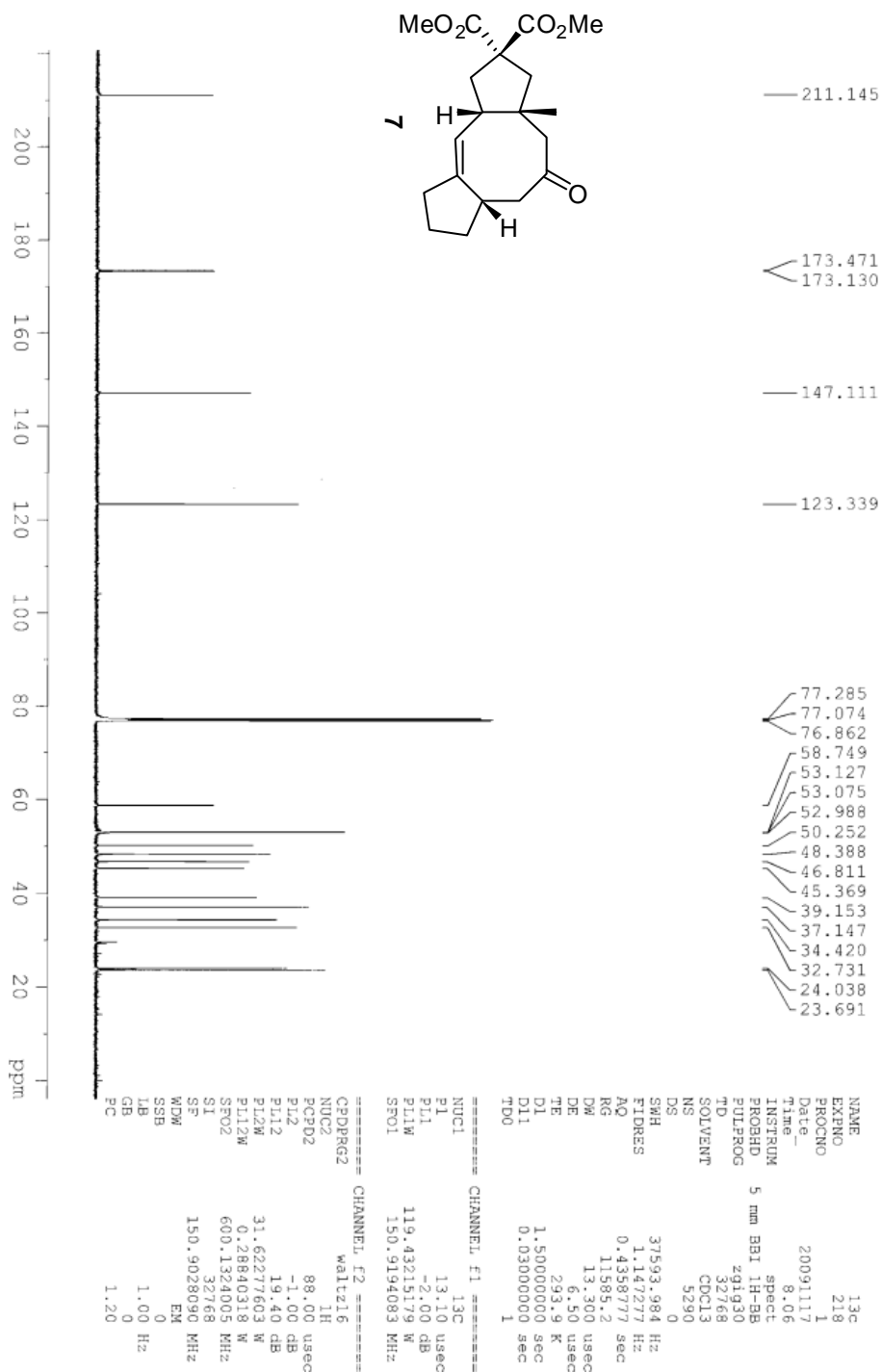
MF-04
 File: MF-04-20070801-C
 Pulse Sequence: zgpg30



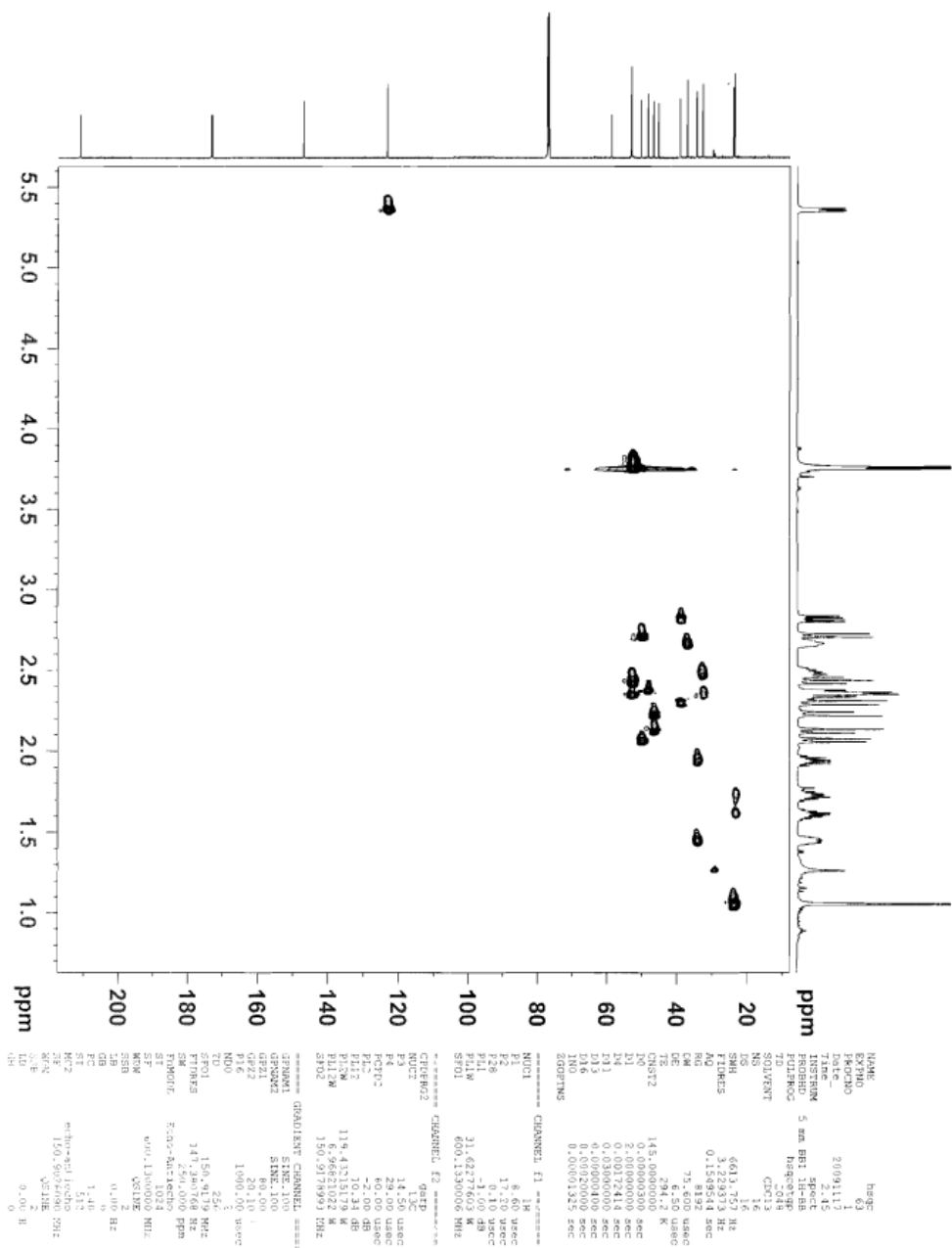
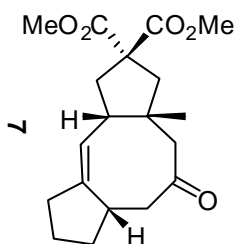
¹H NMR CDCl₃ wy80-B 03616 Yu Zhixiang 2

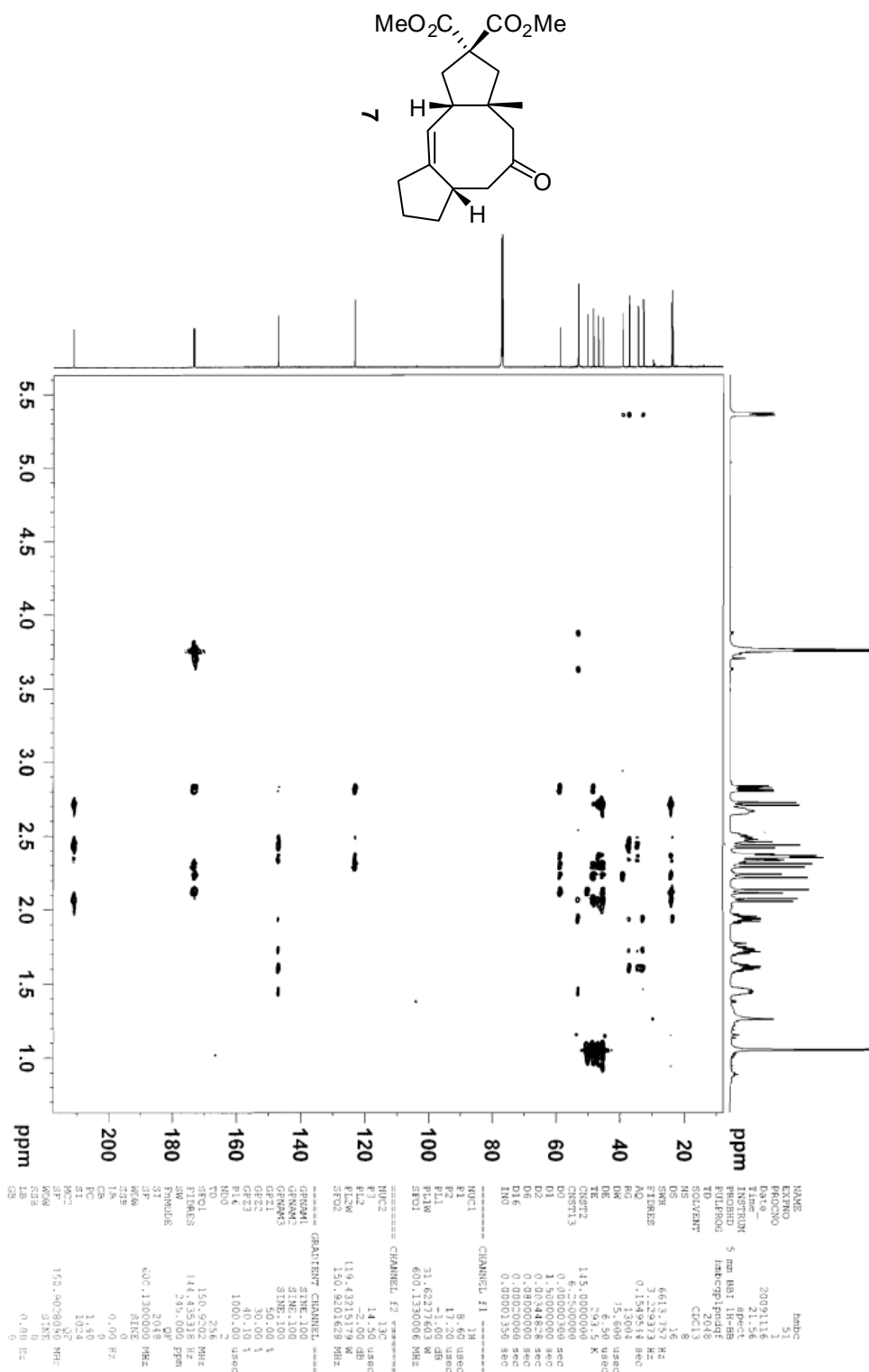


¹³C NMR CDCl₃ wy80-B 03616 Yu Zhixiang 20091116

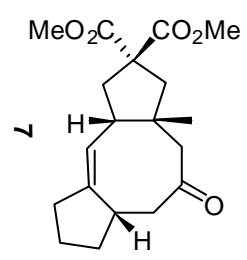
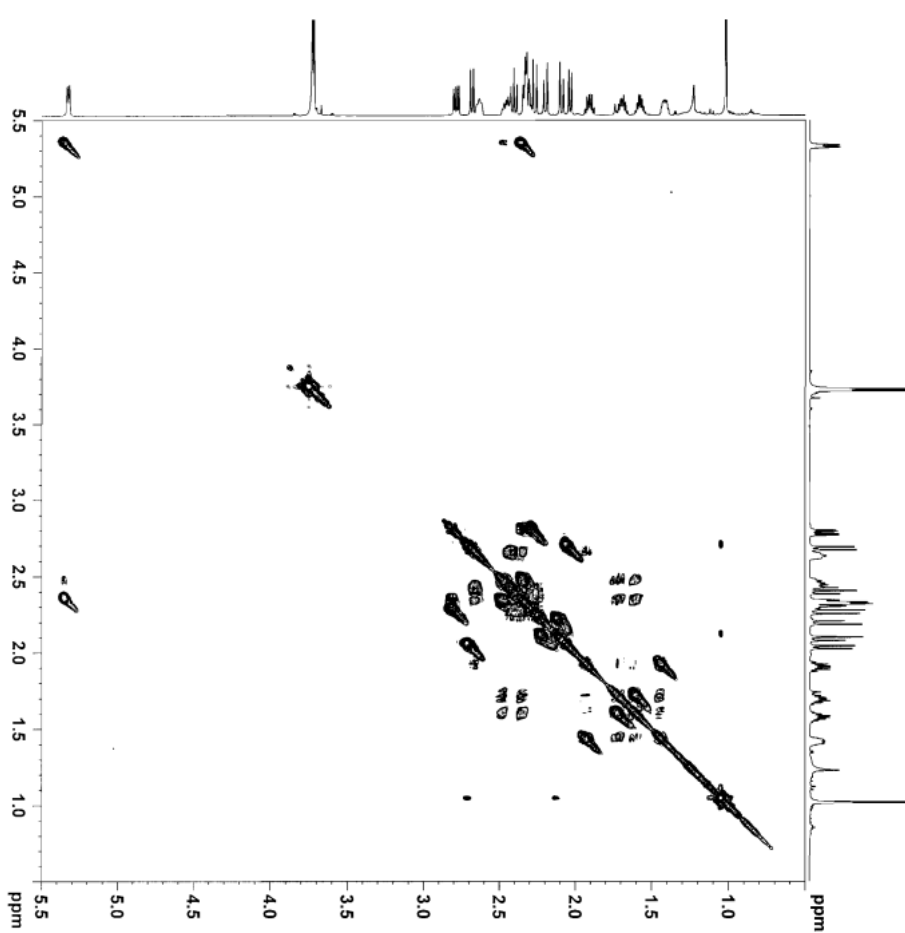


HSQC CDC13 wy80-B 03616 Yu Zhixiang 20091116





COSY CDCl3 wy80-B 03616 Yu Zhixiang 200



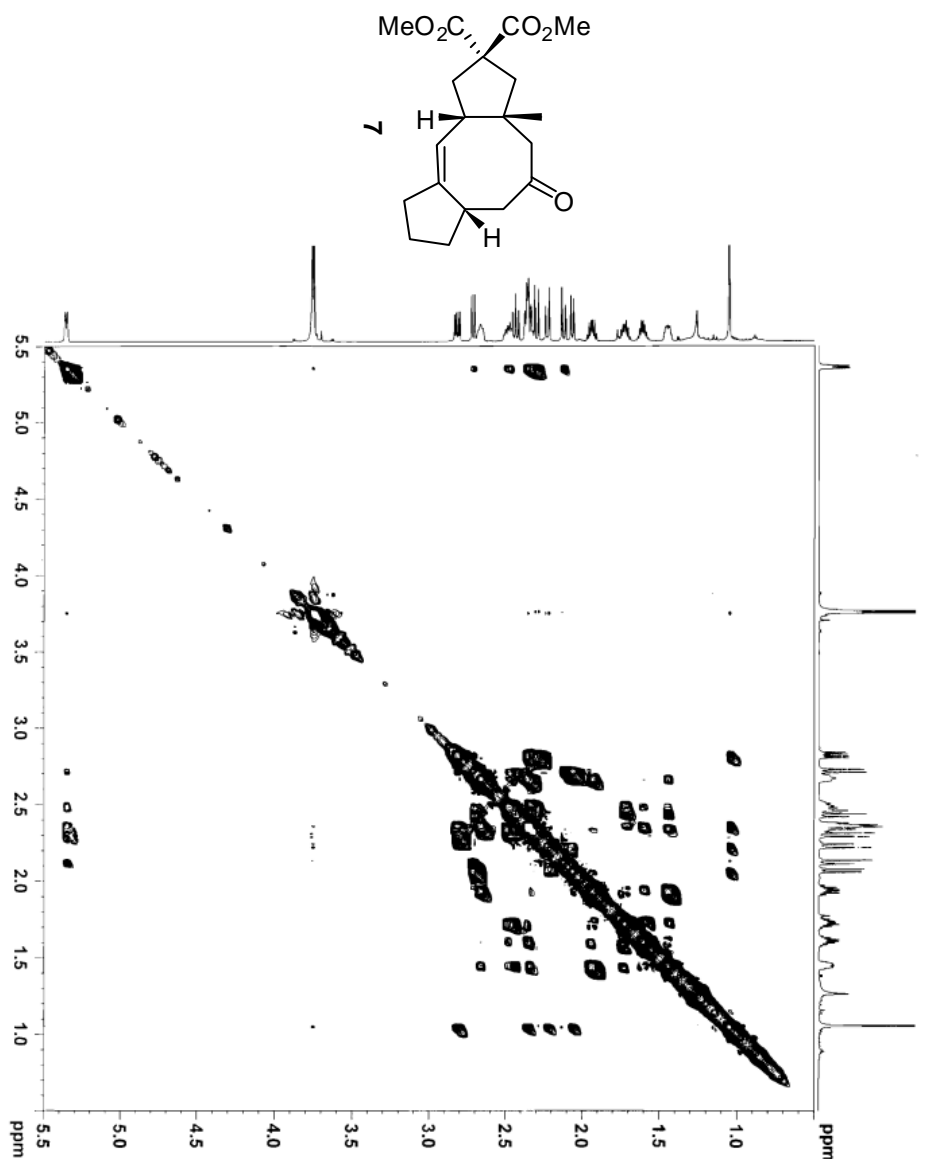
```

NAME          COSY
EXPNO         89
PROCNO        1
Date_         20091117
Time         8.09
INSTRUM       spect
PROBHD        5 mm BBI 1H-4H
PULPROG       zgpg30
TD            2048
SOLVENT       CDCl3
NS            4
DS            16
SWH           6613.757 Hz
FIDRES       3.229373 Hz
AQ           0.1549544 sec
RG            32
DM           75.600 usec
DE           6.50 usec
TE           293.8 K
D0           0.00000300 sec
D1           1.50000000 sec
D13          0.00000400 sec
D16          0.00002000 sec
IN0          0.00015120 sec

===== CHANNEL f1 =====
NUC1          1H
P0            4.30 usec
PL1           -8.60 usec
PT1W         31.62237603 W
SFO1         600.1330006 MHz

===== GRADIENT CHANNEL =====
GPNAM1       SINE.100
GPZ1         10.00 %
P16          1000.00 usec
ND0          1
TD           256
SFO1         600.133 MHz
FIDRES       25.835022 Hz
SW           11.021 ppm
FNUDEC       QF
SI           1024
SF           600.1300000 MHz
WDW          SINE
SSB          0
LB           0.00 Hz
GB          0
PC           1.40
SI           1024
IC2          QF
SF           600.1300000 MHz
WDW          SINE
SSB          0
LB           0.00 Hz
GB          0
  
```

NOESY CDC13 wy80-B 03616 Yu Zhixiang 2C



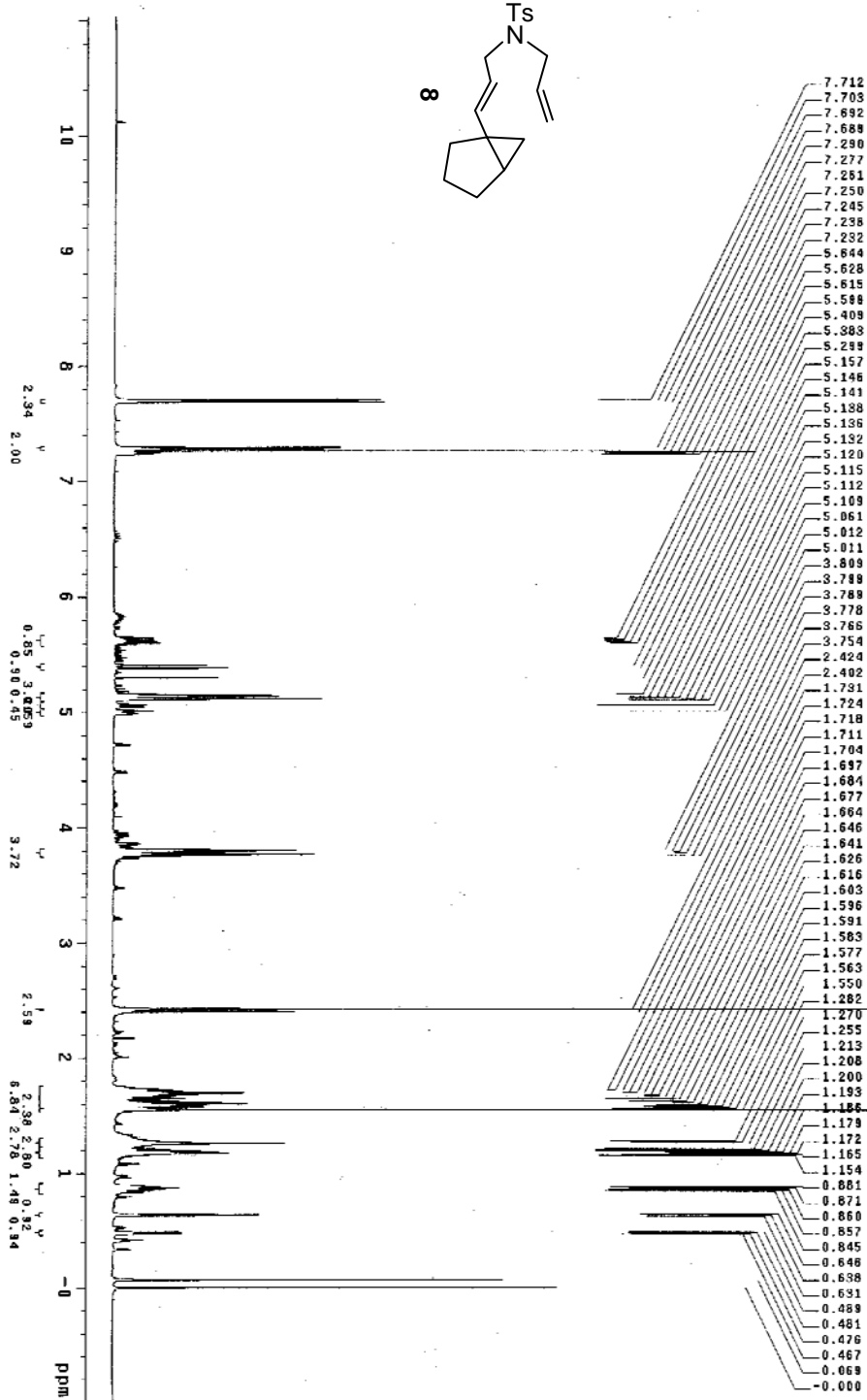
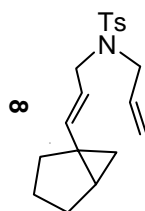
```

NAME          noesy
EXPNO         119
PROCNO        1
Date_         20091117
Time          8.39
INSTRUM       spect
PROBHD        5 mm BBI 1H-BB
PULPROG       noesygpph
TD            2048
SOLVENT       CDC13
NS            16
DS            4
SWH           6613.75 Hz
FIDRES       0.154954 sec
AQ           0.154954 sec
RG            31
DM            75.600 usec
DE            6.50 usec
TE           293.7 K
D0            0.00006465 sec
D1            1.50000000 sec
D8            0.80000001 sec
D16           0.00020000 sec
IN0           0.00015120 sec

===== CHANNEL f1 =====
NUC1          1H
P1            8.60 usec
P2           17.20 usec
PL1          -1.00 dB
PL1W         31.62277603 W
SFO1         600.1330006 MHz

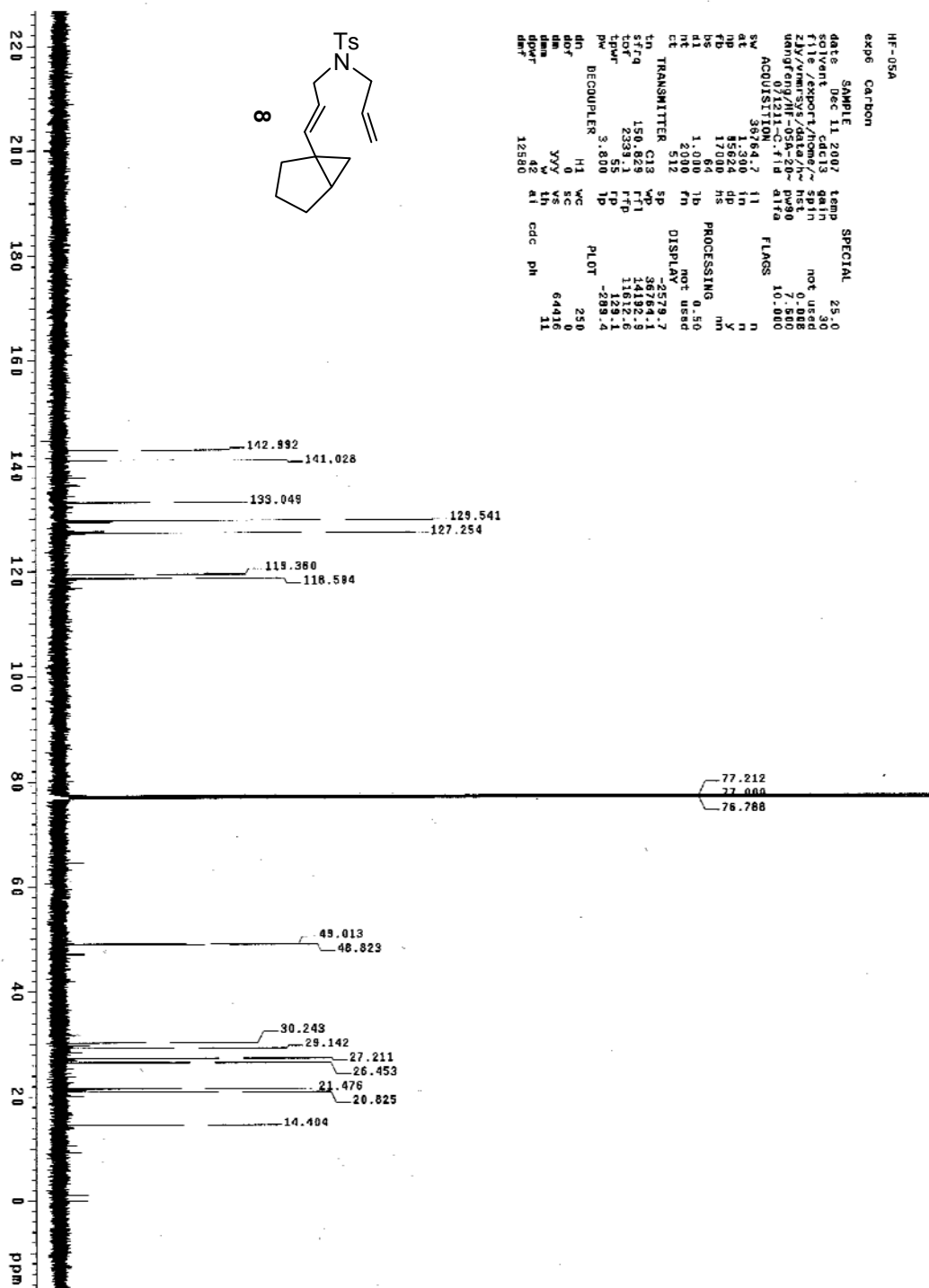
===== GRADIENT CHANNEL =====
GENAM1       SINE.100
SINE.100     SINE.100
GENAM2       40.00 %
GEF2         -40.00 %
P16          1000.00 usec
ND0          1
TD           256
SFO1         600.133 MHz
SADRES       25.835432 Hz
SI           11402 ppm
FMODE        States-TpPI
SI           1024
SF           600.1300000 MHz
WDW          QSINE
SSB          2
LB           0.00 Hz
GB           0
  
```

HF-05A
 File: HF-05A-20071211-H
 Pulse Sequence: s2pu1



HF-05A
expe carbon

SAMPLE
date Dec 11, 2007 temp 25.0
solvent CDCl3
file //export/home/~
zly/vmr/svs/data/hw not used
nanafeng/HF-05A-20~ 0.088
0.7211-C-11d pvs90 7.580
0.7211-C-11d alfa 10.000
ACQUISIT 38764.7 11
SW 1.300 in n
at 1.300 in n
np 35624 dp y
fb 17080 ns
p1 1.000 pb PROCESSING m
nt 2000 fn not used
ct 512 DISPLAY
tn TRANSMITTER C13 SP -2579.7
f1 150.423 F1 34784.1
tor 2338.1 F1 11612.6
tpwr 55 TP 129.1
pw 3.800 TP -293.4
DECOUPLER H1 PLOT 250
dd 0
ddp 0
dam yyy vs 64416
dpr 42 w th 11
dpr 12880 al cdc ph



040612

exp5 Proton

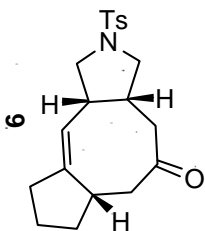
SAMPLE SPECIAL
 date Apr 6 2007 temp not used
 solvent cdc13 gain 8
 file /export/home/~ not used
 y1/vnmr-sys/data/hu- hst 0.008
 dhqfmg/040612-H-1- pw90 11.500
 6.800
 ACQUISITION id 11 f1acs
 sw 9815.4 11
 at 2.043 1n
 op 38386 dp
 fb 4080 hs
 bs 32 1b
 ss 2 1b
 ol 1.000 7n
 nt 4 5p
 cs 4 5p
 tn TRANSMITTER H1 f1
 stf4 559.778 f1d
 tof 559.6 f1d
 tpwr 60 1p
 pw 5.750 1p
 DECOUPLER C13 wc
 do 250
 dof 250
 dm 250
 dmf 35088
 ph 18

7.725
 7.712
 7.347
 7.333
 7.282

3.398
 3.382

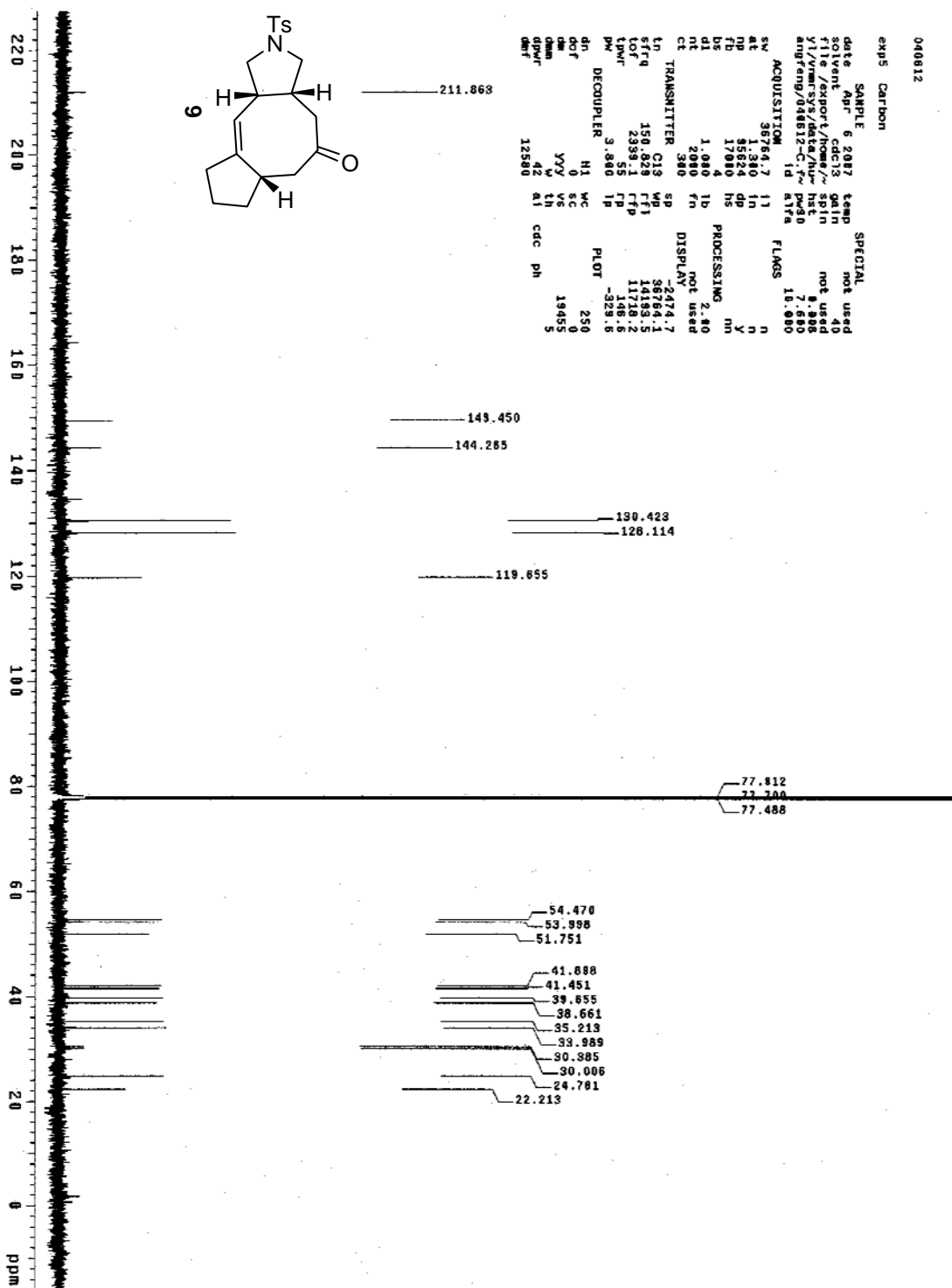
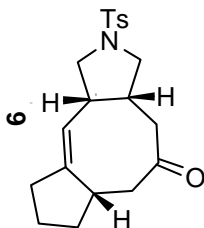
2.453

0.069
 0.000



exp5 Carbon

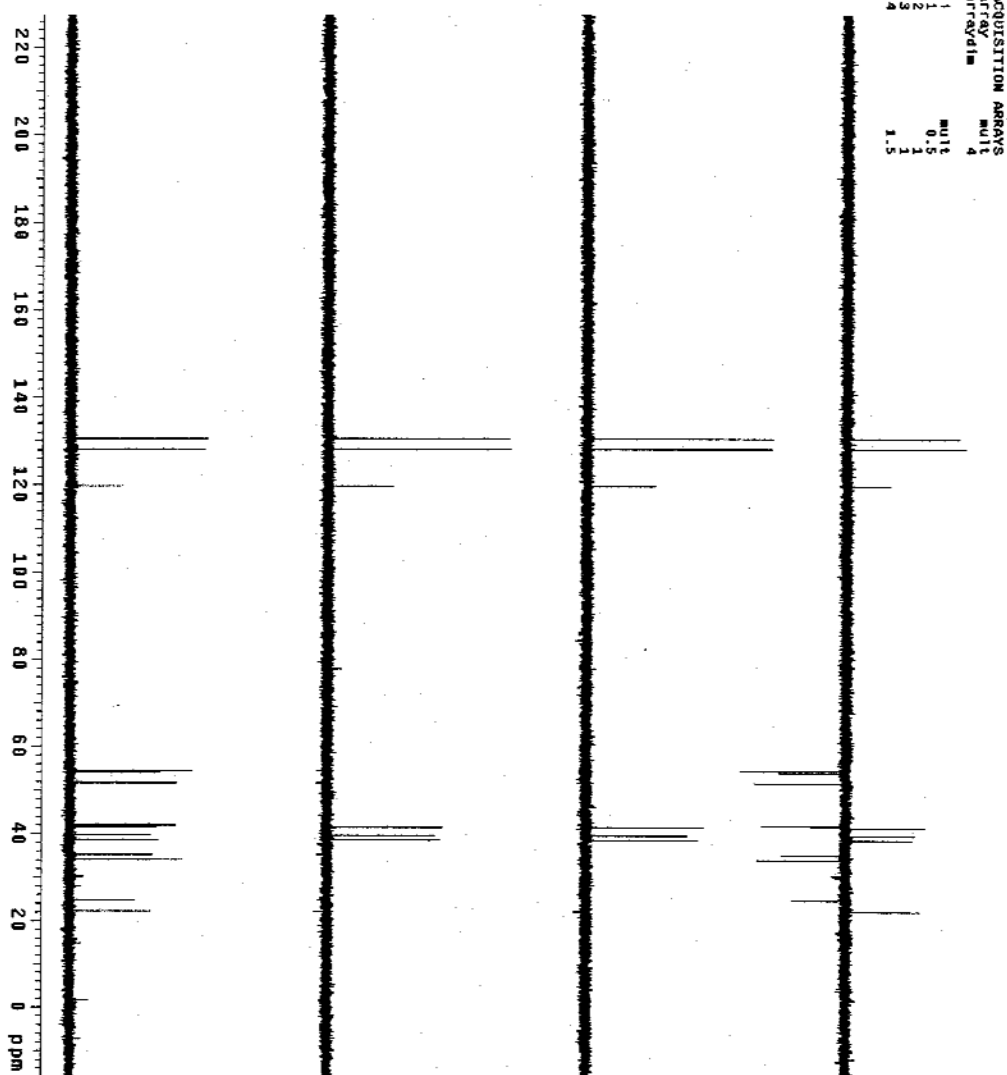
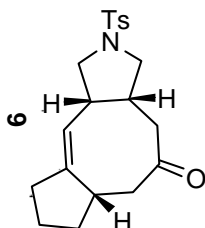
SAMPLE		APR 6 2017		temp		SPECIAL	
date	Apr 6 2017	date	Apr 6 2017	temp	not used	temp	not used
solvent	CDCl3	solvent	CDCl3	gain	40	gain	40
file	/export/home/	file	/export/home/	spin	not used	spin	not used
yl/vnmr	sys/date/	yl/vnmr	sys/date/	nsf	9.886	nsf	9.886
acq/meth	g/400h12-c	acq/meth	g/400h12-c	h1a	15.000	h1a	15.000
ACQUISITION				id	h1a	id	h1a
sv	36764.7	sv	36764.7	1	1	1	1
np	1,310	np	1,310	dp	n	dp	n
at	5652.4	at	5652.4	dp	y	dp	y
fb	1,010	fb	1,010	hs	n	hs	n
ds	1,000	ds	1,000	fr	n	fr	n
nt	2000	nt	2000	fr	n	fr	n
ct	360	ct	360	fr	n	fr	n
TRANSMITTER				display	not used	display	not used
tn	C13	tn	C13	sp	5	sp	5
strq	150.828	strq	150.828	ft1	36764.1	ft1	36764.1
lot	2359.1	lot	2359.1	ftp	11419.5	ftp	11419.5
lpr	3	lpr	3	tp	11718.2	tp	11718.2
pw	3.860	pw	3.860	tp	-328.8	tp	-328.8
DECOUPLER				plot	250.0	plot	250.0
dn	H1	dn	H1	wc	5	wc	5
dot	0	dot	0	sc	19495.5	sc	19495.5
dm	Y/Y	dm	Y/Y	th	5	th	5
opt	42	opt	42	at	cdc	at	cdc
amt	12500	amt	12500	ph	ph	ph	ph



040612

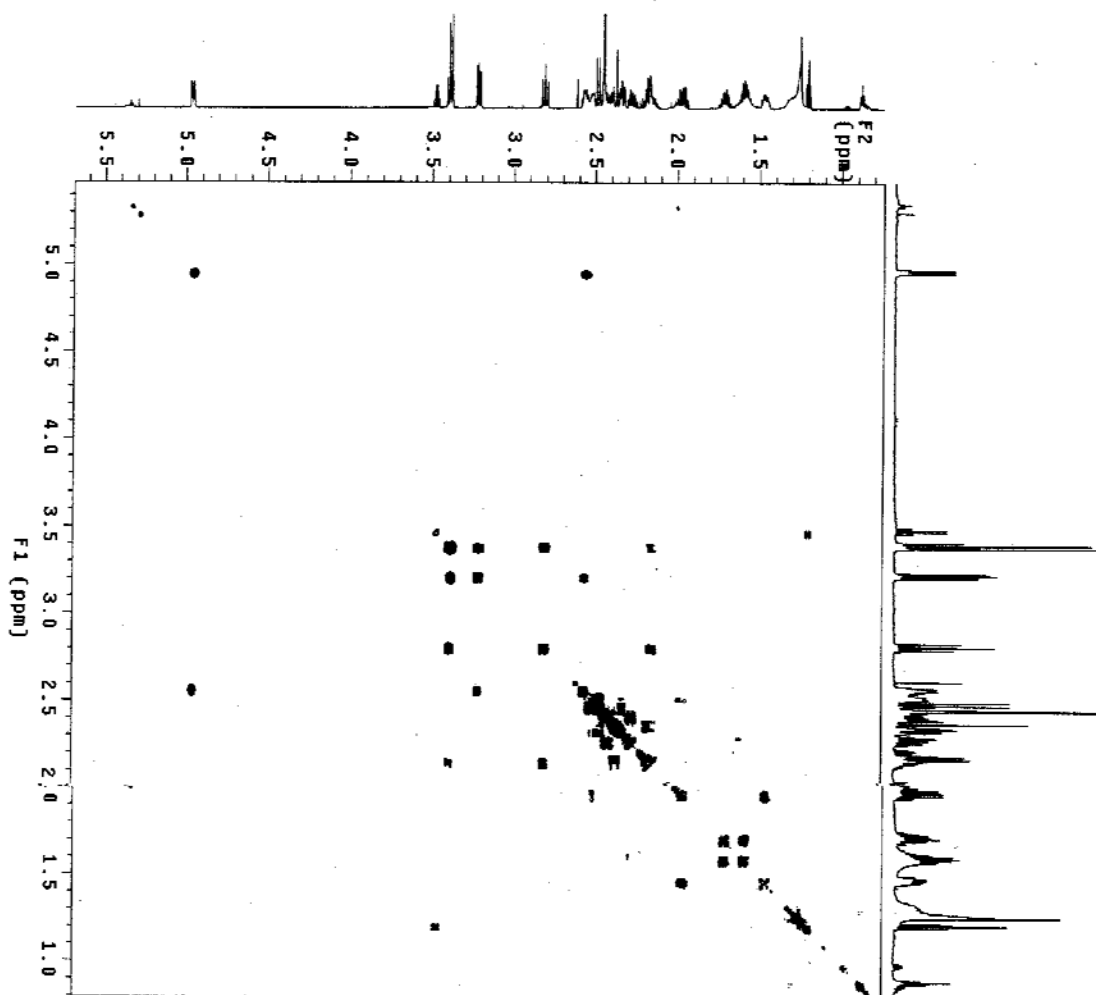
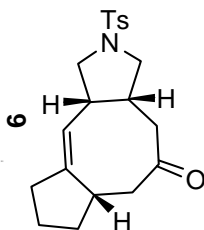
exp3 Dept

SAMPLE		DEPT		ACQUISITION ARRAYS	
date	9 2087	jkh	140.0	array	mult
solvent	cdcl3	mult	arrayed	array	4
sample	cdcl3	mult	not used	array	1
sv	38764.7	temp	20	mult	0.5
acq	1.000	sp	20	mult	0.1
np	73530	sp	20	mult	1.1
bs	64	td	1.00	mult	1.5
ss	2	td	65336		
nt	1.000	wd	48465.6		
ct	300	sp	-2475.2		
tn	2339.1	ai	-154.6		
tof	2339.1	ai	-18.6		
cpwr	7.600	rfi	22445.4		
decoupler	h1	rfp	13665.1		
dn	0	mc	189		
dpwr	42	sc	34081		
dm	my	vs	204.24		
dmr	12580	h2m	7		
pp1v1	15.500	th			



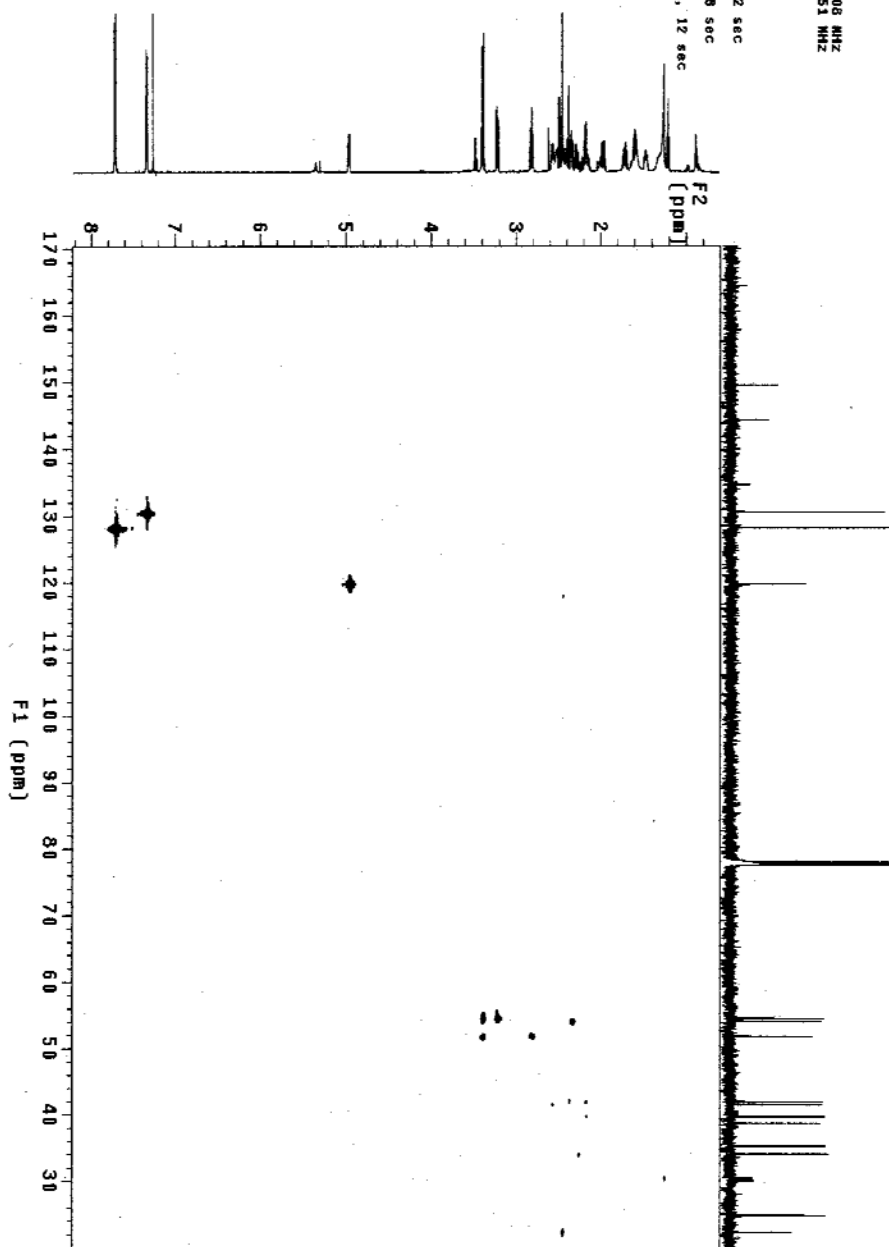
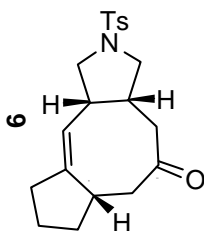
040612

File: 040612-cosy
 Pulse Sequence: gcosy
 Solvent: cdcl3
 Temp: 29.0 C / 298.1 K
 2D F1 (ppm) F2 (ppm)
 F1: 409.6
 F2: 409.6
 VMRS-600 "RICEMMR"
 Relax: delay 1.301 sec
 Acq: time 8.221 sec
 Width 4629.6 Hz
 2D Width 4629.6 Hz
 4 repetitions
 256 increments
 OBSERVE: H1: 399.774938 MHz
 OBSERVE: H2: 399.774938 MHz
 DATA PROCESSING
 Sine bell 0.111 sec
 F1 DATA PROCESSING
 Sine bell 0.111 sec
 F1 size 4096 x 4096
 Total time 27 min, 4 sec



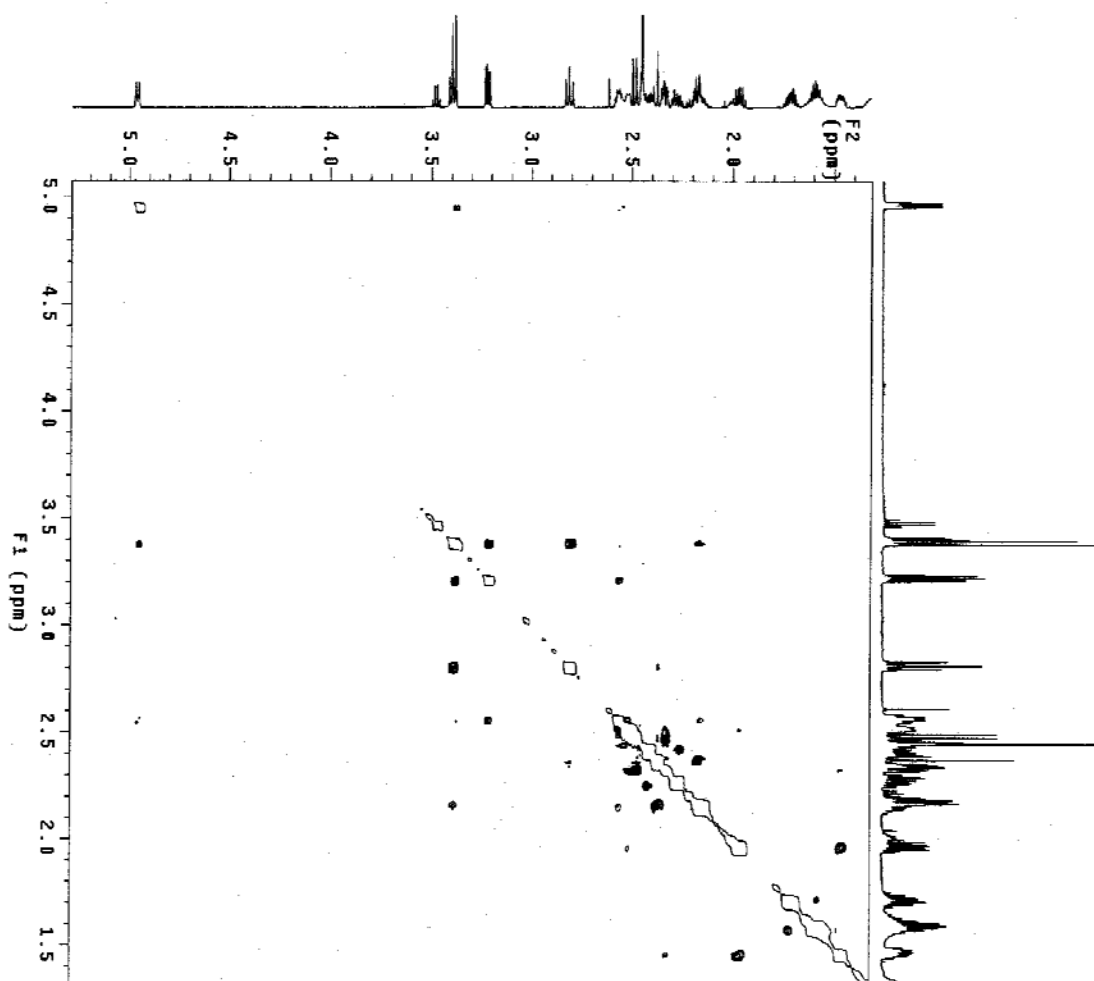
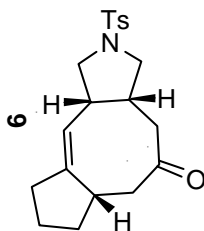
040612

File: 040612-hsac
Pulse Sequence: gmsqc
Solvent: cdcl3
Temp: 25.0 C / 238.1 K
Operator: YJ
File: 040612-hsac
NAME: 000 "RICDMR"
Relax. delay 1.981 sec
Acq. time 484.2 sec
Width 39487.8 Hz
2D Width 39487.8 Hz
8 repetitions
512 increments
OBSERVE N1, 539.7744908 MHz
DECOUPLE C13, 150.8310851 MHz
Power 36 dB
on during acquisition
or during relaxation
DATA PROCESSING
Gauss apodization 0.032 sec
F1 DATA PROCESSING
Gauss apodization 0.008 sec
FT size 4096 x 4096
Total time 1 hr, 47 min, 12 sec




040512

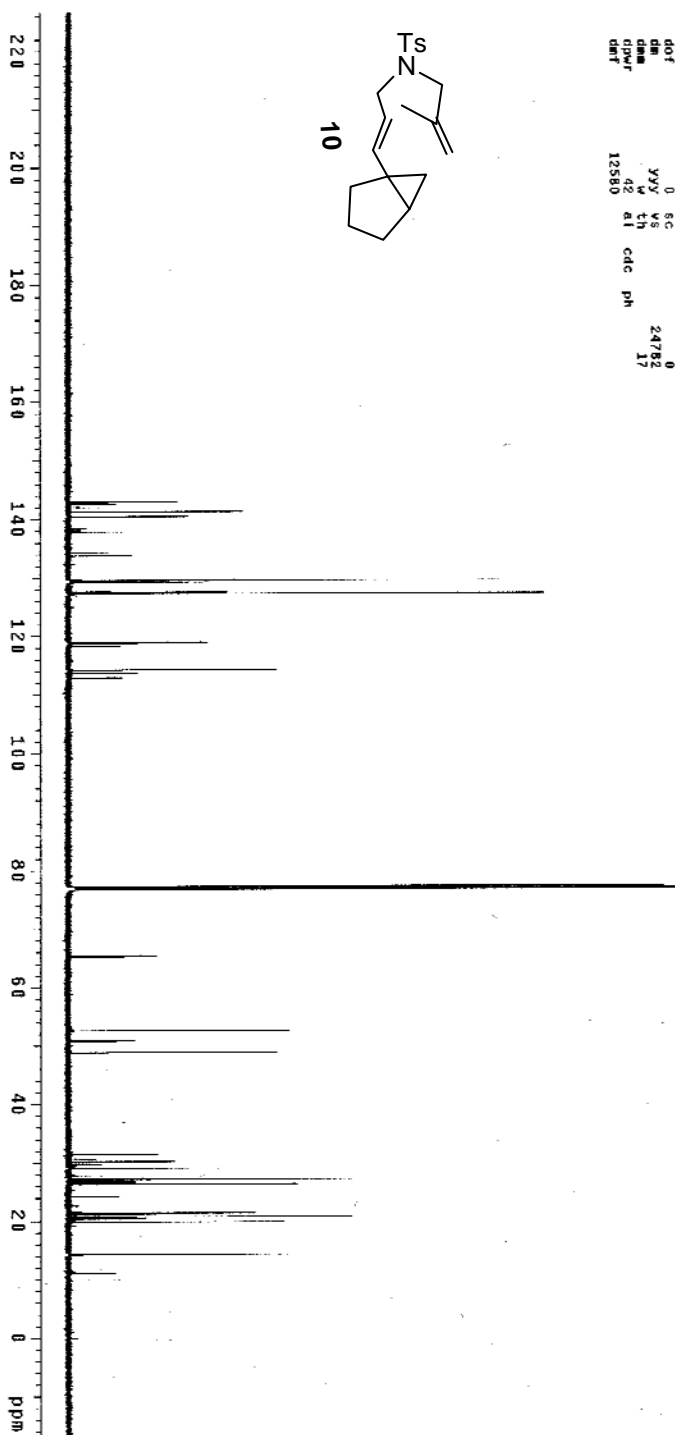
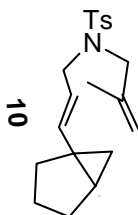
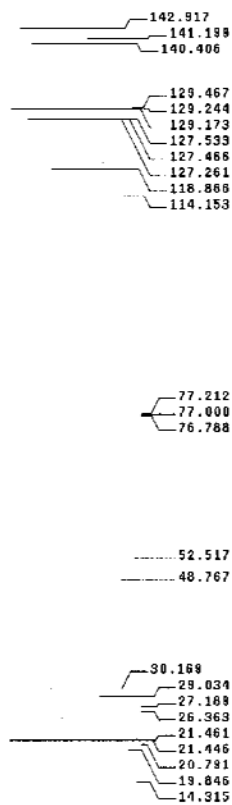
File: 040512-Moesy-1
 Pulse Sequence: MUESY
 Solvent: cdcl3
 Temp: 25.0 C / 298.1 K
 Omega: 125.761 MHz
 File: 040512-Moesy-1
 VMRS-600 "RDCMR"
 Relax: delay 1.000 sec
 Mixing: 0.400 sec
 Acq. time 0.224 sec
 Width: 4582.0 Hz
 2D Width: 4582.0 Hz
 32 F2 partitions
 32 F1 partitions
 Offset: 11.5531774906 MHz
 DATA PROCESSING
 Gauss apodization 0.104 sec
 F1 DATA PROCESSING
 Gauss apodization 0.080 sec
 F1 size 4096 x 4096
 Total time 19 hr, 6 min, 52 sec



10

CC1=C(C2CC3C(C1)C=C(C2)C3)N(C4=CC=C(C=C4)C(=O)C5=CC=CC=C5)C=C1

HF-07
 8KPB Carbon
 SAMPLE
 date Dec 11 2007 temp 25.0
 solvent cdc13 gain 30
 file /export/home/~ not used
 zly/vnmr/sy/data/h- hst 0.008
 uang/eng/HF-07-240- pw20 10.000
 241.153 f1d alfa
 ACQUISITION
 SW 36764.7 f1
 at 1.300 in n
 np 85824 dp y
 pb 17000 hs PROCESSING nm
 bs 64 lb not used
 us 1.000 f0
 nt 2000 f1 DISPLAY
 ct 1344
 TRANSMITTER
 tn C13 sp -2582.0
 sftq 150.829 rft 36764.1
 tot 2339.1 rfp 14195.2
 lprf 155.55 tp 11612.6
 pw 3.800 tp -238.0
 DECOUPLER H1 WC PLOT 250
 dn 0 sc 0
 dof 0 sc 0
 dm yyy vs 24782
 dnm w th 17
 dipw 42 at cdc ph
 dnt 12580



040613

exp5 Proton

SAMPLE
 date apr 6 2017 temp not used
 solvent cdcl3 gain 8
 file /export/home/~ not used
 y1/vnmr/sy/data/hu- het 8.088
 angfreq/040613-H-1- pw90 11.500
 id a17a 6.600

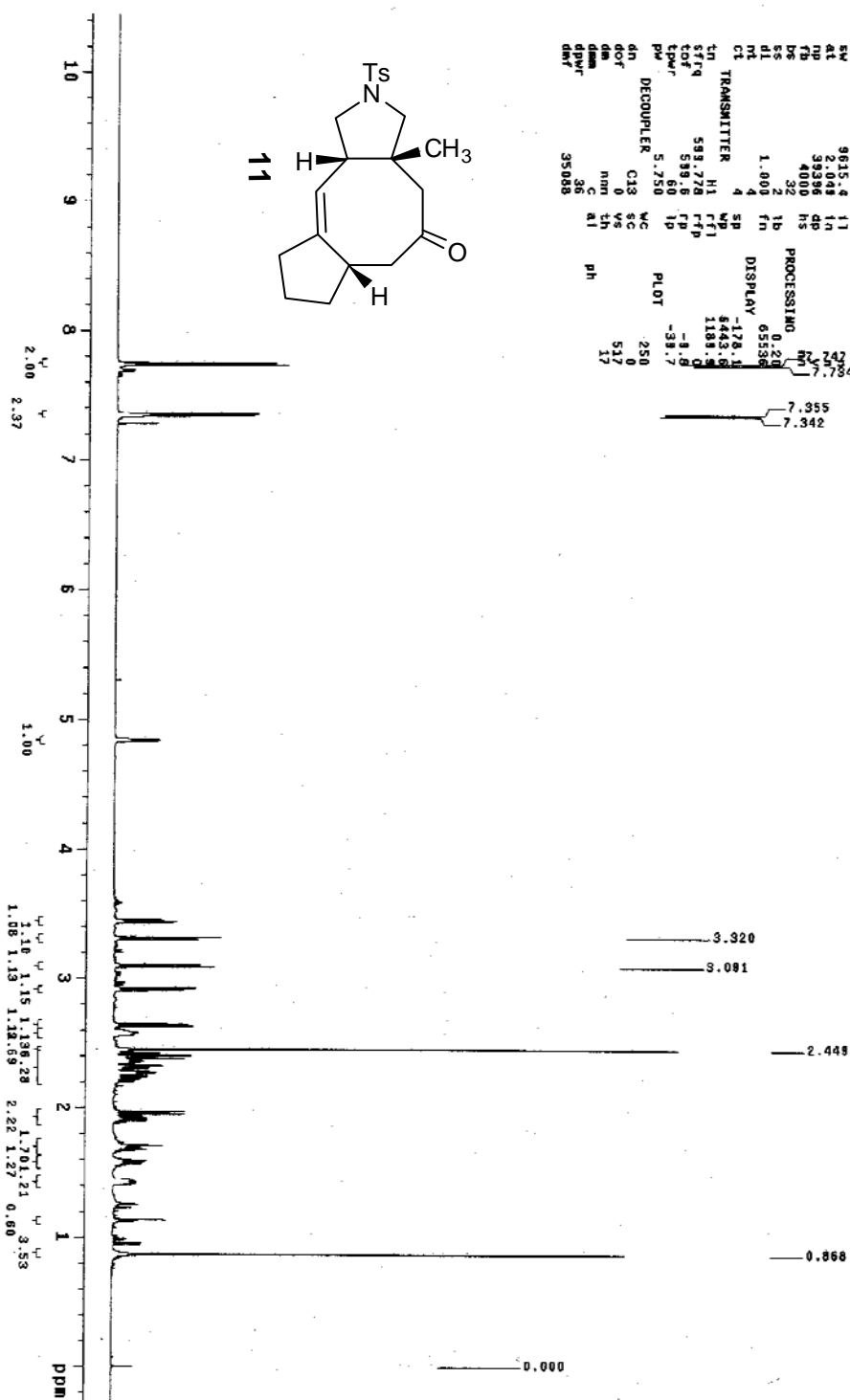
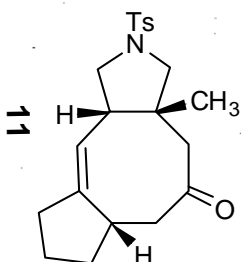
ACQUISITION
 sw 3615.4 11
 at 2.048 10
 np 39396 dp
 fb 4000 hs
 bs 32 1b
 ss 1.003 fm
 ul 4
 ct 4

PROCESSING
 0.20
 65536
 7.355
 7.342

TRANSMITTER
 h1 ffl
 sfreq 589.728
 cdf 589.8 ffp
 tpr 5.750 1p
 pw DECOUPLER C13
 dn 0 vs
 dof 0 th
 dm nm
 dpr 35088

DISPLAY
 -128.1
 6443.6
 1189.4
 -8.9
 -38.7
 250
 517
 17

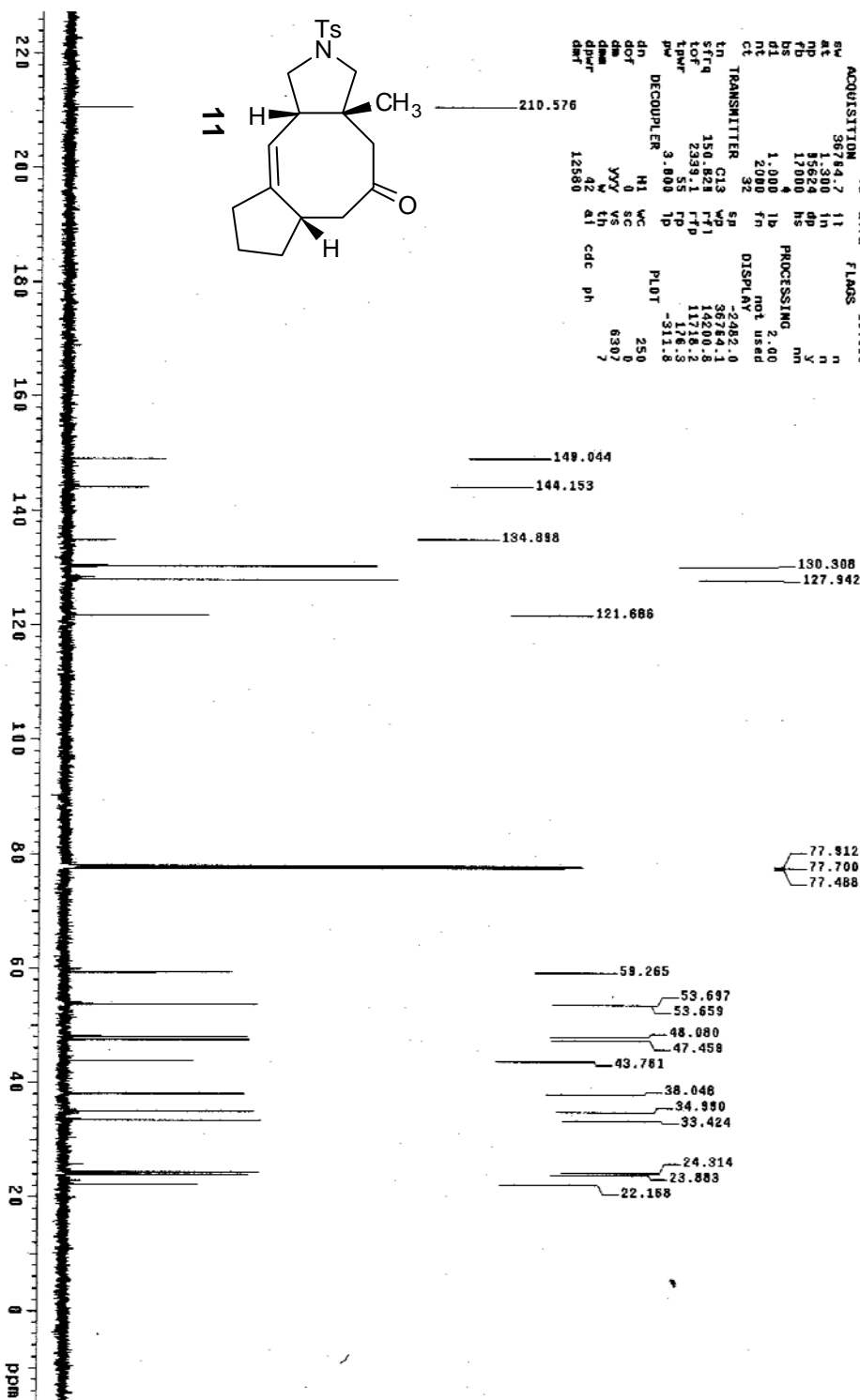
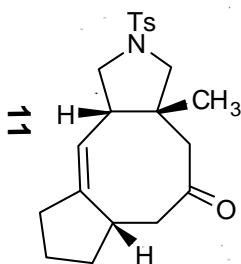
PLOT



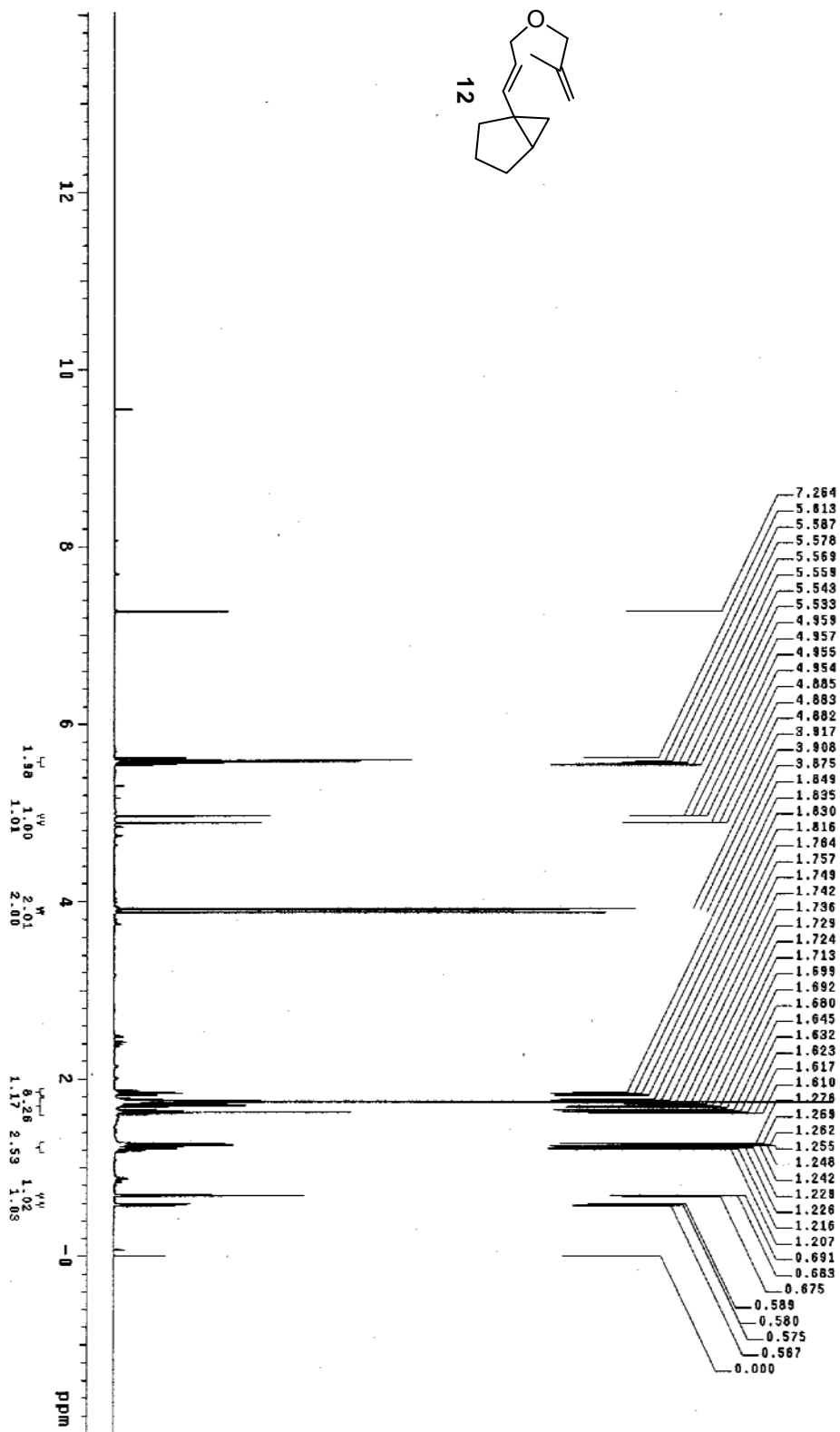
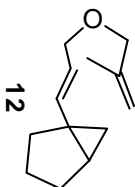
040613

exp5 Carbon

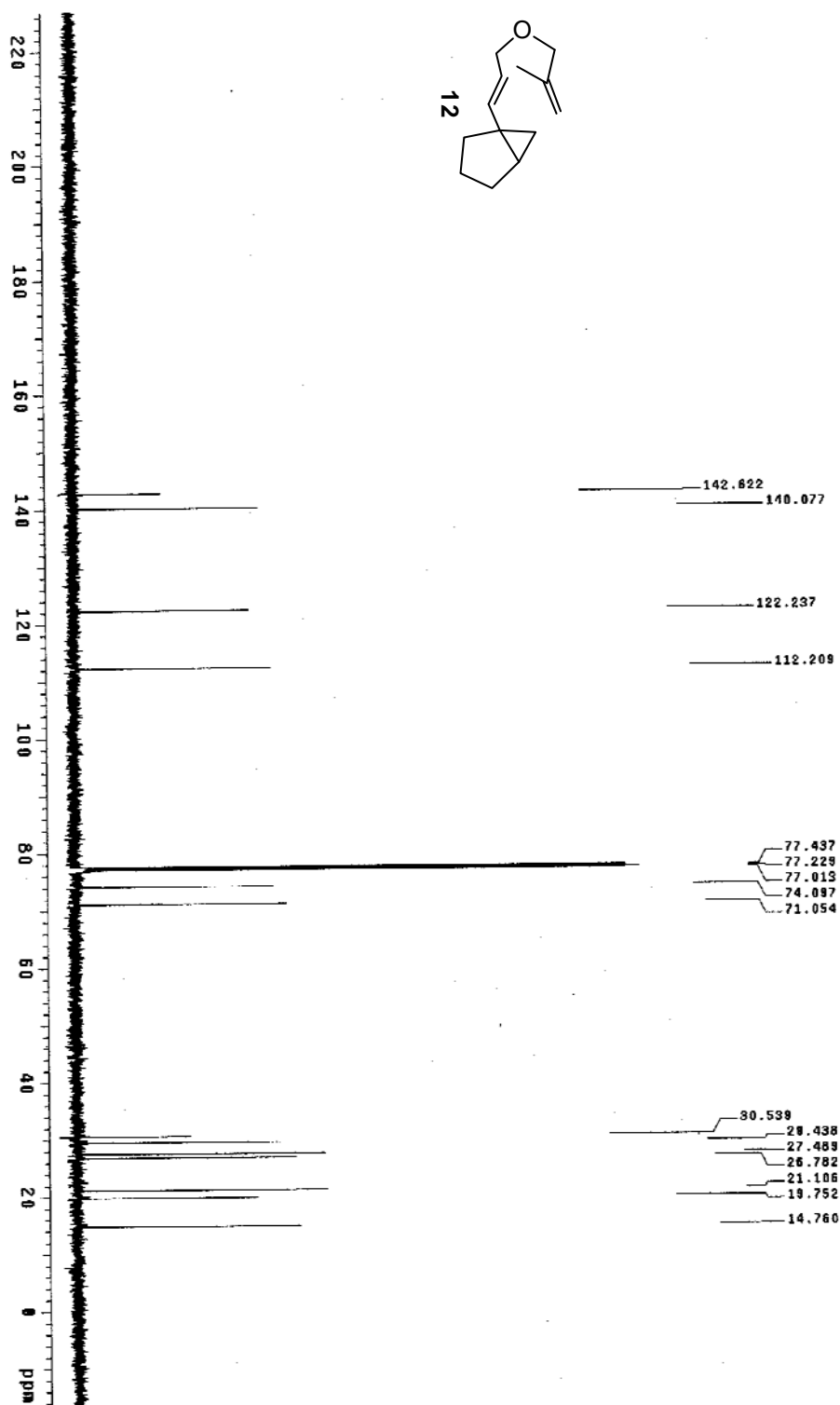
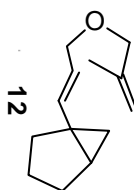
SAMPLE 6 2007 temp not used
 date Apr cdc13 gain 40
 solvent cdc13 not used
 file /export/home/~ not used
 y1/vm1/sv/data/hu- hst 5.006
 anbr/emg/040613-C-1-7- p420 7.600
 18 atfa 18.000
 ACQUISITION 18 atfa 18.000
 sw 36764.7 11 n
 at 1.300 1n n
 np 85624 dp y
 fb 17000 hs m
 bs 4 1b PROCESSING nm
 st 1.000 2.00
 nt 2000 fn not used
 ct 32 DISPLAY
 TRANSMITTER 32 sp -2482.0
 tn C13 wp 36764.1
 sftq 150.824 f71 14200.8
 tof 2339.1 f7p 11718.2
 tpwr 3.808 1p 176.3
 pw DECOUPLER H1 WC PLDT -311.8
 dn 0 250
 dof 0 sc d
 da yyy vs 6307
 dm w th
 dpar 42 at cdc ph
 det 12560



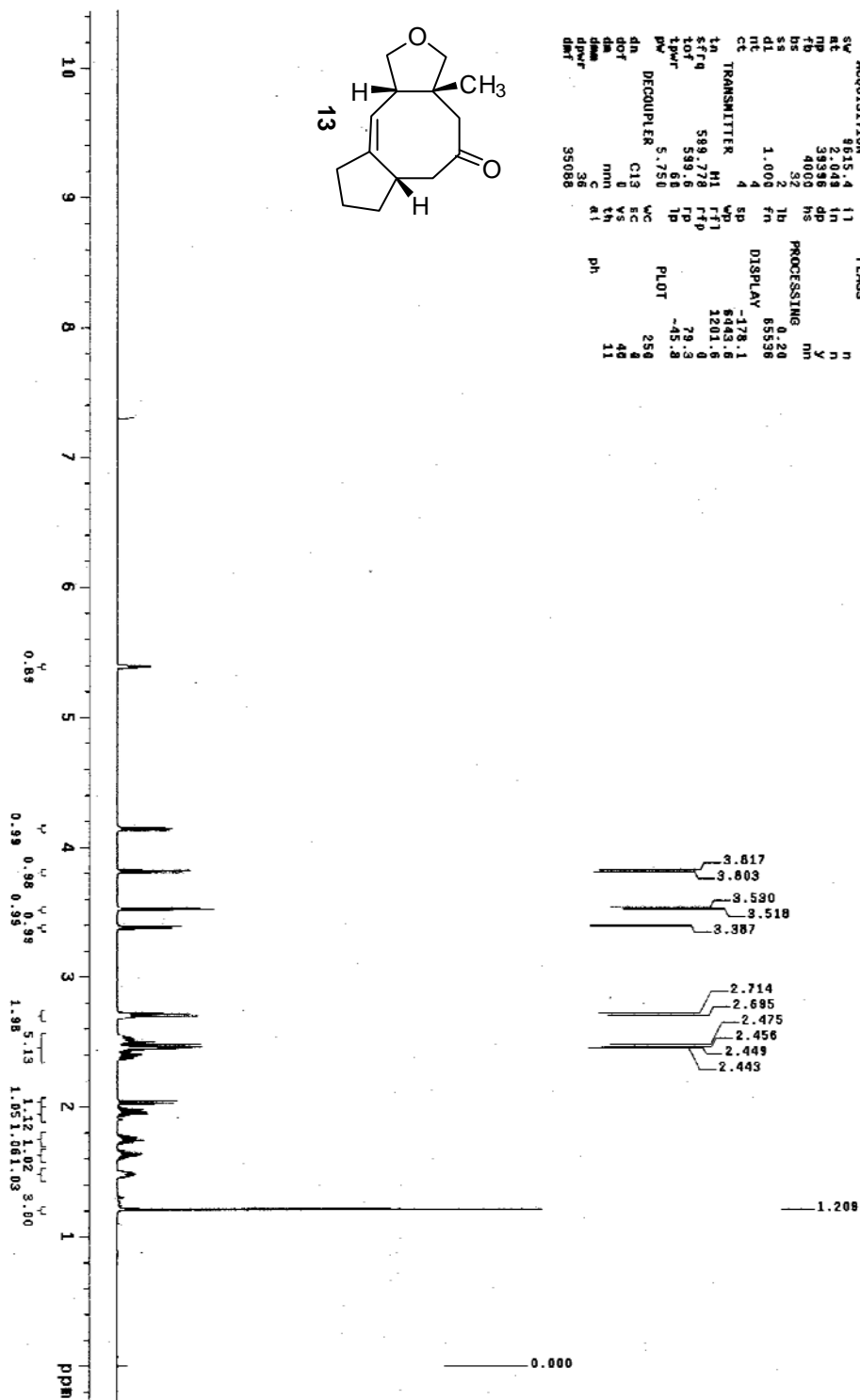
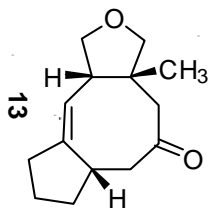
HF-11
 File: HF-11-20076891-H
 Pulse Sequence: szpu1



HF-11
File: HF-11-20070801-C
Pulse Sequence: zgpg1



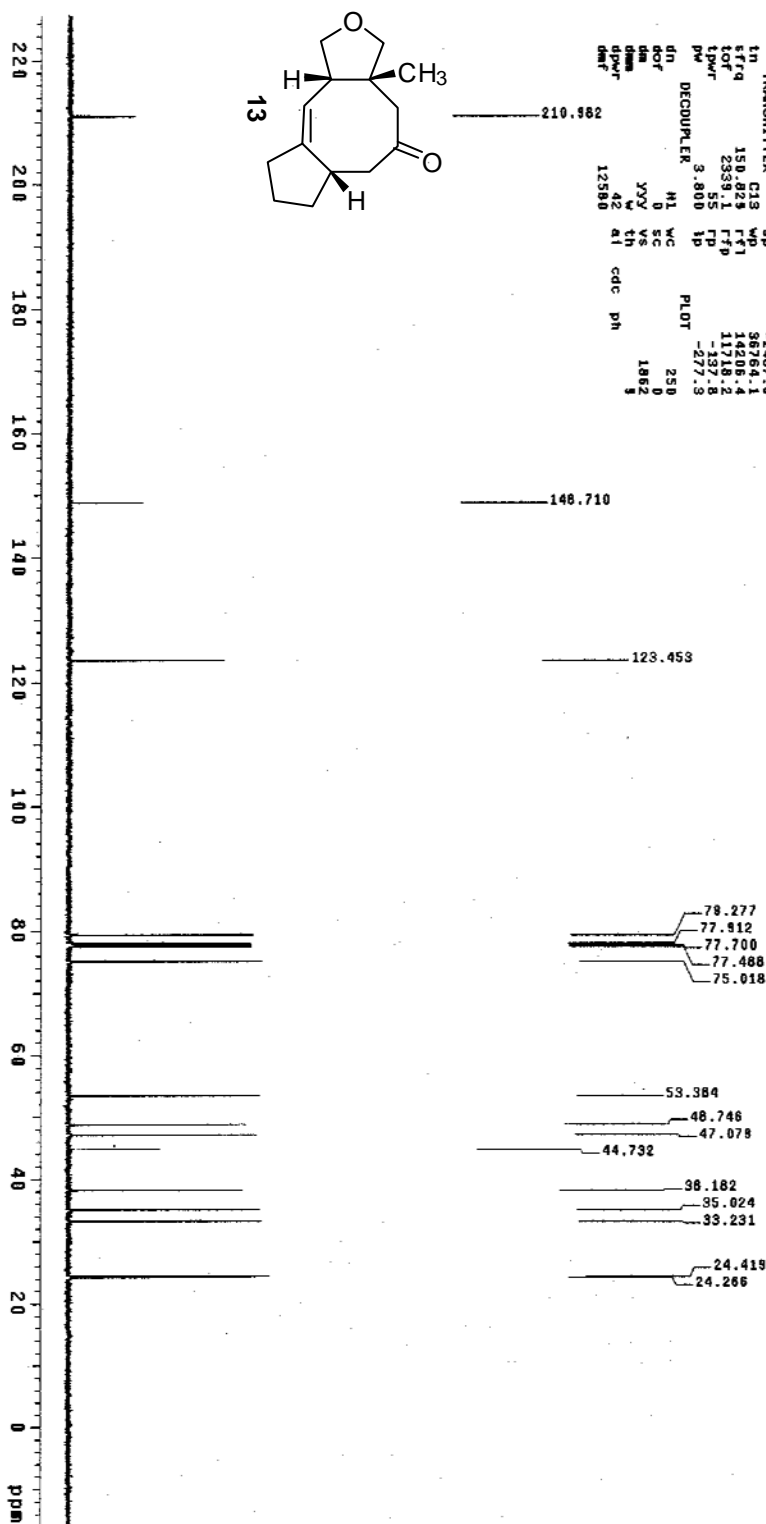
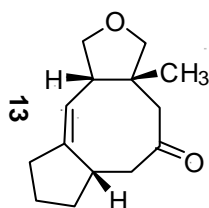
040603
 emp5 Proton
 SAMPLE SPECIAL
 date Apr 8 2007 temp not used
 solvent cdc13 gain 20
 file /export/home/~ not used
 x1/vnmr-sys/data/hu- hst 0.008
 ang/eng/040603-4.1- pw20 11.500
 id at18a 8.600
 ACQUISITION
 sw 8615.4 11
 at 2.043 1n
 np 38386 dp
 fb 4000 hs
 bs 32 7b
 ss 1.000 85358
 dl 4
 ct 4 5p
 TRANSMITTER
 tn 4 5p
 \$frq 589.778 rfp
 totf 589.6 rfp
 tpwr 68 7p
 pw 5.750 250
 DECOUPLER C13 250
 dn 1000 40
 dot 0 vs
 dm mn th
 dprv c 40
 dmf 36 11
 35088
 PROCESSING
 n
 y
 nm
 0.20
 85358
 DISPLAY
 -178.1
 643.6
 1201.6
 79.3
 -45.8
 PLOT
 250
 40
 11
 ph



049609

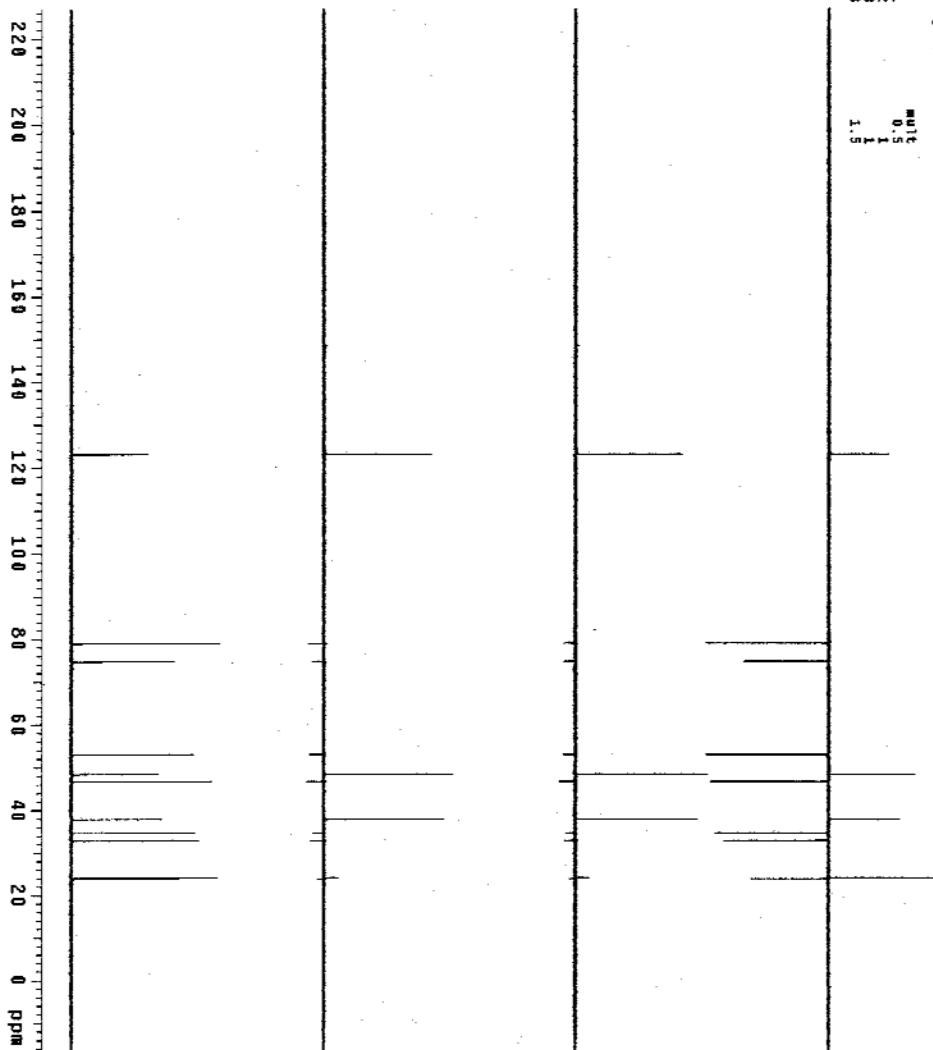
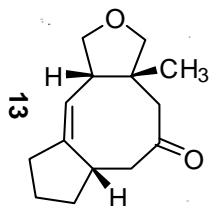
exps Carbon

SAMPLE
date Apr 6 2007 temp not used
solvent Cdc13 gain 40
file /export/home/~ not used
YI/VMR/Sys/date/Hu- net 0.008
angfreq/00000-C-13- p990 7.000
ACQUISITION id 61768 10.000
SW 36764.7 13
at 1.300 in n
np 85624 dp y
fb 17000 hz
ds 1.000 1b
nt 2000 fm not used
ct 28
TRANSMITTER C13 SP -2487.6
in 150.823 wd 36764.1
efrq 2359.1 f1 14206.4
tort 5.5 f2 11219.2
bpr 3.800 1p -277.6
pw DECOUPLER N1 WC PLOT 250
dn dnf 0 SC 1862
dof dnf 0 SC 1862
dm dnf 0 SC 1862
dpr dnf 0 SC 1862
dmf dnf 0 SC 1862



040809
 exp5 Dept
 SAMPLE
 date Apr 8 2007 DEPT 140.0
 solvent cdc13 mult arrayed
 sample
 ACQUISITION
 sw 36764.7 temp not used
 at 1.000 gain 26
 np 73530 spin PROCESSING 20
 ds 44 1b 1.00
 si 32 32 fn SPECTRUM 65536
 nt 1.000 32 mp 36763.6
 ct 32 sp -2545.8
 tn TRANSMITTER C13 1p 37.6
 tof 2339.1 ai cdc ph
 tpwr 7.600 rf1 REFERENCE 2546.9
 pw DECOUPLER H1 rfp PLOT 0
 dn 0 wc 180
 dot 42 sc 0
 dpwr my vs 2800
 dam ccw hzmm 204.24
 dmf 12580 th 8
 ppiw1 57
 pp 15.500

ACQUISITION ARRAYS
 array mult
 arrayd/m mult
 1 1
 2 0.5
 3 1
 4 1.5



040608

F11a: gc05y

Pulse Sequence: gc05y

Solvent: cdc13

Temp: 25.8 C / 298.1 K

Operator: Y "RICDMR"

VMRS-800 "RICDMR"

Relax: delay 1.301 sec

Acq: time 0.163 sec

Width: 3140.7 Hz

2D: 3140.7 Hz

4 repetitions

128 increments

Observed: 599.7744908 MHz

DATA PROCESSING

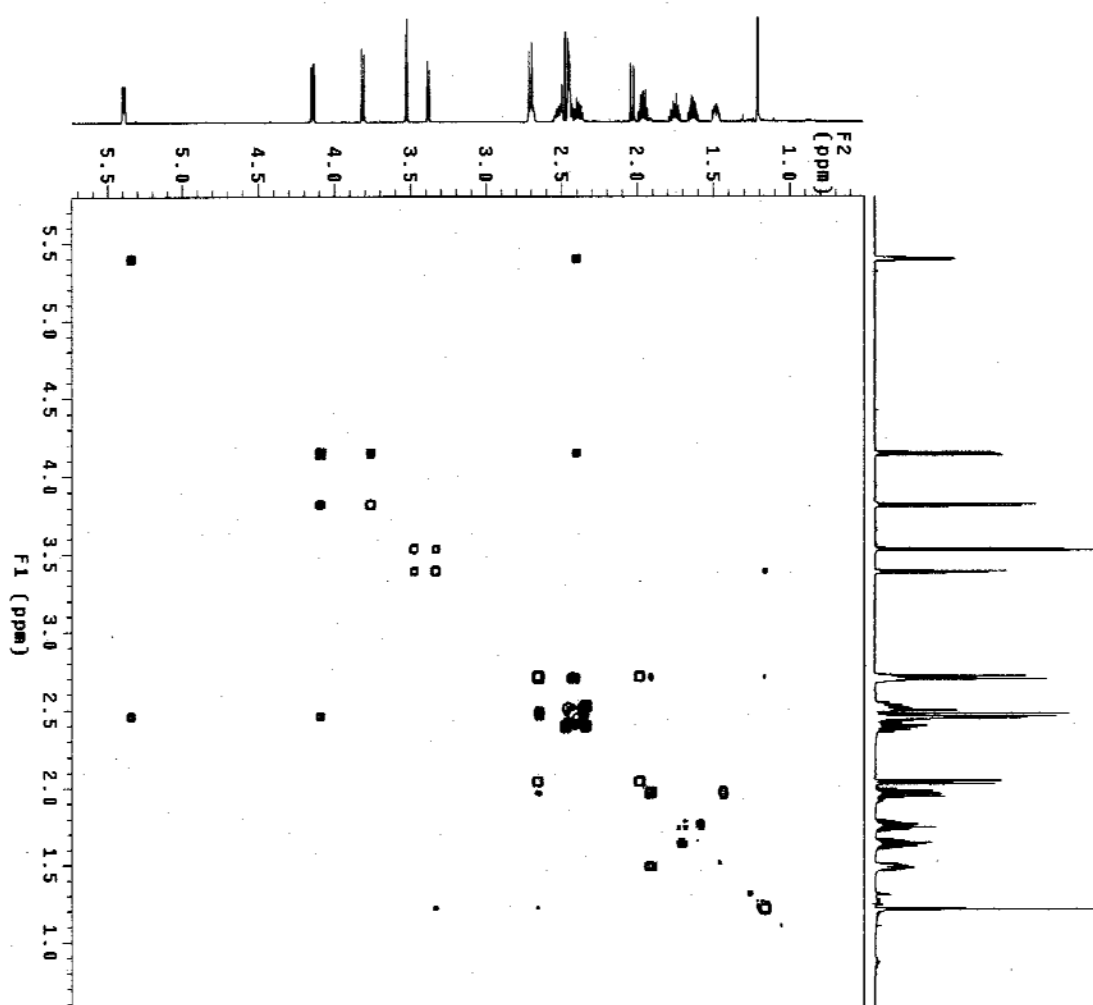
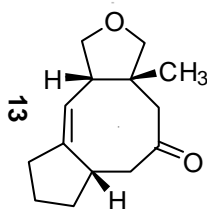
SI: 1024

F1: 0.082 sec

F2: 0.021 sec

FT size: 1024 x 1024

Total time: 13 min, 18 sec



840603

File: 040603-hsac-0410

Pulse Sequence: ghsqc

Solvent: cdcl3

Ambient temperature

Operator: Y1

File: 040603-hsac-0410

VMRS-800 "RICHMR"

Relax. delay 1.301 sec

Acq. time 0.191 sec

Width 3239.5 Hz

2D Width 36764.7 Hz

Repetitions

512

Observed

159.7744808 MHz

Decouple C13, 150.8282151 MHz

Power 36 dB

on during acquisition

off during delay

W40 AutoX modulated

DATA PROCESSING

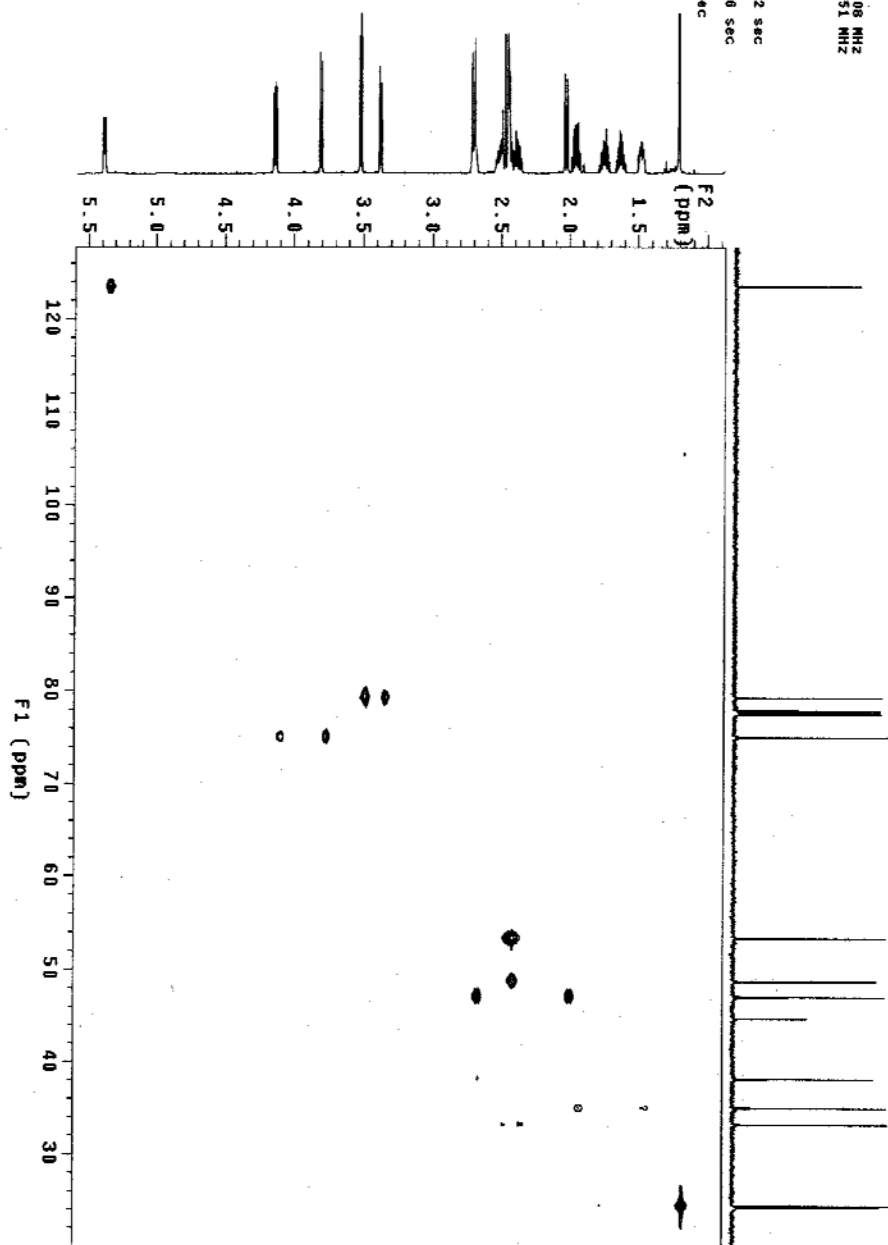
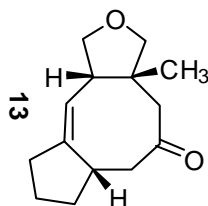
Phase apodization 0.492 sec

FT size 2048 x 4096

Gauss apodization 0.406 sec

FT size 2048 x 4096

Total time 27 min, 27 sec



046609

File: 046609-NOESY

Pulse Sequence: NOESY

Solvent: cdcl3

Ambient temperature

Operator: YI

File: 046609-NOESY

NAME: 500 "RDCMR"

Relax. delay 1.500 sec

Acq. time 0.150 sec

Width 3255.2 Hz

2D Width 3255.2 Hz

16 repetitions

2 x 128 increments

0.000 sec/1.000 Hz

0.000 sec/1.000 Hz

0.000 sec/1.000 Hz

0.000 sec/1.000 Hz

0.000 sec/1.000 Hz

0.000 sec/1.000 Hz

0.000 sec/1.000 Hz

0.000 sec/1.000 Hz

0.000 sec/1.000 Hz

0.000 sec/1.000 Hz

0.000 sec/1.000 Hz

0.000 sec/1.000 Hz

0.000 sec/1.000 Hz

0.000 sec/1.000 Hz

0.000 sec/1.000 Hz

0.000 sec/1.000 Hz

0.000 sec/1.000 Hz

0.000 sec/1.000 Hz

0.000 sec/1.000 Hz

0.000 sec/1.000 Hz

0.000 sec/1.000 Hz

0.000 sec/1.000 Hz

0.000 sec/1.000 Hz

0.000 sec/1.000 Hz

0.000 sec/1.000 Hz

0.000 sec/1.000 Hz

0.000 sec/1.000 Hz

0.000 sec/1.000 Hz

0.000 sec/1.000 Hz

0.000 sec/1.000 Hz

0.000 sec/1.000 Hz

0.000 sec/1.000 Hz

0.000 sec/1.000 Hz

0.000 sec/1.000 Hz

0.000 sec/1.000 Hz

0.000 sec/1.000 Hz

0.000 sec/1.000 Hz

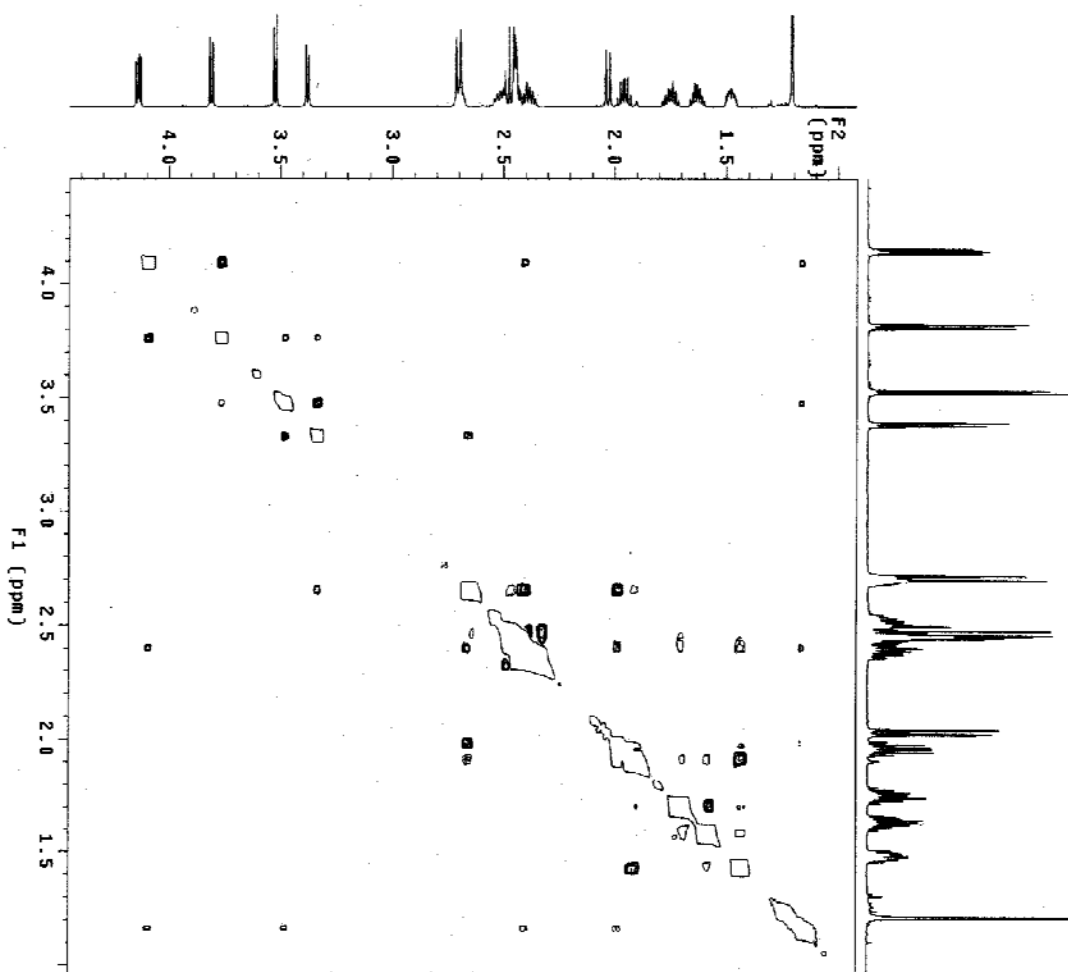
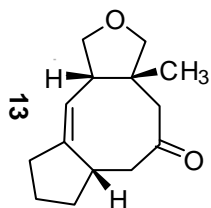
0.000 sec/1.000 Hz

0.000 sec/1.000 Hz

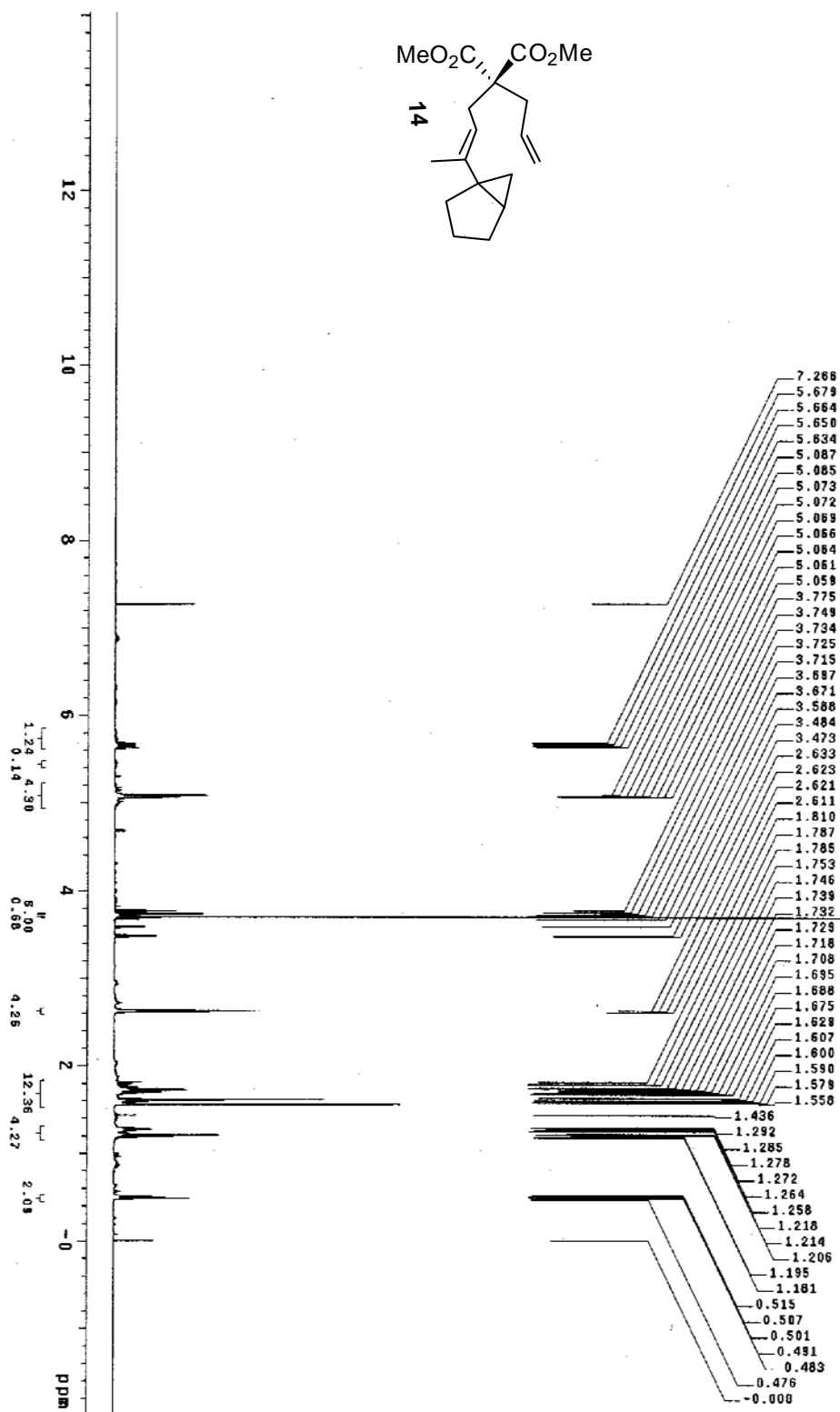
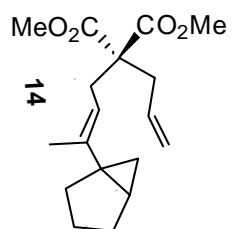
0.000 sec/1.000 Hz

0.000 sec/1.000 Hz

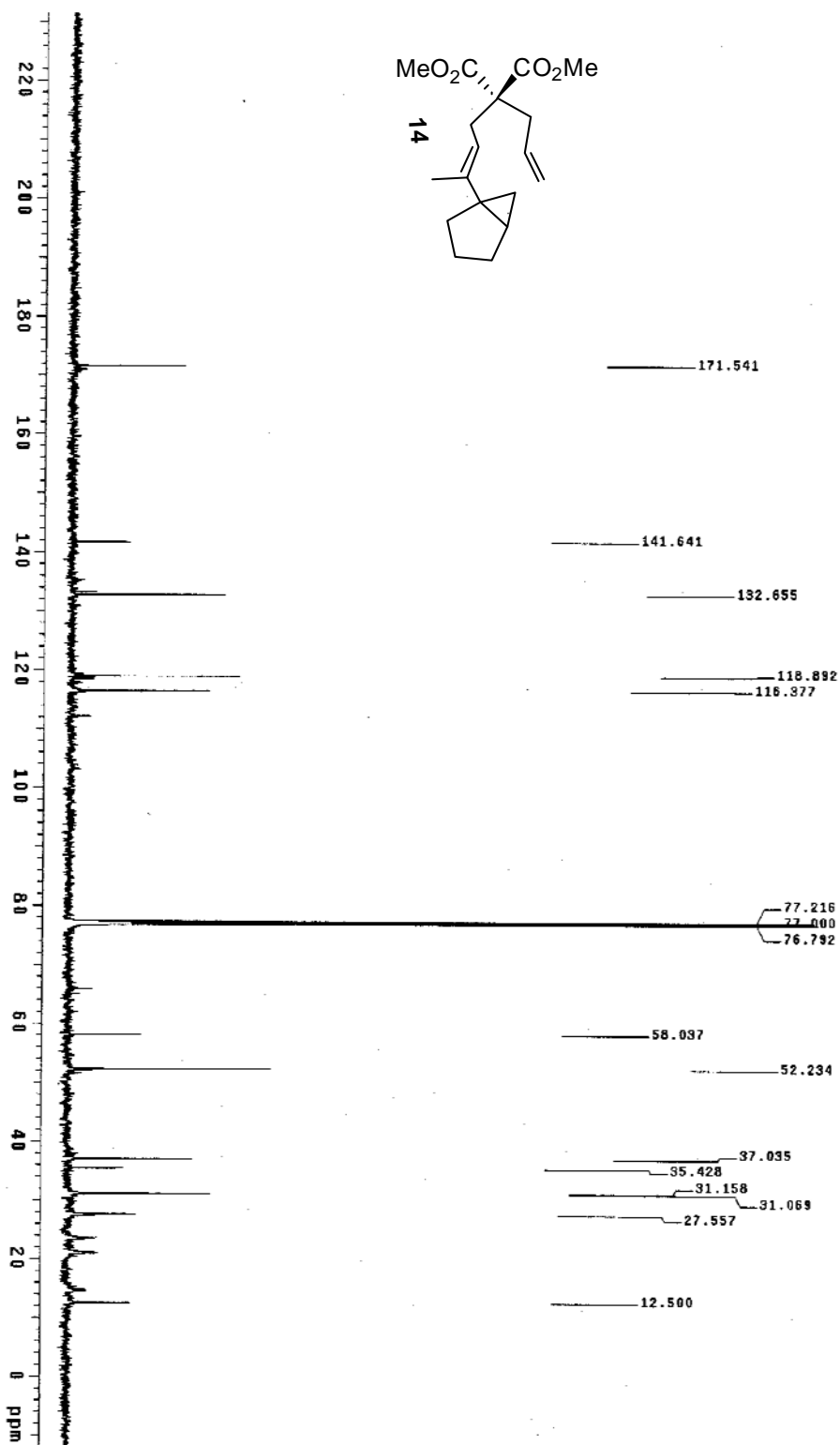
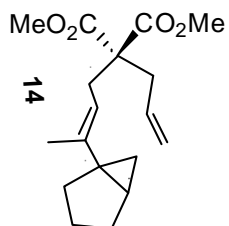
0.000 sec/1.000 Hz



MF-02
 File: MF-02-20070801-H
 Pulse Sequence: szpu1



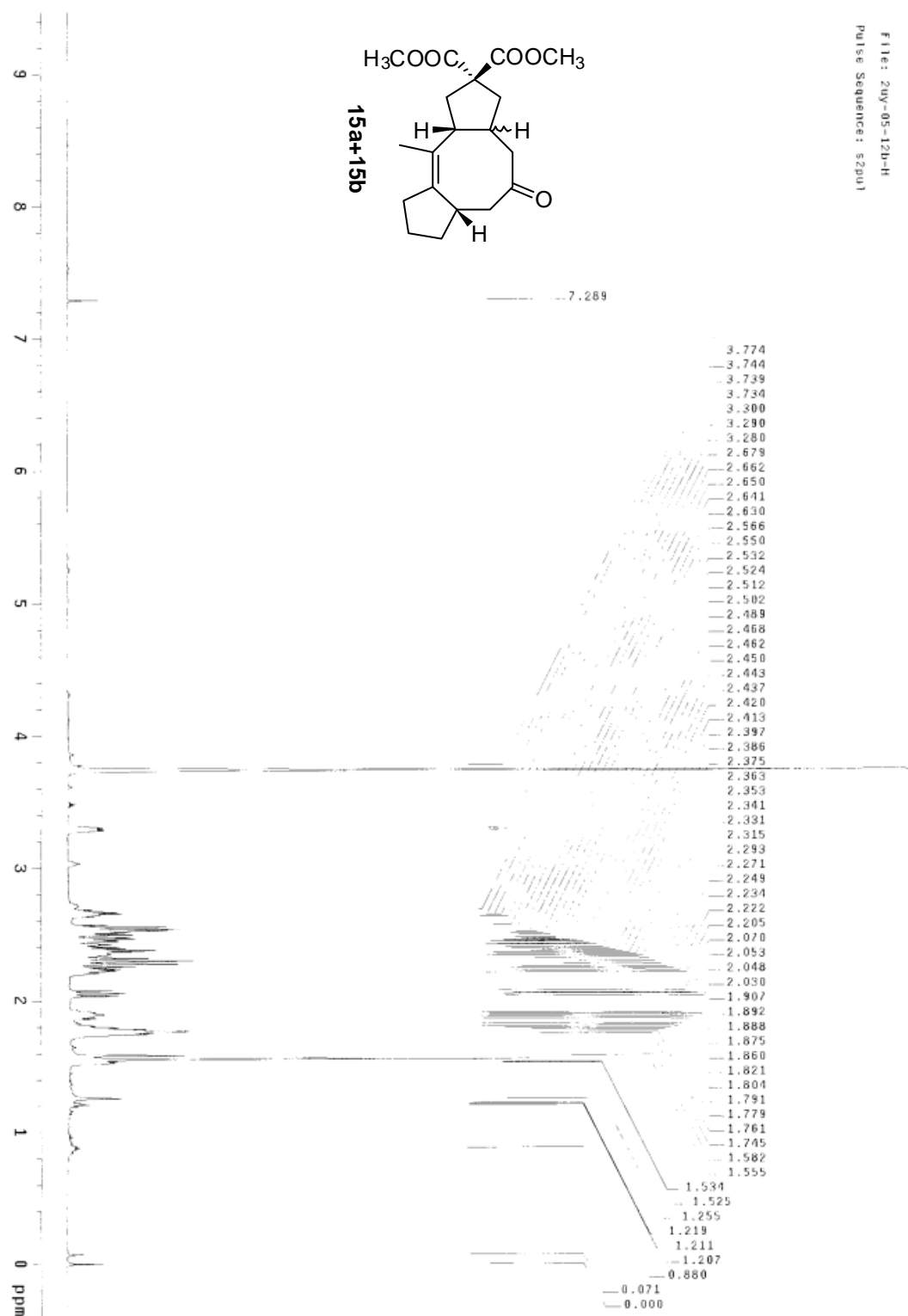
HF-02
 File: HF-02-20070801-C
 Pulse Sequence: szpu1



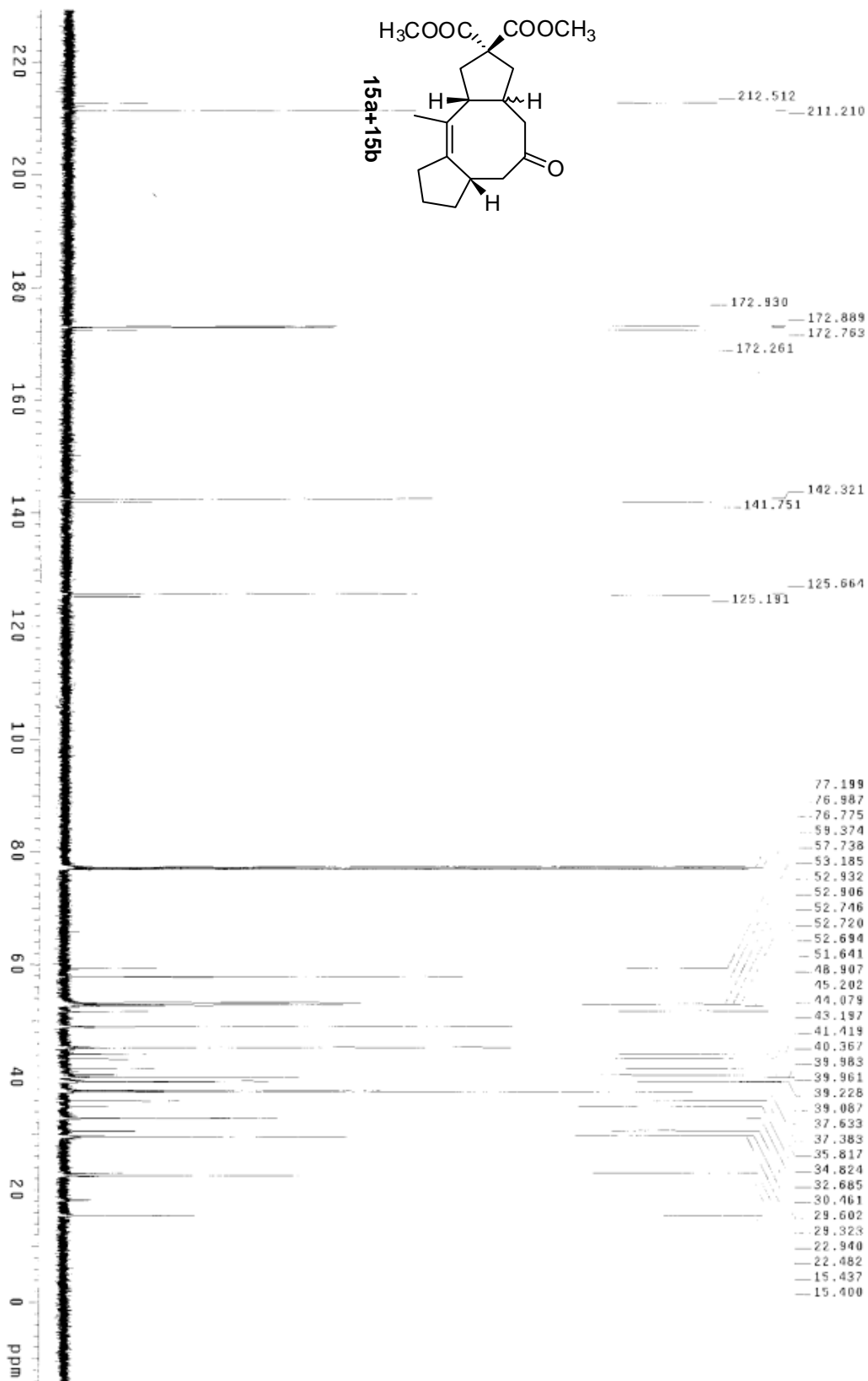
2uy-05-12b

File: 2uy-05-12b-H

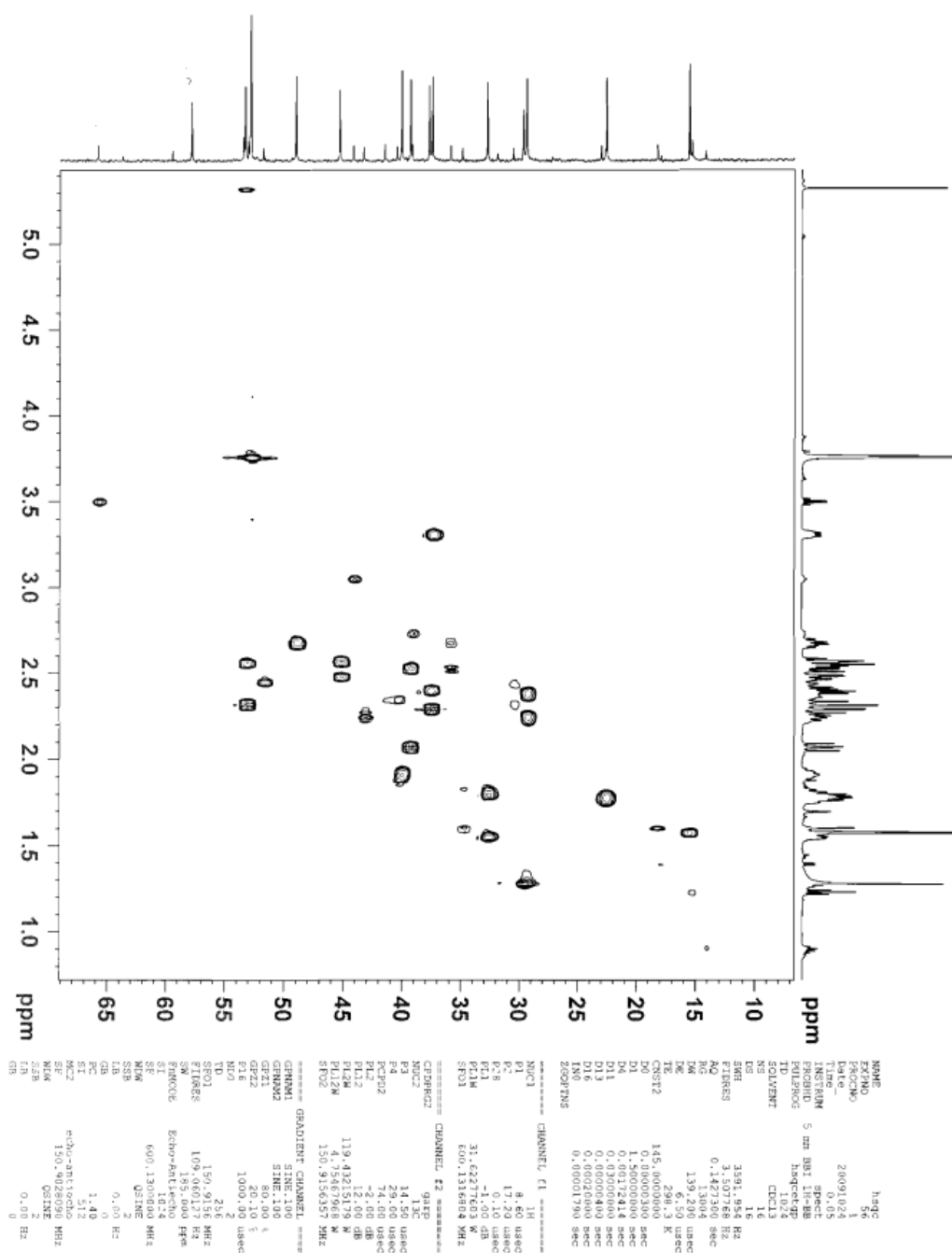
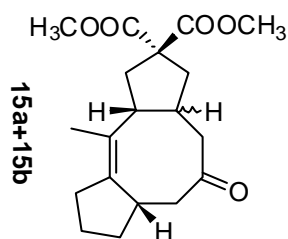
Pulse Sequence: zgpg30

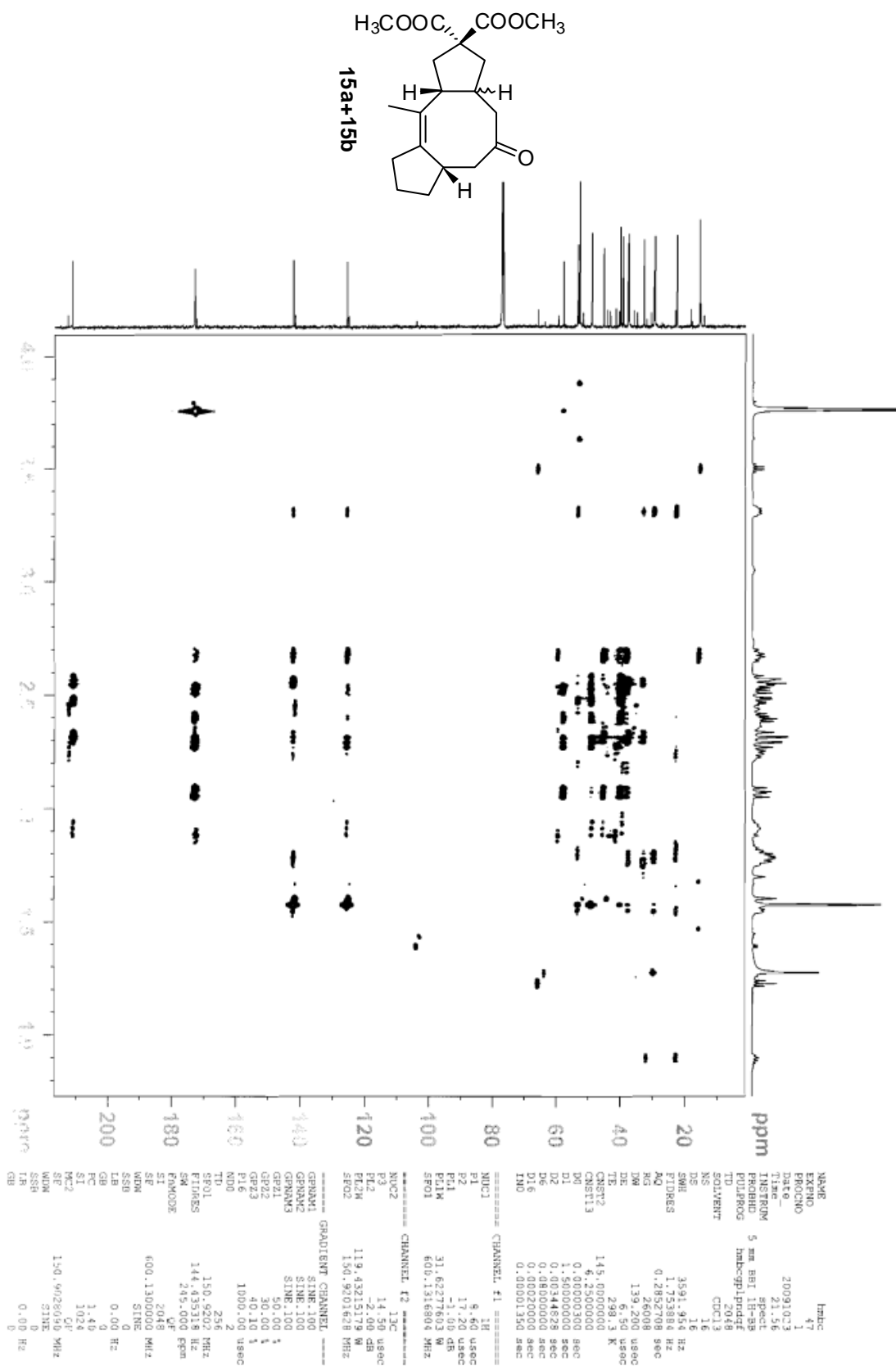


File: 2uy-05-12b-C
Pulse Sequence: szpu1

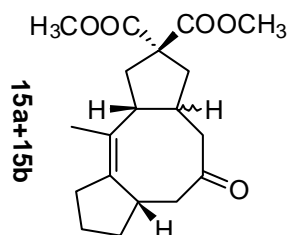
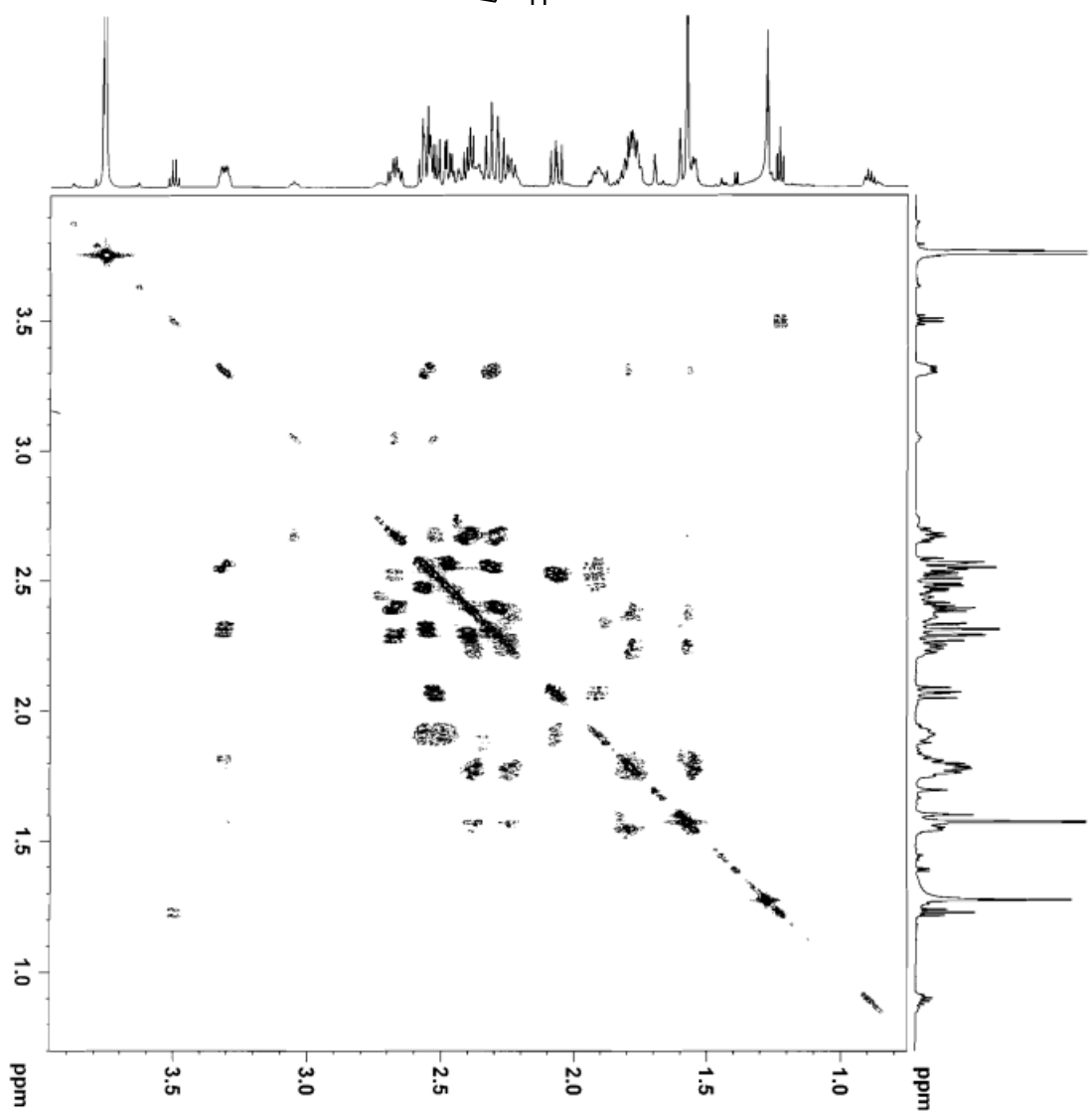


HSQC 2KY-05-12B CDCl3 298K 091023





COSY 2KY-05-12B CDCl3 298K 091023



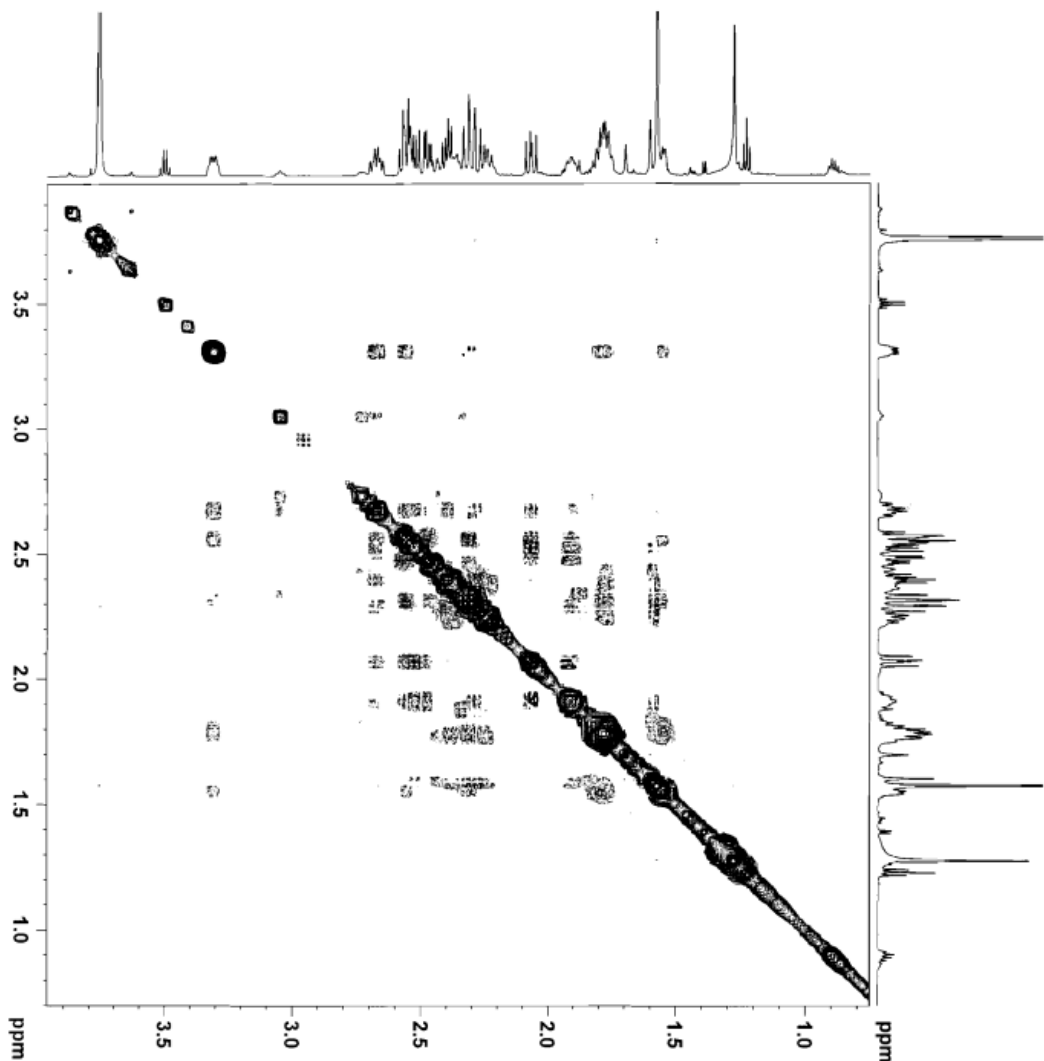
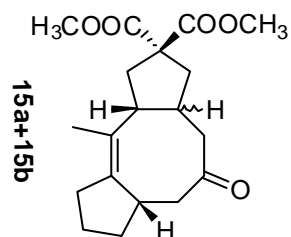
```

NAME          cosy
EXPNO         79
PROCNO        1
Date_         20091023
Time          19.50
INSTRUM       spect
PROBHD        5 mm BBI 1H-1H
PULPROG       cosyprgf
TD            2048
SOLVENT       CDCl3
NS            16
DS            16
SWH           3591.954 Hz
FIDRES        1.753884 Hz
AQ            0.2852708 sec
RG            181
DE            139.200 usec
TE            298.2 K
D0            0.00000300 sec
D1            1.50000000 sec
D13           0.00000400 sec
D16           0.00002000 sec
IN0           0.00027840 sec

===== CHANNEL f1 =====
NUC1          1H
P0            4.30 usec
PL            8.60 usec
PL1          -1.00 dB
PL1W          31.62271603 W
SFO1          600.1316804 MHz

===== GRADIENT CHANNEL =====
GPNAM1        SINE.100
GPR1          10.00 %
PI6           1000.00 usec
ND0           1
TD            256
SFO1          600.1317 MHz
FIDRES        14.031126 Hz
SW            5.985 ppm
FIRMODE       QF
SI            1024
SE            600.1300000 MHz
WDW           SINE
SSB           0
LB            0.00 Hz
GB            0
PC            1.40
SI            1024
MC2           QF
SF            600.1300000 MHz
WDW           SINE
SSB           0
LB            0.00 Hz
GB            0
  
```


NOESY 2KY-05-12B CDCl3 298K 091023



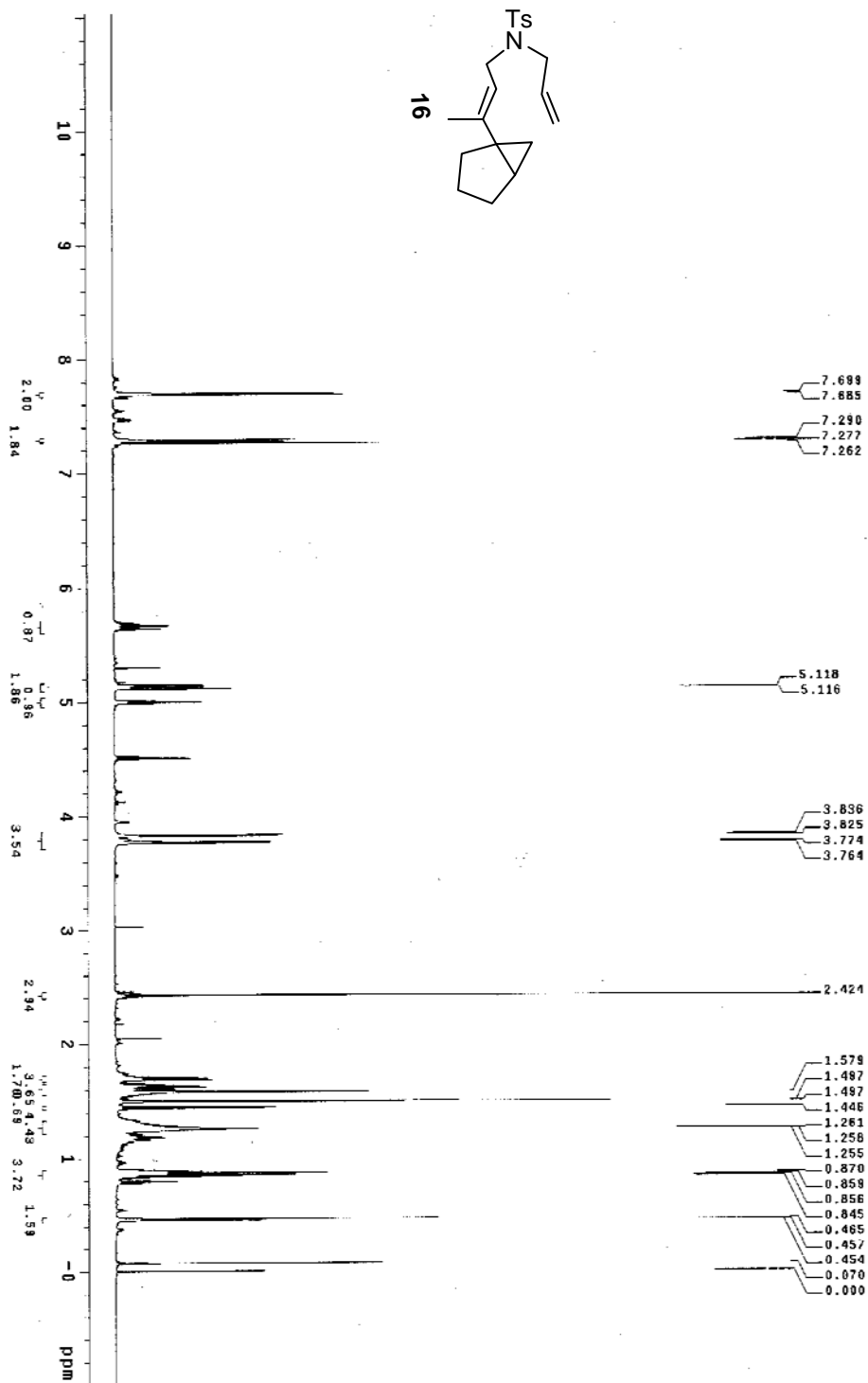
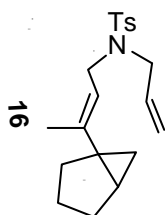
```

NAME      noesy
EXPNO     100
PROCNO    1
Date_     20091024
Time      1.59
INSTRUM   spect
PROBHD    5 mm BBI 1H-BB
PULPROG   noesygph
TD         2048
SOLVENT   CDCl3
NS         16
DS         16
SWH        3591.954 Hz
FIDRES     1.753884 Hz
AQ         0.2852708 sec
RG         45.3
DW         139.200 usec
DE         6.50 usec
TE         298.3 K
D0         0.00012825 sec
D1         1.50000000 sec
D8         0.80000001 sec
D16        0.00020000 sec
INO        0.00027840 sec

===== CHANNEL f1 =====
NUC1       1H
P1         8.60 usec
P2         17.20 usec
PL1        -1.00 dB
PL1W       31.62277603 W
SFO1       600.1316804 MHz

===== GRADIENT CHANNEL =====
GPNAM1     SINE.100
GPNAM2     SINE.100
GP21       40.00 %
GP22       -40.00 %
P16        1000.00 usec
ND0         1
TD         256
SFO1       600.1317 MHz
FIDRES     14.031126 Hz
SFO        5.985 ppm
PRMODE     States-TFPI
SI         States-TFPI
SF         600.1300000 MHz
WDW         GSIINE
SSB         2
LB         0
GB         0
  
```

HF-06
 File: HF-06-20071211-H
 Pulse Sequence: zgpg30

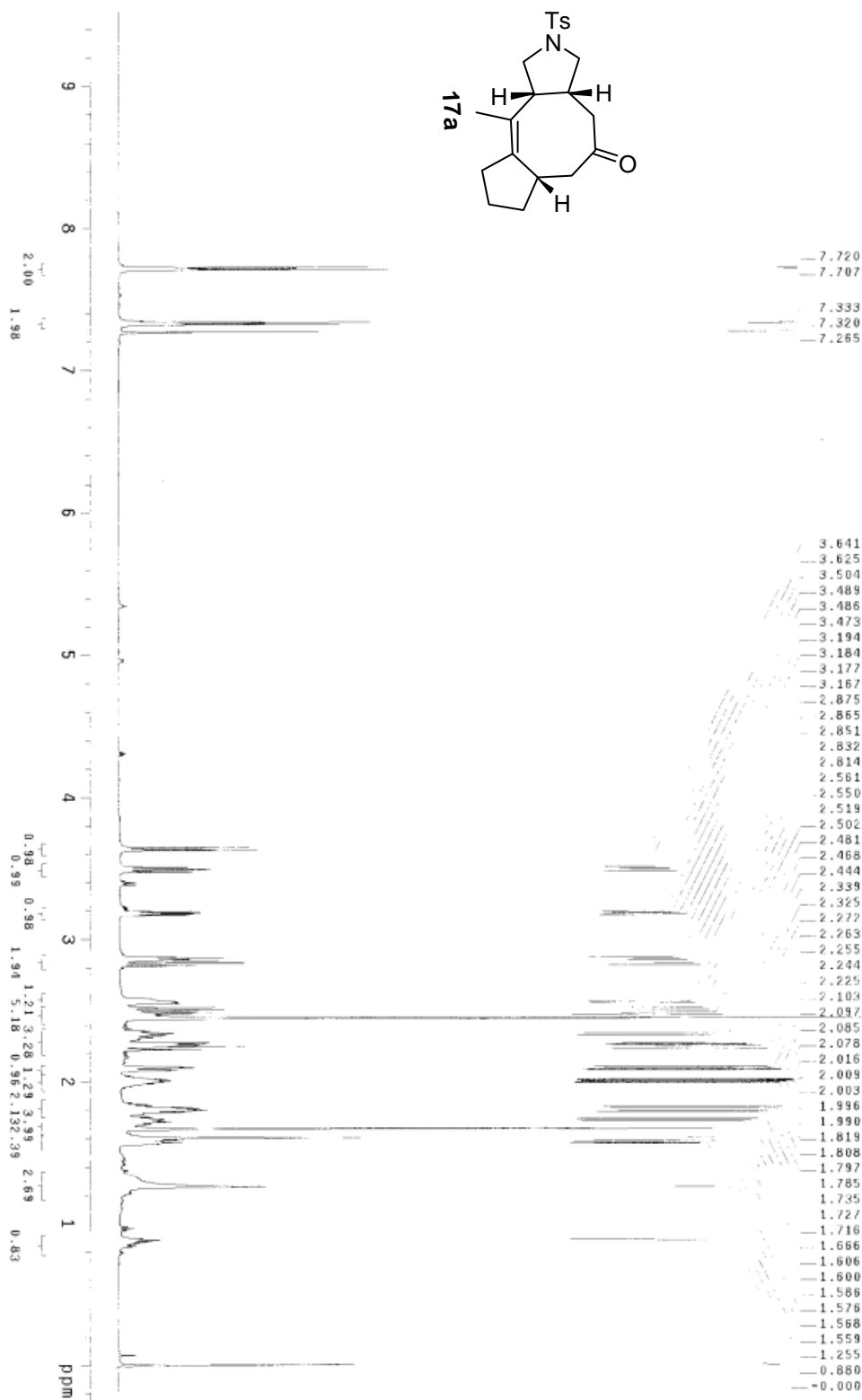
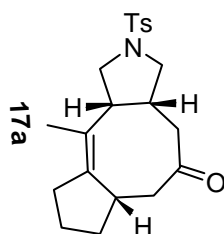


16



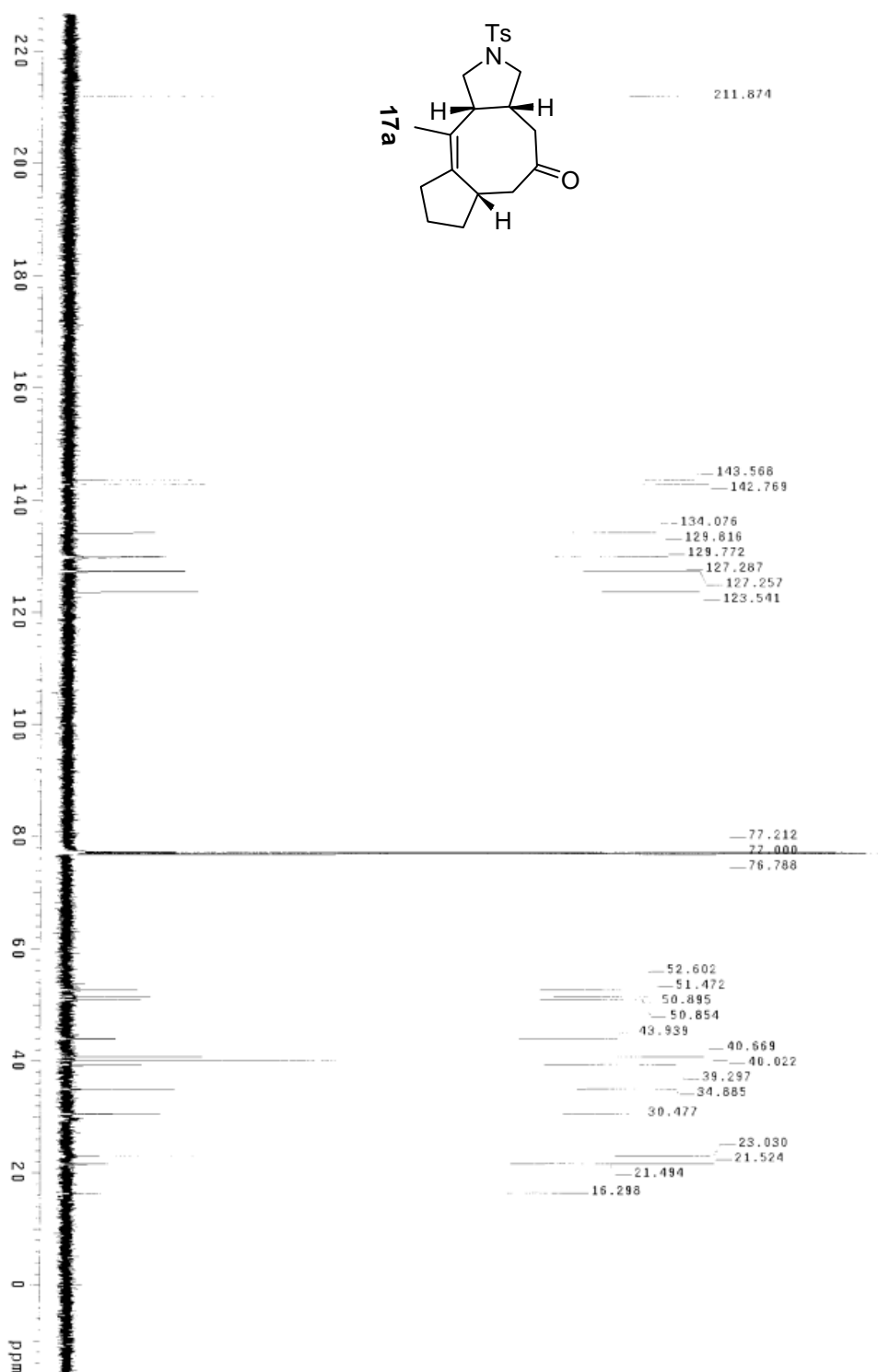
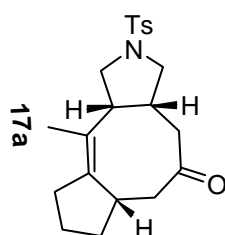
ZKY-05-118

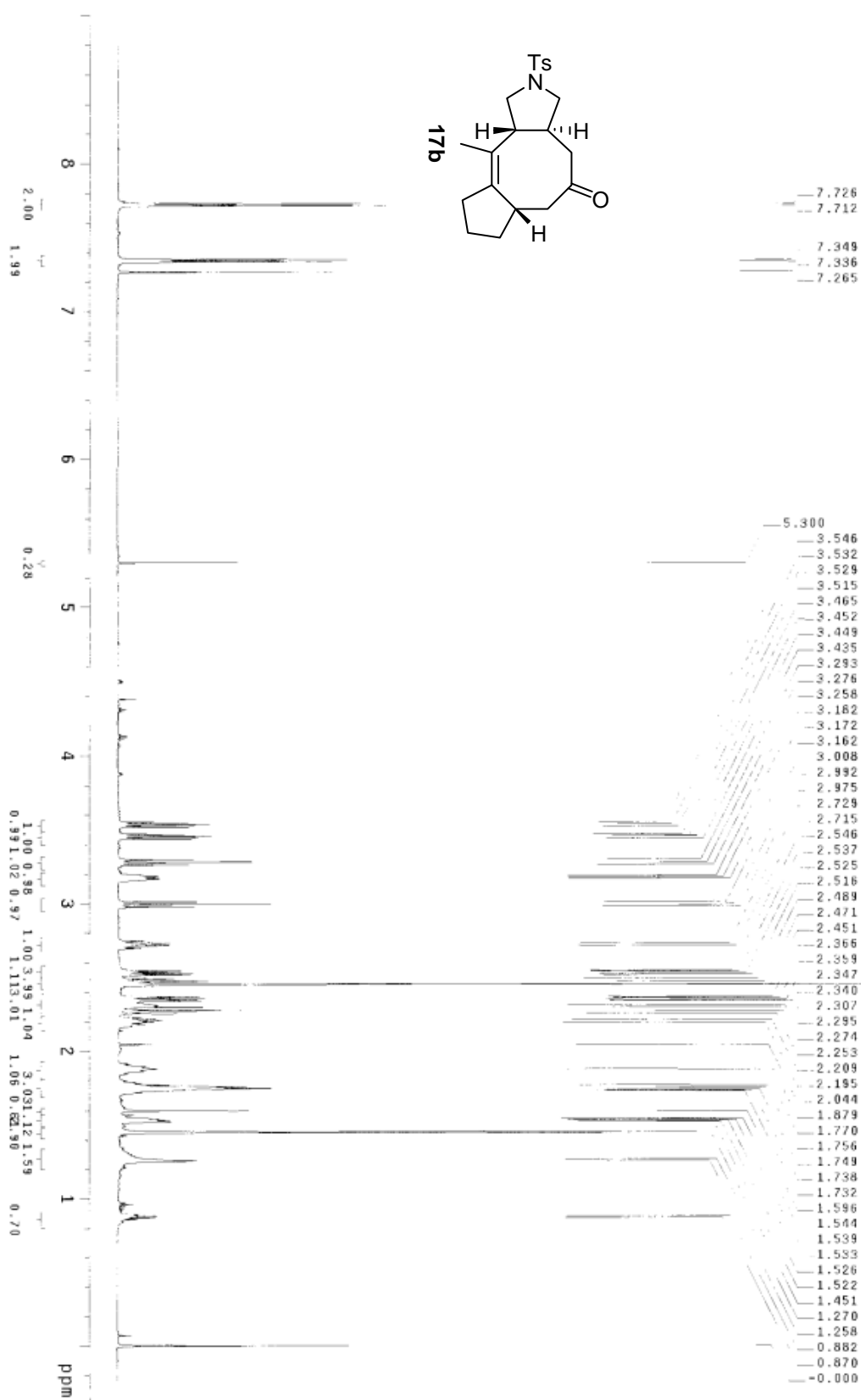
File: 20y-05-11b-H
Pulse Sequence: s2pul



2ky-05-11b

File: 2uy-05-11b-C
Pulse Sequence: szpul

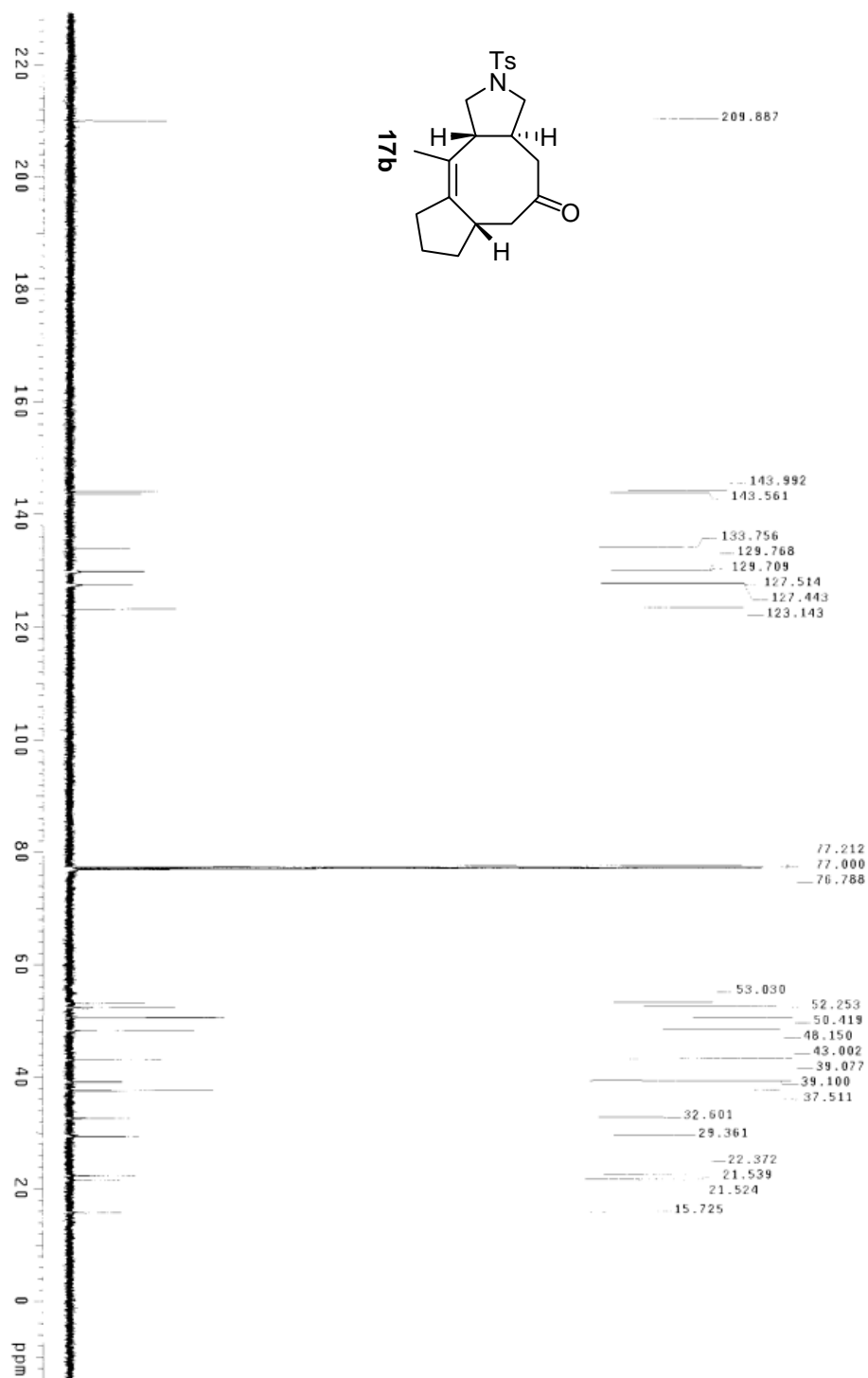




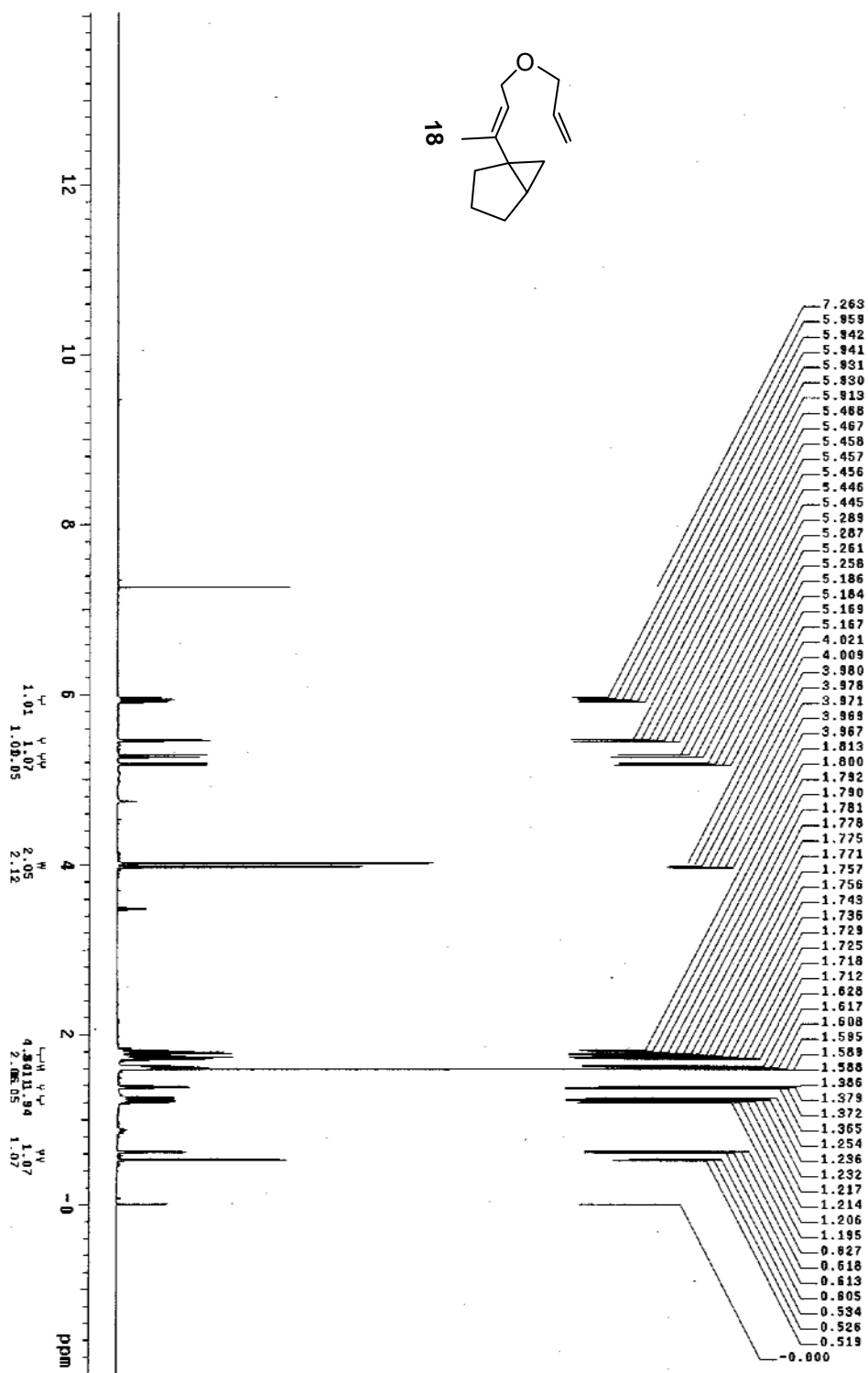
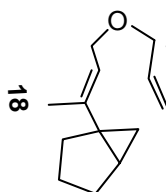
2xy-05-11c

File: 2ur-05-11c-C

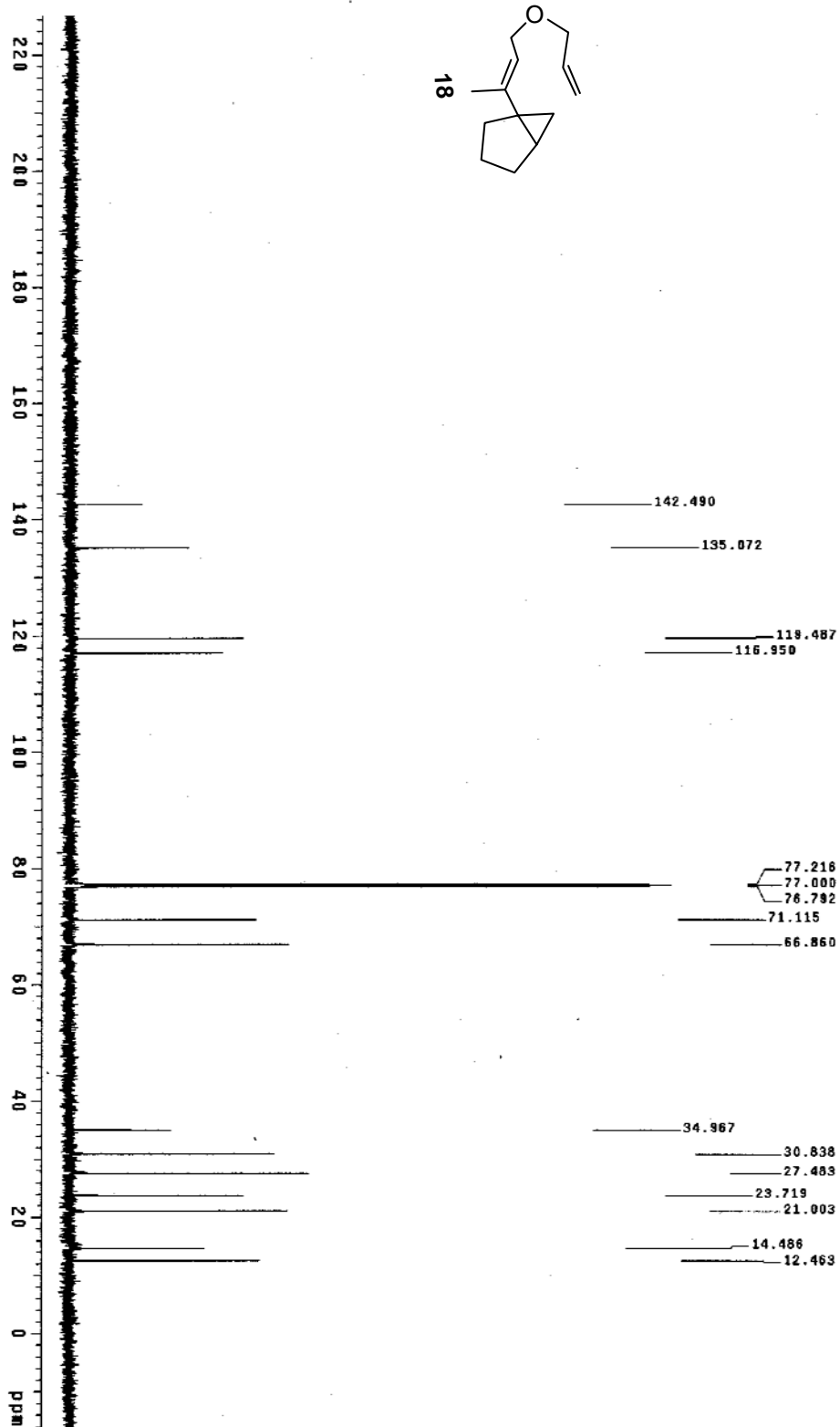
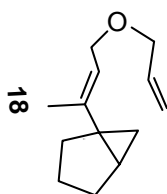
Pulse Sequence: szpu1



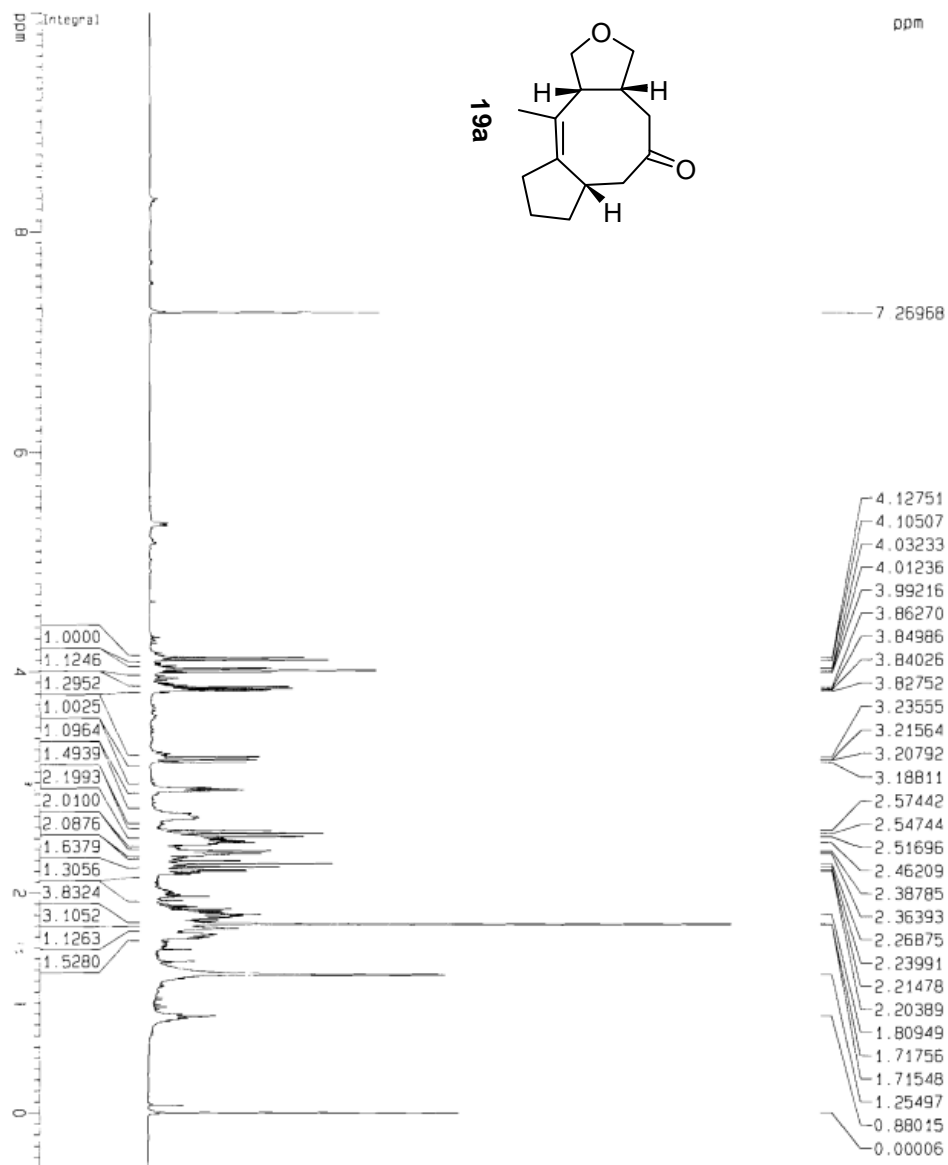
HF-10
 File: HF-18-20070801-H
 Pulse Sequence: szpul



MF-10
File: MF-10-20070801-C
Pulse Sequence: szpul



241-05-27b



Current Data Parameters
NAME n03485
EXPNO 1
PROCNO 1

F2 - Acquisition Parameters
Date_ 20091028
Time 13.56

INSTRUM ARX400
PROBHD 5 mm Multinu
PULPROG zg
TD 32768
SOLVENT CDCl3
NS 32

DS 0
SWH 6097.561 Hz
FIDRES 0.186083 Hz
AQ 2.6870260 sec

RG 360
DM 82.000 usec
DE 102.50 usec

TE 300.0 K
D1 2.00000000 sec
P1 3.00 usec

DE 102.50 usec
SF01 400.1321971 MHz
NUCLEUS 1H

F2 - Processing parameters

SI 16384
SF 400.130055 MHz
WDW no
SSB 0
LB 0.00 Hz
GB 0
PC 4.00

1D NMR plot parameters

CX 20.00 cm
F1P 10.000 ppm
F1 4001.30 Hz
F2P -0.500 ppm
F2 -200.07 Hz
PPMCM 0.52500 ppm/cm
HZCM 210.06825 Hz/cm

19a

Chemical Structure: A complex polycyclic molecule with a central eight-membered ring containing a ketone and an ether bridge. It has a cyclopentyl group and a methyl group attached to the ring system.

1D NMR Spectrum Data:

Chemical Shift (ppm)
77.297
76.979
76.662
71.392
71.085
52.575
45.258
41.204
40.070
39.322
34.975
30.325
23.064
16.563

040610

exp5 Dept

SAMPLE

date Nov 29 2007

solvent cdc13

sample

ACQUISITION

sw 36764.7

at 1.000

np 73530

bs 64

ss -2

dl 1.000

nt 512

ct 512

TRANSMITTER

tn C13

tof 2339.1

tpwr 55

pw 7.600

DECOUPLER

dn H1

dof 0

dpwr 42

din nny

dmn vs

dmf CCW

pp1v1 hzmm

pp 15.500

DEPT

140.0

SPECIAL

1.5

temp

25.0

gain

40

spin

not used

PROCESSING

1.00

fn

65536

SPECTRUM

21197.4

wp

512

sp

128.2

rfp

-52.8

ai

REFERENCE

2546.9

rfp

0

71.649
71.337

52.827

45.522

41.467

40.329

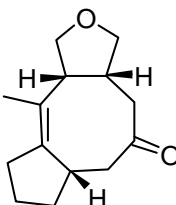
39.578

35.233

30.576

23.322

16.813



19a

130 120 110 100 90 80 70 60 50 40 30 20 10 ppm

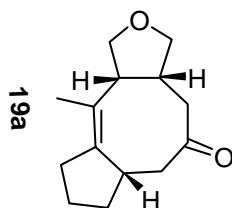
840610

File: GC05Y

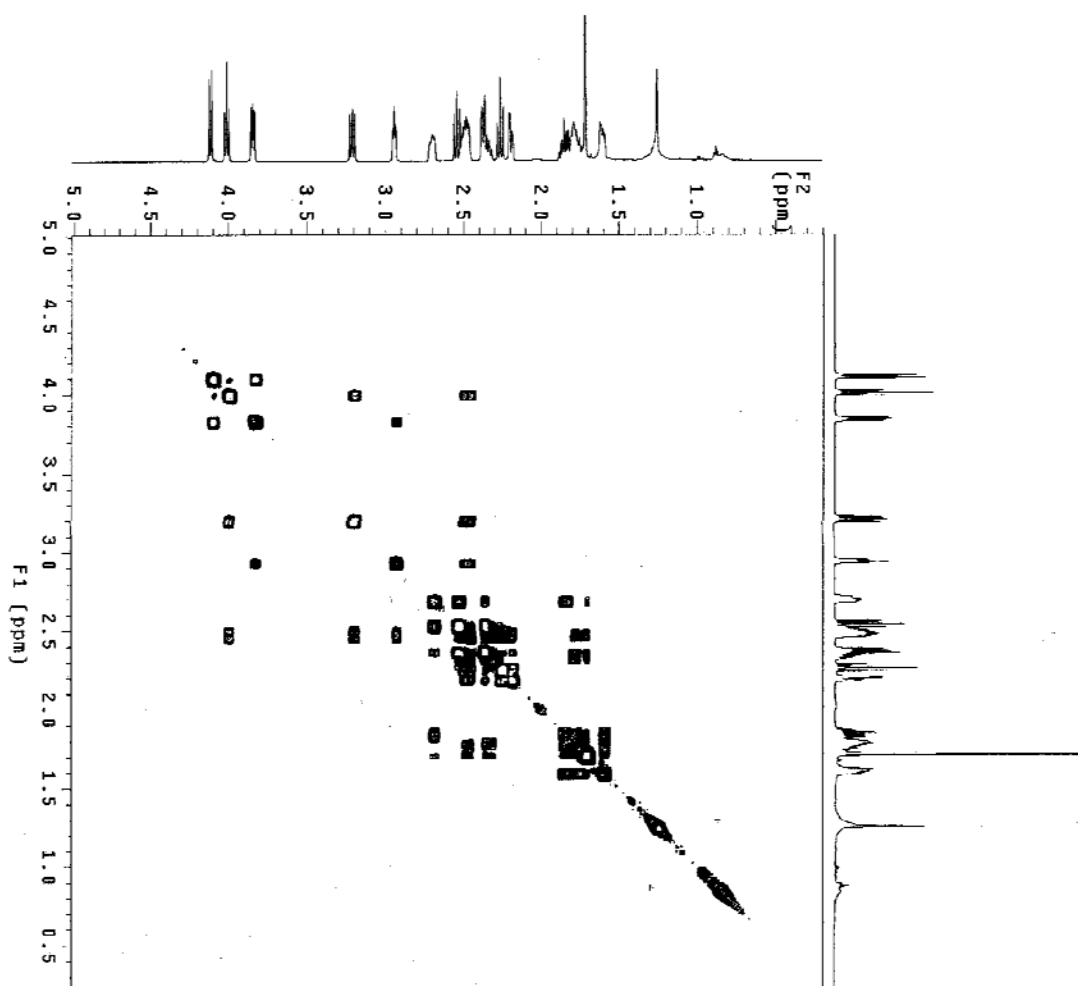
Pulse Sequence: gCOSY

Solvent: cdcl3
Temp: 25.0 C / 298.1 K
Operator: JY
VNMRS-600 ¹HCDNMR

Relax. delay 1.301 sec
Acq. time 0.177 sec
Width 2633.5 Hz
2D Width 2633.5 Hz
8 repetitions
322 increments
F2: 599.7744900 MHz
DATA PROCESSING
Sf. sine bell 0.088 sec
F1 DATA PROCESSING
Sf. sine bell 0.022 sec
F1 size 1024 x 1024
Total time 26 min, 12 sec



19a



040610

File: 040610-GHSQC-2

Pulse Sequence: ghsqc

Solvent: cdcl3

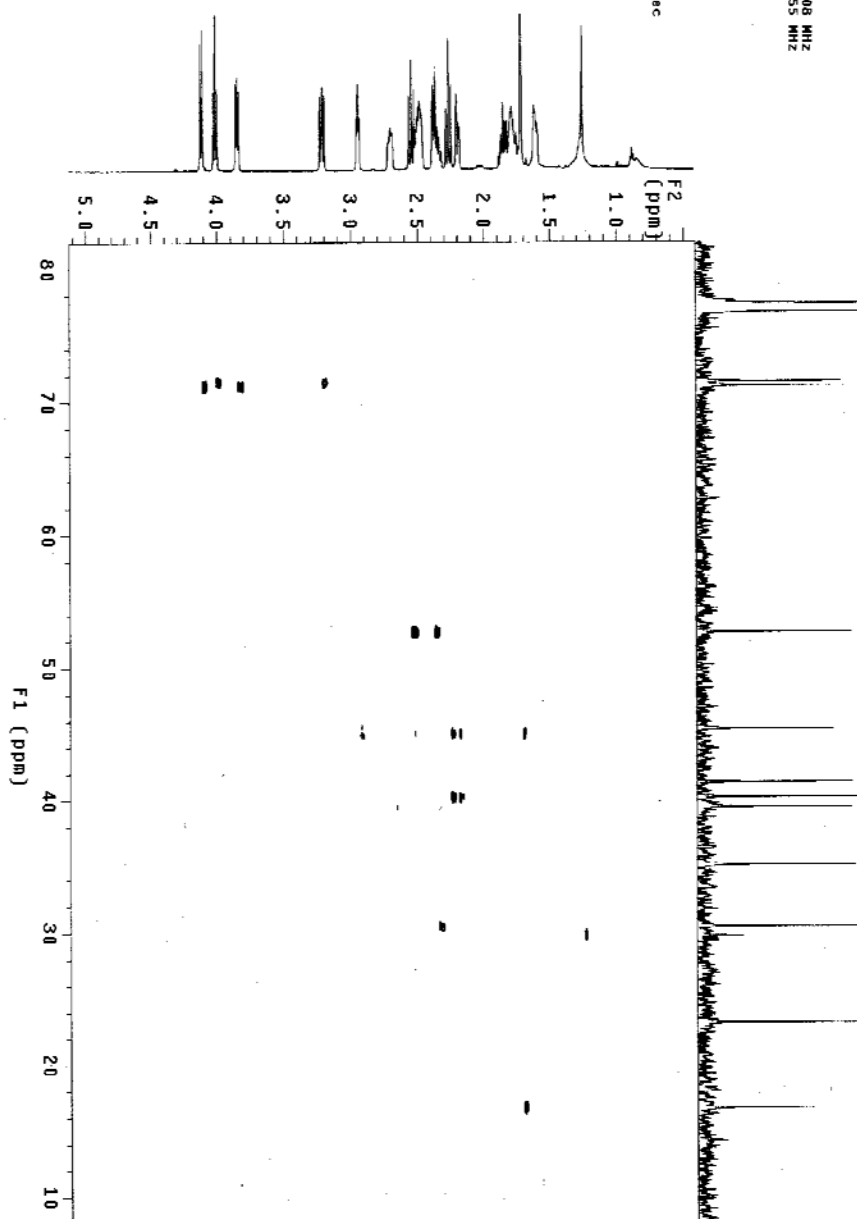
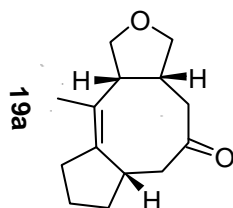
Temp: 25.0 C / 298.1 K

Observed: 13C

File: 040610-GHSQC-2

VMRS-600 "R1CDMR"

Relax. delay 1.301 sec
Acq. time 0.189 sec
Width 2828.1 Hz
2D Width 11150.7 Hz
80 repetitions
2.0000000000000000
Observed: 13C
Decouple: 13C, 150.820155 MHz
Power 36 dB
on during acquisition
off during delay
13C-13C decoupled
Data acquisition
Data processing
Sine bell 0.110 sec
F1 Data processing
Sine bell 0.111 sec
F1 size 2048 x 2048
Total time 27 min, 22 sec



040610

File: NOESY

Pulse Sequence: NOESY

Solvent: cdcl3

Temp: 25.0 C / 298.1 K

Operator: zly

VMRS-500 "RICHMR"

Relax. delay 1.000 sec

Mixing 0.300 sec

Acq. time 0.187 sec

Scan 271.2 Hz

20 Width 2741.2 Hz

16 Repetitions

2 x 128 increments

OBSERVE H1, 599.7744900 MHz

DATA PROCESSING

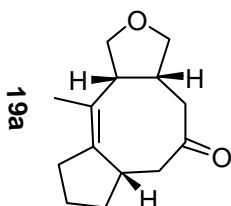
Gauss apodization 0.086 sec

F2 DATA PROCESSING

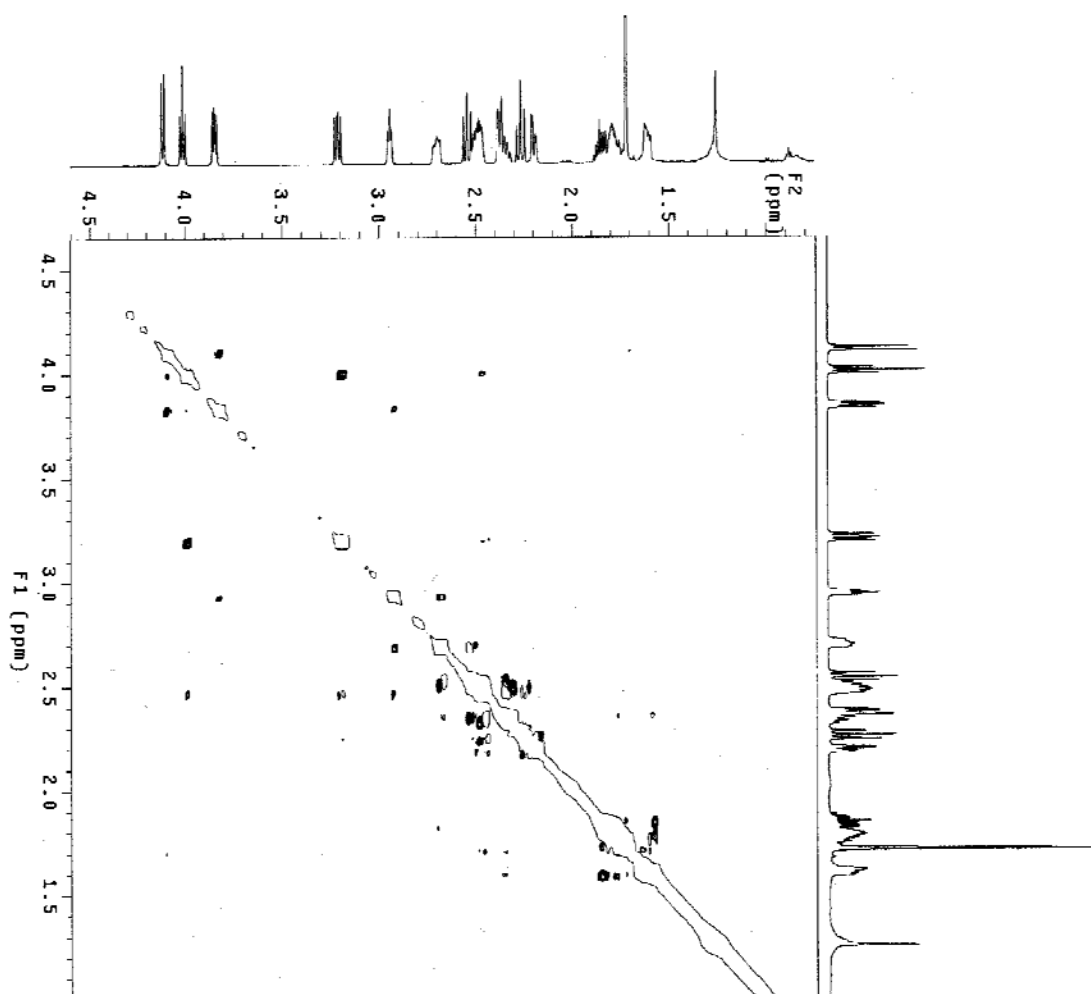
Gauss apodization 0.020 sec

FI size 1024 x 1024

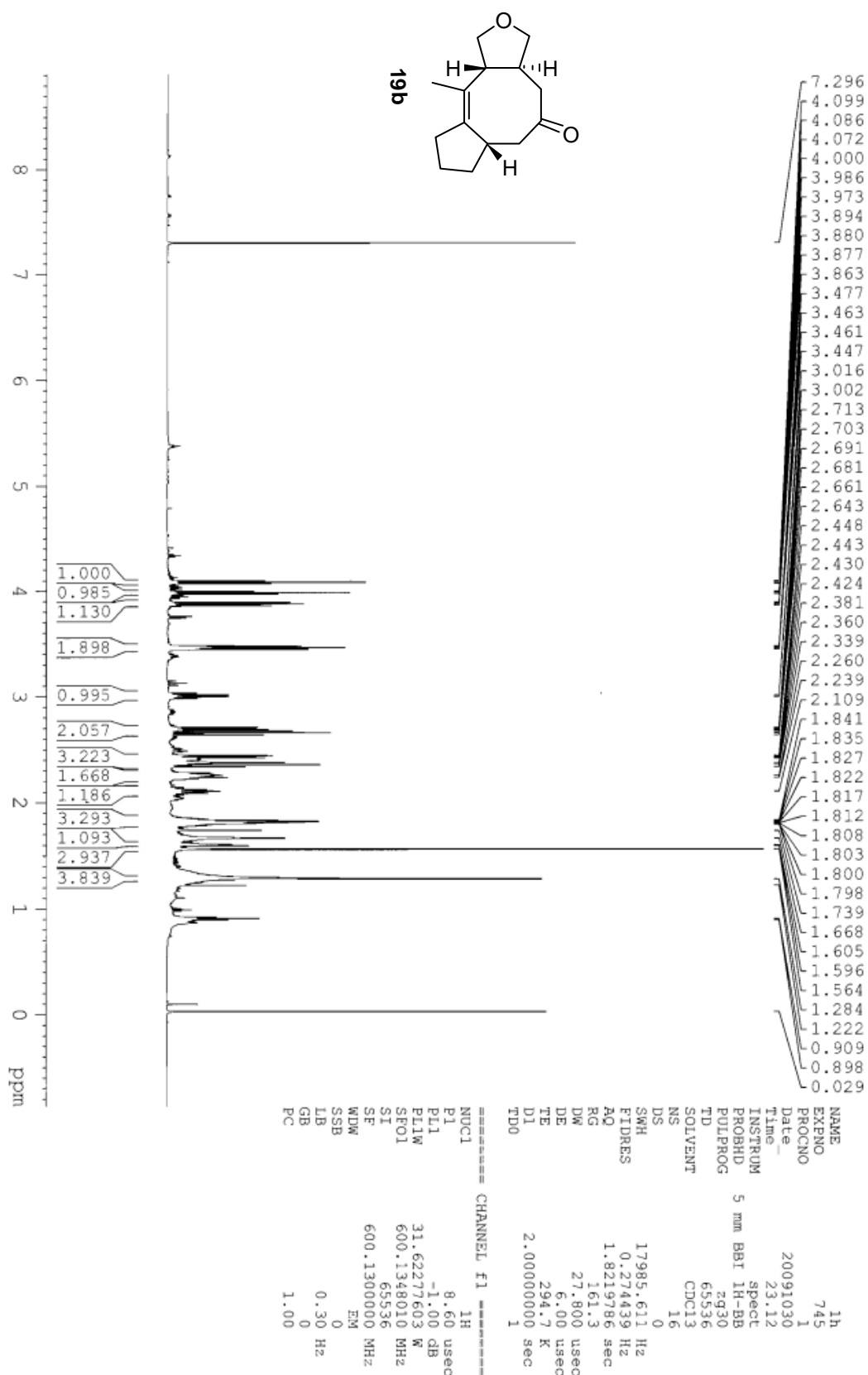
Total time 1 hr, 45 min, 4 sec

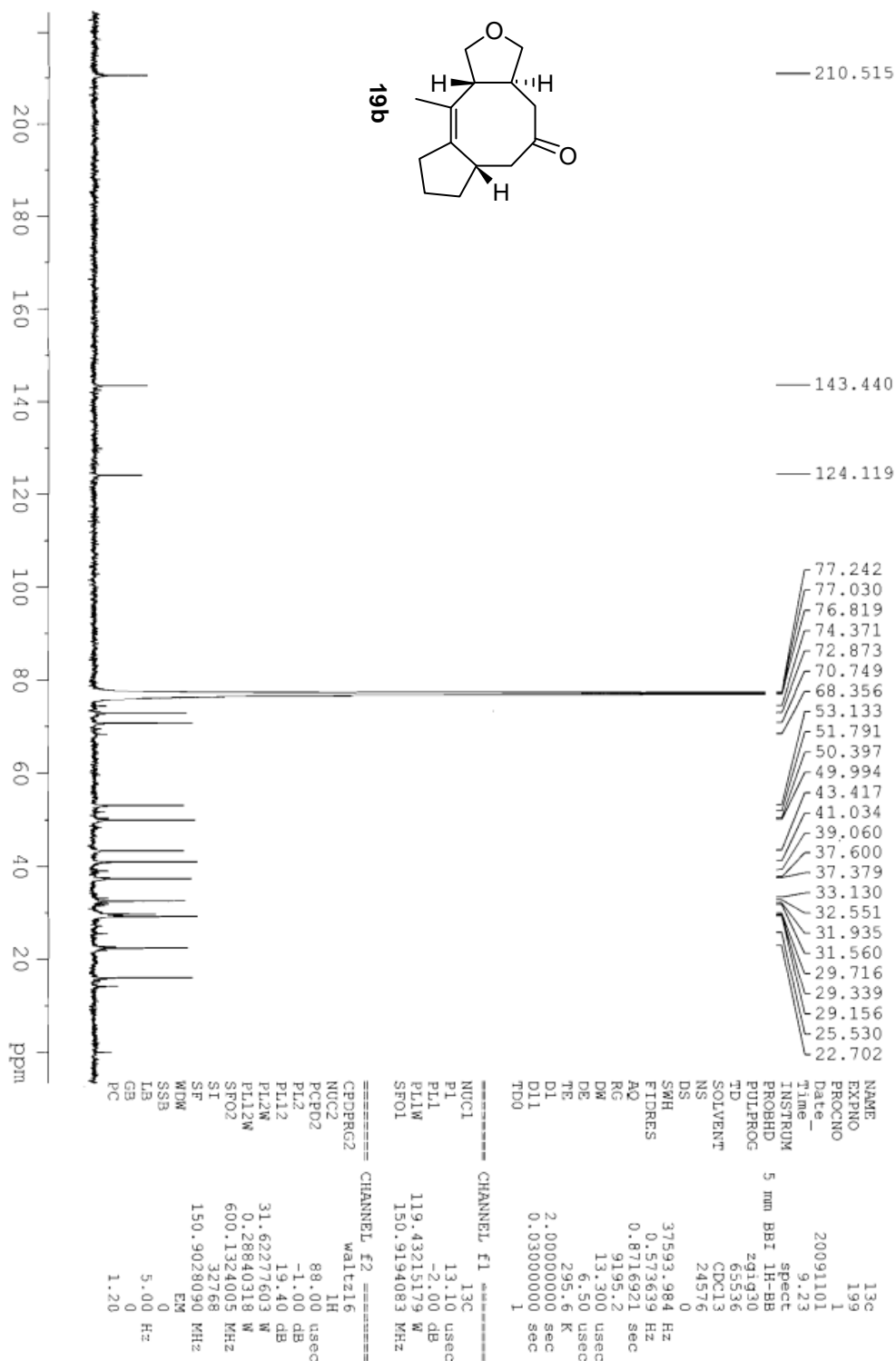


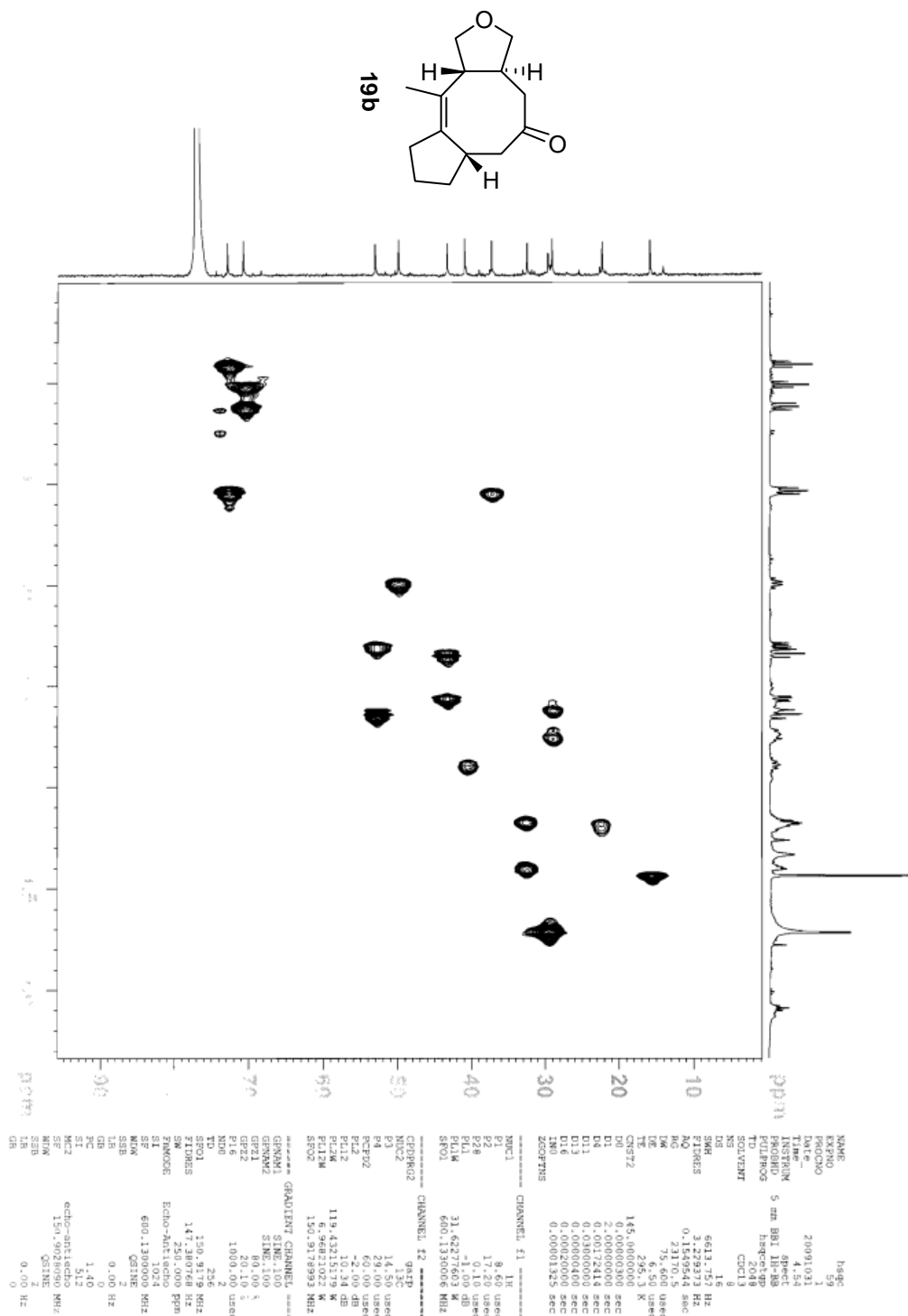
19a

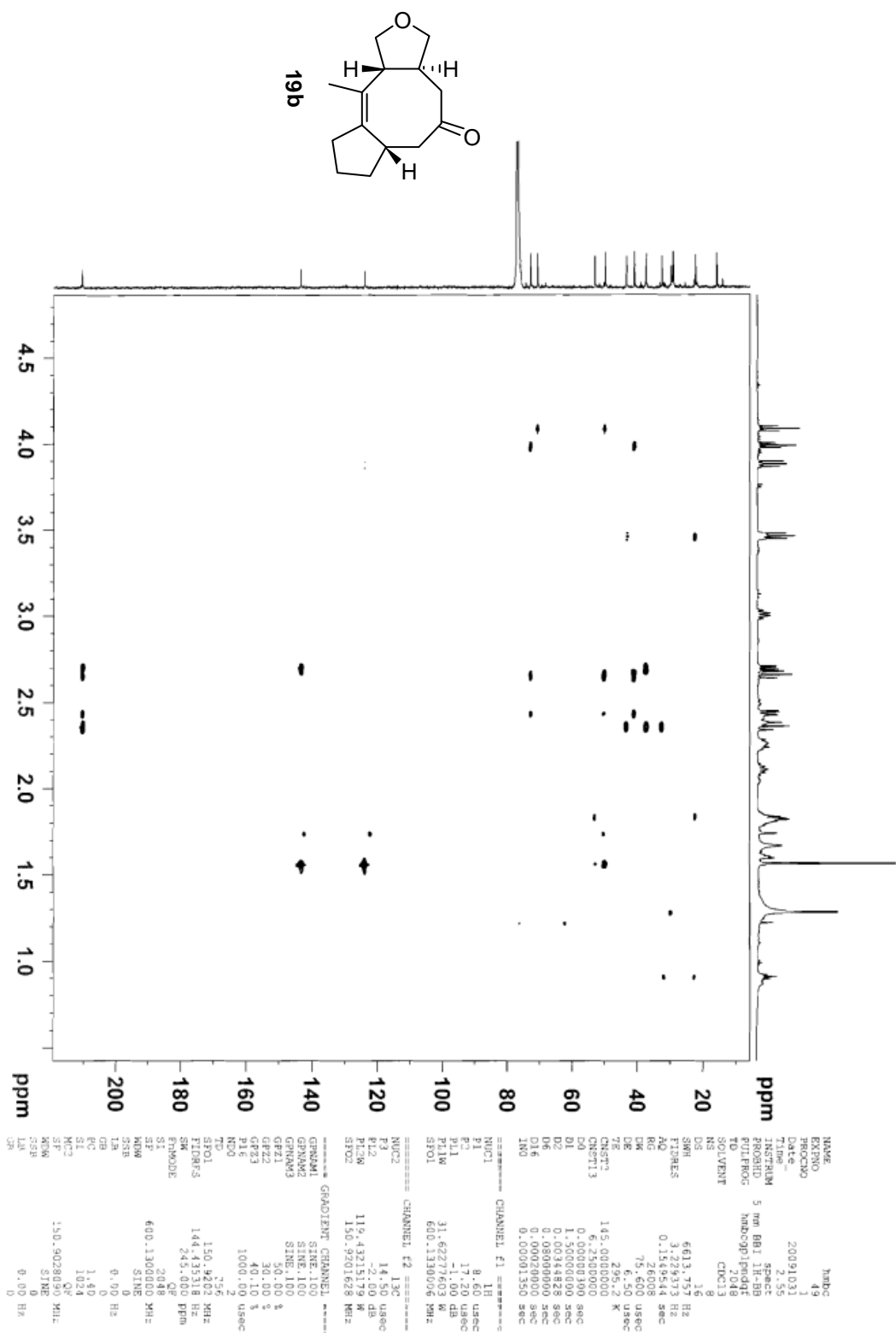


¹H NMR CDCl₃ ZKY-05-22C Yu Zhixiang 2009103

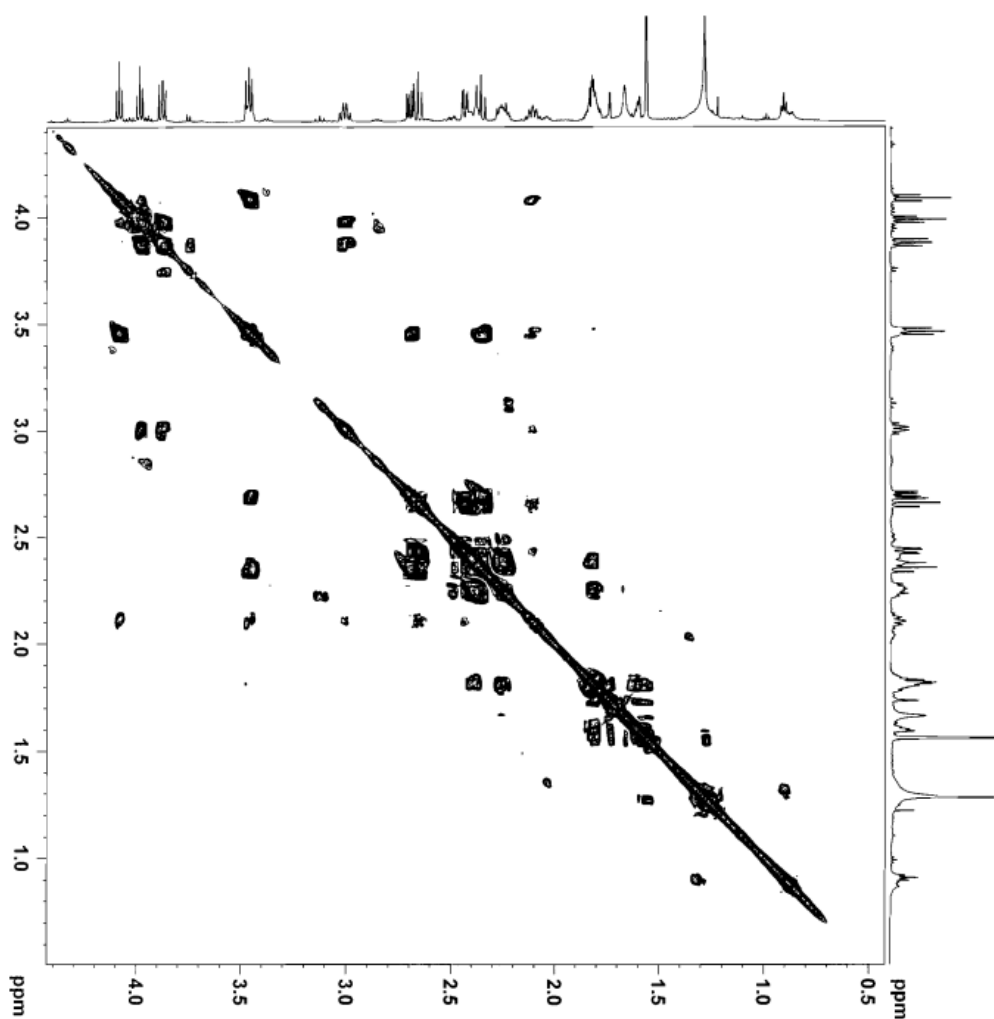
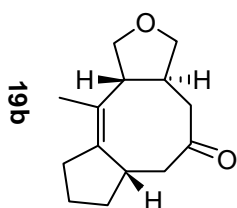








COSY CDC13 ZKY-05-22C Yu Zhixiang 20091030

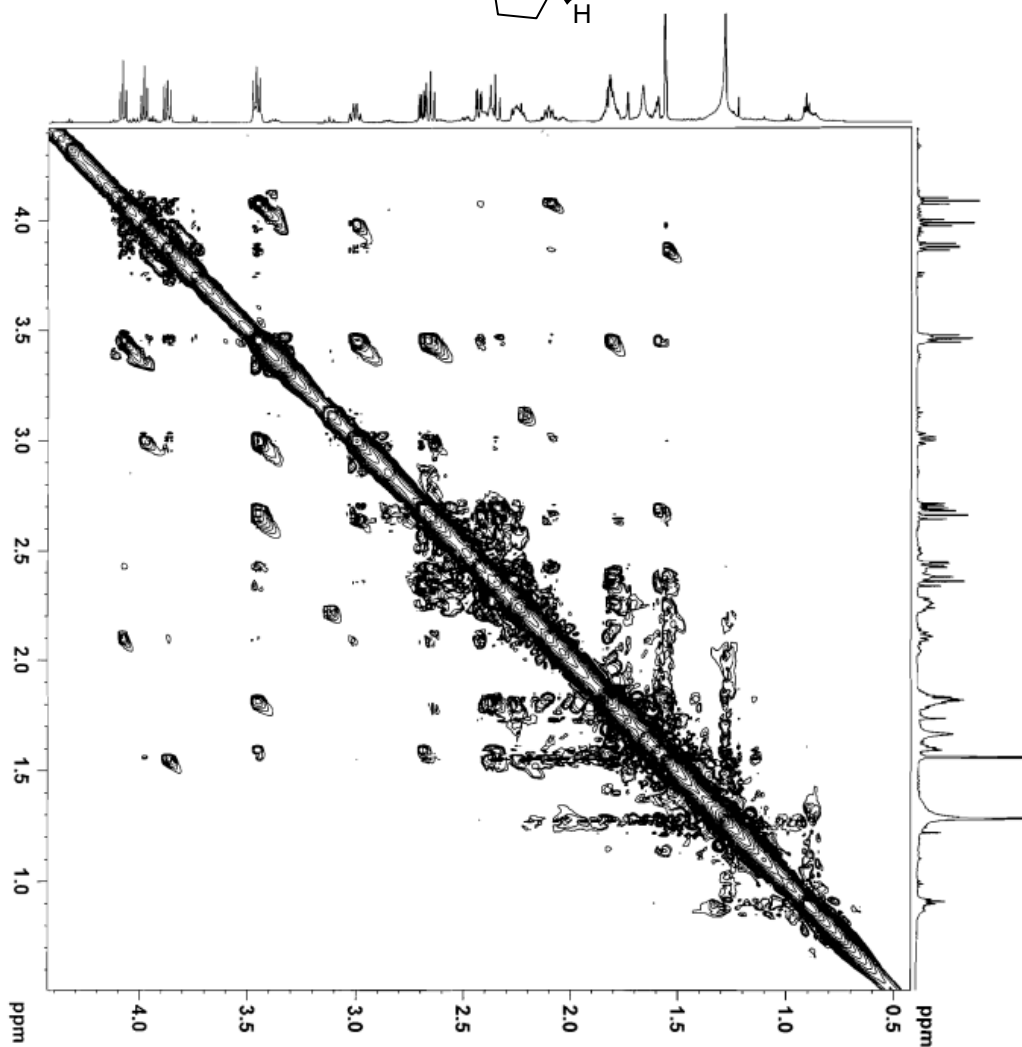
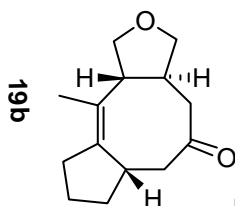


```

NAME          COSY
EXPNO         84
PROCNO        1
Date_         20091031
Time         3.56
INSTRUM       spect
PROBHD        5 mm BBI 1H-BB
PULPROG       cosyprgf
TD            2048
SOLVENT       CDC13
NS            8
DS           16
SRH           6613.757 Hz
FIDRES        3.228273 Hz
AQ            0.1349344 sec
RG            514.7
DE            75.600 usec
WE            6.50 usec
TE            295.0 K
D0            0.00000300 sec
D1            1.500000000 sec
D13           0.00000400 sec
D16           0.00002000 sec
INO           0.00015120 sec

===== CHANNEL f1 =====
NUC1          1H
P0            4.30 usec
PL            8.60 usec
PL1           -1.00 dB
PL1W          31.62271603 W
SFO1          600.1330006 MHz

===== GRADIENT CHANNEL =====
GENDMT        SINE,100
GRZ1          10.00 %
P16           1000.00 usec
ND0           1
TD            256
SFO1          600.133 MHz
FIDRES        25.835022 Hz
SW            11.021 ppm
FMODE         OF
SI            1024
SF            600.1300000 MHz
WDW           SINE
SSB           0
LB            0.00 Hz
GB            0
MC2           OF
SF            600.1300000 MHz
WDW           SINE
SSB           0
LB            0.00 Hz
GB            0
  
```



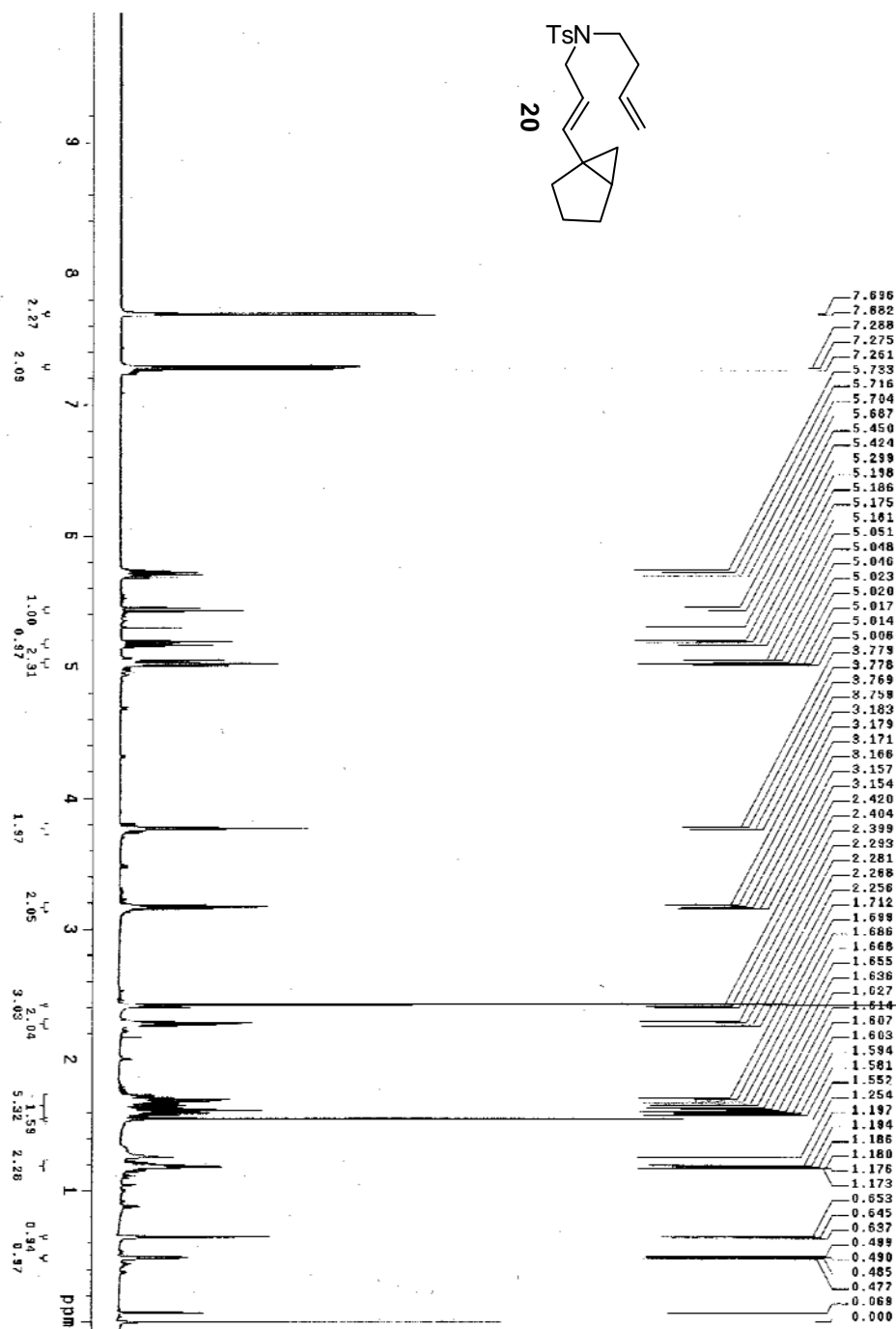
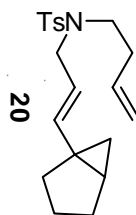
```

NAME          noesy
EXPNO         108
PROCNO        1
Date_         20091031
Time          10.17
INSTRUM       spect
PROBHD        5 mm BBI 1H-BB
PULPROG       noesypph
TD            2048
SOLVENT       CDCl3
NS            8
DS            16
SWH           5387.931 Hz
FIDRES       2.630826 Hz
AQ           0.1901972 sec
RG           161.3
DE           92.800 usec
TE           295.5 K
D0           0.00008185 sec
D1           2.00000000 sec
D8           0.50000000 sec
D16          0.00020000 sec
IN0          0.00018560 sec

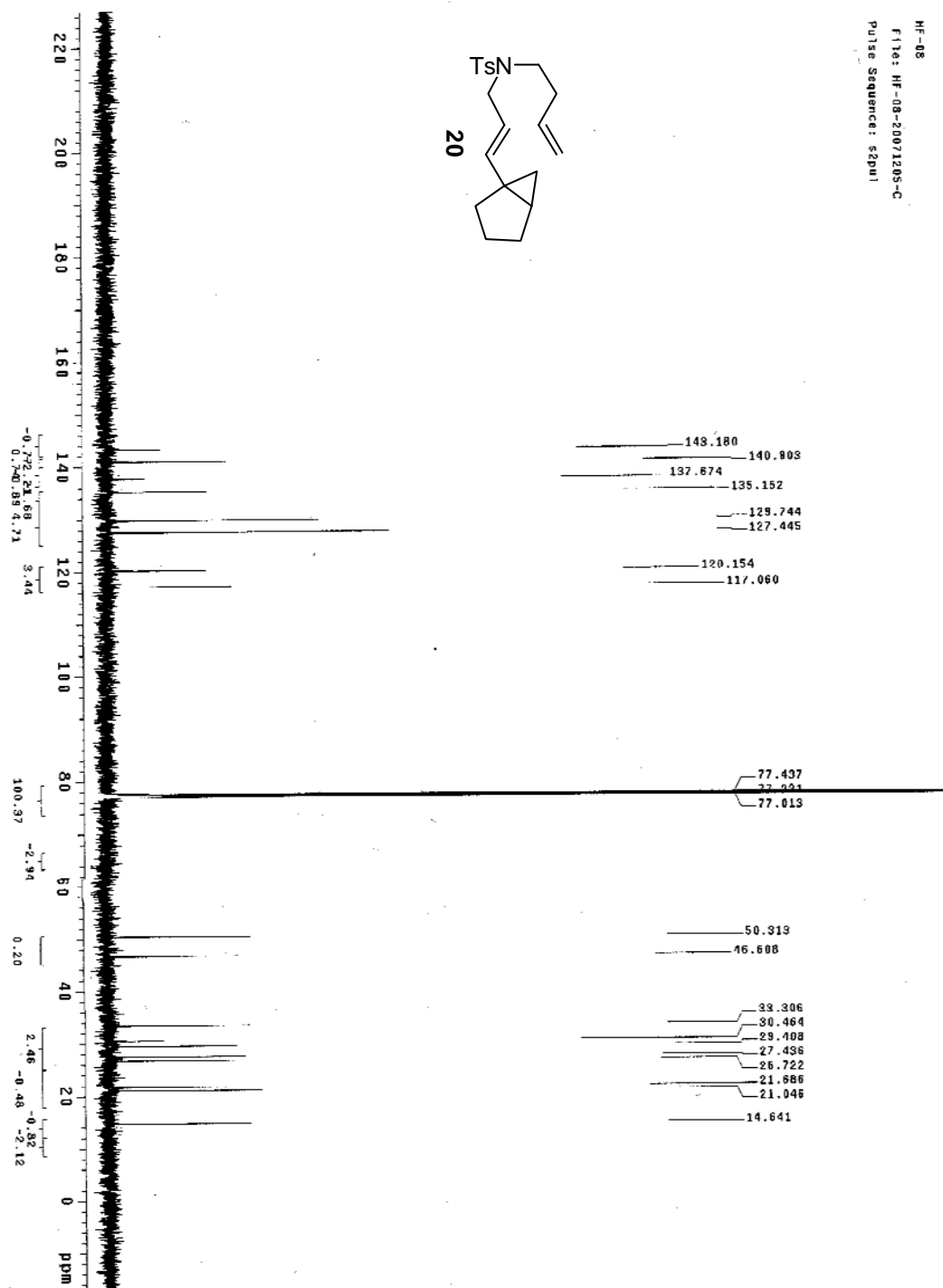
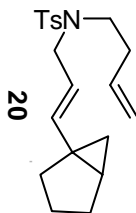
===== CHANNEL f1 =====
NUC1          1H
P1           8.60 usec
P2           17.20 usec
PL1          -1.00 dB
PL1W         31.62277603 W
SFO1         600.1324005 MHz

===== GRADIENT CHANNEL =====
GPNAM1       SINE.100
SINE.100    SINE.100
GPNAM2       SINE.100
SINE.100    SINE.100
GPZ1         40.00 %
GPZ2         -40.00 %
P16          1000.00 usec
ND0          1
TD           256
SFO1         600.1324 MHz
FIDRES       21.046597 Hz
SW           8.978 ppm
FMODE        States-TEPI
SI           1024
SF           600.1300000 MHz
WDW          QSINE
SSB          2
LB           0.00 Hz
GB           0
PC           1.40
SI           1024
MC2          States-TEPI
SF           600.1300000 MHz
WDW          QSINE
SSB          2
LB           0.00 Hz
GB           0
  
```

HF-08
 File: HF-08-20071205-H
 Pulse Sequence: szpu)

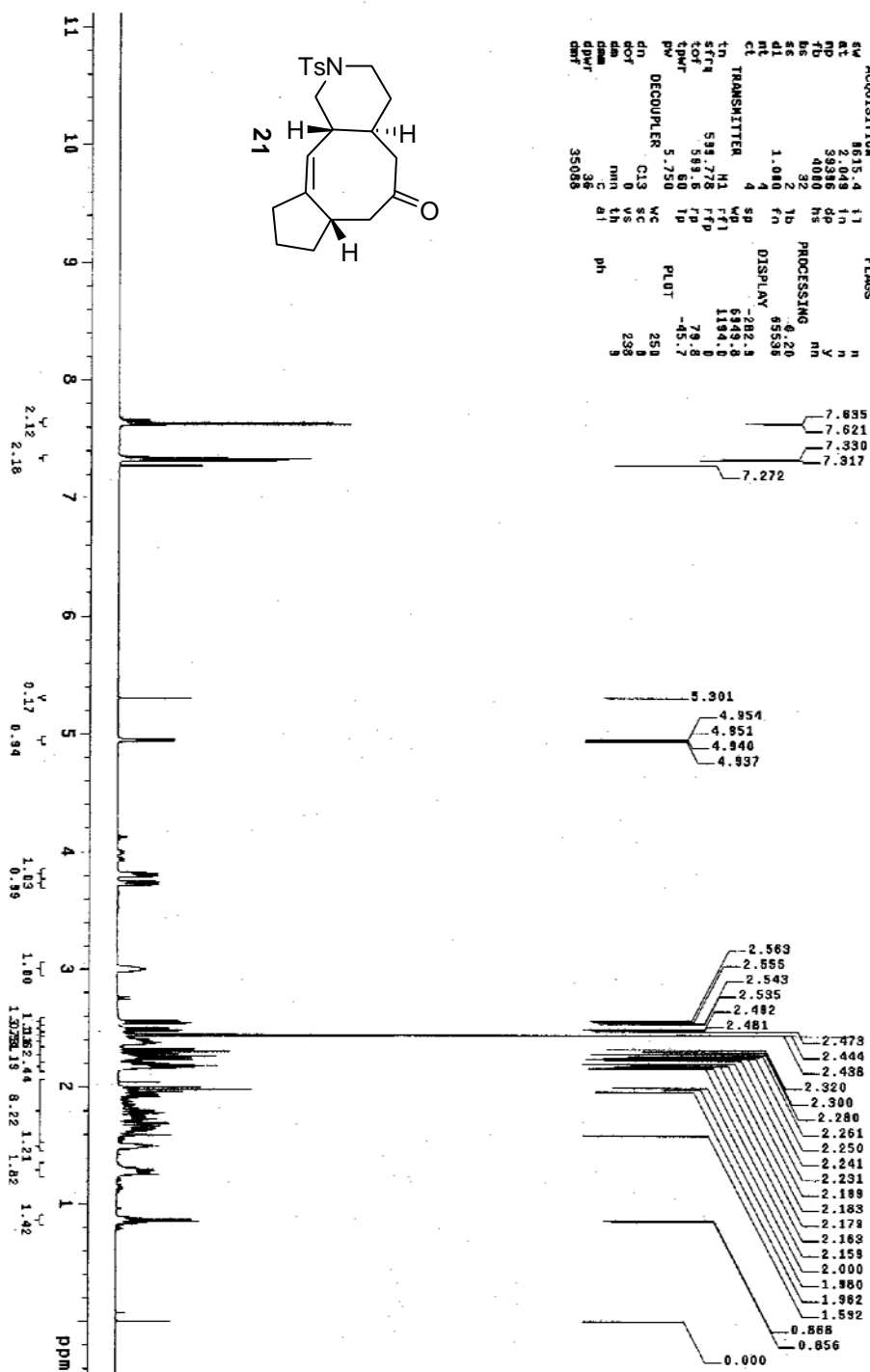
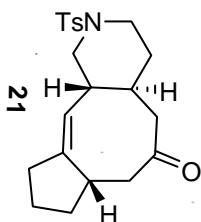


HF-08
 F116: HF-08-20071205-C
 Pulse Sequence: zgpg30



exp5 Proton

SAMPLE		2007		temp		SPECIAL	
date	Apr	5	2007	temp	not used		
solvent	CHCl3	CHCl3	gain	not used	20		
Y1	exp/orig	1	spin	not used	20		
Y2	exp/orig	1	spin	not used	20		
exp/eng	040827-H	f3	pw30	not used	11.500		
		id	at20		6.6000		
		id	at20				
ACQUISITION							
sw	8635.4	1	1	FLAGS			
at	2.0000	1	1				
rs	2.0000	1	1				
fb	2.0000	1	1				
bs	3000	32	2				
se	1.000	2	1b				
dl	1.000	4	4				
cl	1.000	4	4				
PROCESSING							
tn	TRANSMITTER	h1	f1f1				
sftr	533.778	7f	7f				
cor	583.5	7f	7f				
tptr	5.750	60	1p				
pv	DECODER	5.750	mc				
dn	C13	3	3				
eor	3	3	3				
dm	min	3	3				
dpr	36	36	36				
am	35008	35008	35008				
DISPLAY							
sp	282.8	3	3				
mc	582.8	3	3				
sftr	1194.0	1	1				
cor	79.8	8	8				
tptr	-45.7	7	7				
PLOT							
mc	251.8	3	3				
dn	3	3	3				
eor	3	3	3				
dm	3	3	3				
dpr	36	36	36				
am	35008	35008	35008				



040607

exps Carbon

SAMPLE SPECIAL
 date Apr 6 2007 temp not used
 solvent cdcl3 gain 40
 file /export/home/~ not used
 y1/vmrays/data/hu- hst 0.008
 angfreq/040607-C1- pw90 7.500
 id atla 10.000
 ACQUISITION
 sw 38764.7 11
 at 1.300 1n
 np 85824 dp
 fb 17000 hs
 bs 4
 n1 1.000 1b
 n2 2000 fn
 ct 36 not used
 TRANSMITTER
 tn C13 sp
 sfr 150.829 rft1
 tot 2359.1 rfp
 tprt 55 1p
 pw 9.000 1p
 DECOUPLER H1 wc
 dn 0 sc
 dof 0 vs
 dm yy
 dpr 42 th
 dnt at cdc ph
 PLOT 250
 7465 7

